

Supplementary Information

Convergent and Divergent Synthesis of Dihydroisoquinoline-1,4-diones Enabled by a Photocatalytic Skeleton-Editing [4+2] Strategy

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I. General Information

General remarks: ^1H and ^{13}C NMR spectra were recorded in CDCl_3 (unless otherwise noted) on a Bruker AVANCE 600 MHz or a Bruker AVANCE 400 MHz spectrometer. Chemical shifts in ^1H NMR spectra were reported in parts per million (ppm) on the δ scale from an internal standard of TMS (0.00 ppm). Data for ^1H NMR were reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, brs = broad), coupling constant in Hertz (Hz) and integration. Data for ^{13}C NMR spectra were reported in terms of chemical shift in ppm from the central peak of CDCl_3 (77.16 ppm). High-resolution electrospray ionization and electronic impact mass spectrometry were performed on a Thermo Scientific Q Exactive mass spectrometer (mass analyzer type: Orbitrap).

Materials and methods: Unless otherwise noted, all reactions and substrates preparation were conducted in flame-dried glassware under a nitrogen atmosphere using anhydrous solvent re-distilled according to Purification of Laboratory Chemicals (Fifth Edition). Commercially available reagents were used without further purification. Thin layer chromatography (TLC) was performed on Jiangyou TLC silica gel plates HSG F254 and visualized by UV light or staining with anisaldehyde or potassium permanganate. Flash column chromatography was performed over silica gel (300-400 mesh). Photochemical reactions were carried out under irradiation from 30 W blue LED (composed of 30 LED units each with 1.0 W).

II. Reaction Condition Optimization

Table S1. Investigation of the amount of 2,4,6-collidine for two-component reaction.^a

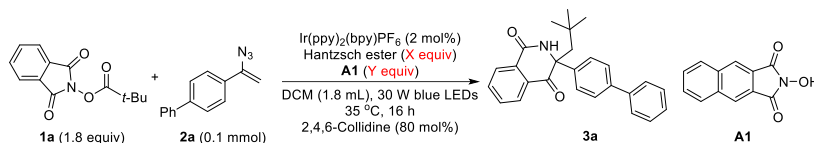
Reaction scheme: 1a (1.8 equiv) + 2a (0.1 mmol) $\xrightarrow[\text{2,4,6-Collidine (X mol\%)}]{\text{Ir(ppy)}_2\text{(bpy)PF}_6 \text{ (2 mol\%)}, \text{Hantzsch ester (4.2 equiv)}, \text{A1 (2.7 equiv)}, \text{DCM (1.8 mL)}, 30 \text{ W blue LEDs}, 35 \text{ }^\circ\text{C}, 16 \text{ h}}$ 3a

Structure A1: O=C1C(=O)N1c2ccc3ccccc3cc2

| Entry | 2,4,6-Collidine (X mol%) | Conv. of 2a (%) | Yield of 3a (%) |
|-------|--------------------------|------------------------|------------------------|
| 1 | 0 | >95 | 43 |
| 2 | 10 | >95 | 42 |
| 3 | 20 | >95 | 50 |
| 4 | 40 | >95 | 57 |
| 5 | 80 | >95 | 62 |
| 6 | 100 | >95 | 52 |
| 7 | 150 | >95 | 37 |
| 8 | 200 | >95 | 33 |

^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (0.27 mmol), Ir(ppy)₂(bpy)PF₆ (0.002 mmol), Hantzsch ester (0.42 mmol) and 2,4,6-collidine (X mol%) in DCM (1.8 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 16 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

Table S2. Investigation of the amount of **A1** and Hantzsch ester for two-component reaction.^a



| Entry | (X equiv) | (Y equiv) | Conv. of 2a (%) | Yield of 3a (%) |
|-------|-----------|-----------|------------------------|------------------------|
| 1 | 4.2 | 2.2 | >95 | 55 |
| 2 | 4.2 | 2.5 | >95 | 67 |
| 3 | 4.2 | 3.0 | >95 | 60 |
| 4 | 4.2 | 3.5 | >95 | 47 |
| 5 | 3.5 | 2.5 | >95 | 58 |
| 6 | 3.8 | 2.5 | >95 | 62 |
| 7 | 4.0 | 2.5 | >95 | 68 (63) |
| 8 | 4.5 | 2.5 | >95 | 60 |

^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (Y equiv), Ir(ppy)₂(bpy)PF₆ (0.002 mmol), Hantzsch ester (X equiv) and 2,4,6-collidine (0.08 mmol) in DCM (1.8 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 16 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard. Isolated yield after flash chromatography is shown in the parentheses.

Table S3. Investigation of the PC for two-component reaction.^a

$\text{1a (1.8 equiv)} + \text{2a (0.1 mmol)} \xrightarrow[\text{DCM (1.8 mL), 30 W blue LEDs, 35 } ^\circ\text{C, 16 h}]{\text{PC (2 mol\%), Hantzsch ester (4.0 equiv), A1 (2.5 equiv)}} \text{3a}$

| Entry | PC | Conv. of 2a (%) | Yield of 3a (%) |
|-------|---|------------------------|------------------------|
| 1 | Ir(ppy) ₂ (bpy)PF ₆ | >95 | 68 (63) |
| 2 | 4CzIPN | >95 | 56 |
| 3 | 4CzIPN-Cl | >95 | 52 |
| 4 | Ir(ppy) ₃ | >95 | trace |
| 5 | Ru(bpy) ₃ (PF ₆) ₂ | >95 | 34 |
| 6 | Mes-Acr-3,6- <i>t</i> Bu ₂ -Ph ⁺ BF ₄ ⁻ | >95 | trace |

Ir(ppy)₂(bpy)PF₆

Ir(ppy)₃

Ru(bpy)₃(PF₆)₂

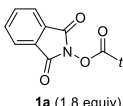
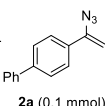
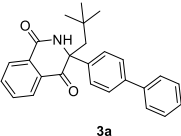
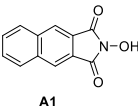
Mes-Acr-3,6-*t*Bu₂-Ph⁺ BF₄⁻

4CzIPN

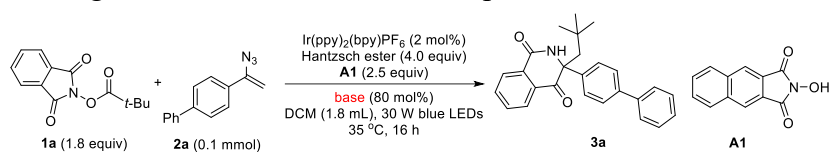
4CzIPN-Cl

^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (0.25 mmol), PC (0.002 mmol), Hantzsch ester (0.40 mmol) and 2,4,6-collidine (0.08 mmol) in DCM (1.8 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 16 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard. Isolated yield after flash chromatography is shown in the parentheses.

Table S4. Investigation of the solvent for two-component reaction.^a

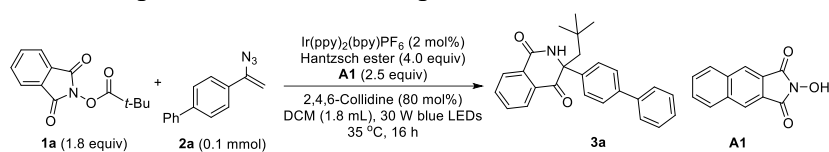
| <div><div><div><div><p>1a (1.8 equiv)</p></div><div><p>2a (0.1 mmol)</p></div></div><div><div><p>3a</p></div><div><p>A1</p></div></div></div><div><div><p>Ir(ppy)₂(bpy)PF₆ (2 mol%) Hantzsch ester (4.0 equiv) A1 (2.5 equiv) 2,4,6-Collidine (80 mol%) solvent (1.8 mL), 30 W blue LEDs 35 °C, 16 h</p></div></div></div> | | | |
|---|-----------|------------------------|------------------------|
| Entry | Solvent | Conv. of 2a (%) | Yield of 3a (%) |
| 1 | DCE | >95 | 41 |
| 2 | 1,1,2-TCE | >95 | 35 |
| 3 | PhCl | >95 | 44 |
| 4 | MeCN | >95 | 22 |
| 5 | THF | >95 | ND |

^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (0.25 mmol), Ir(ppy)₂(bpy)PF₆ (0.002 mmol), Hantzsch ester (0.40 mmol) and 2,4,6-collidine (0.08 mmol) in solvent (1.8 mL) was irradiated with 30 W blue LEDs at room temperature under N₂ for 16 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard. ND = not detected.

Table S5. Investigation of the base for two-component reaction.^a

| Entry | Base | Conv. of 2a (%) | Yield of 3a (%) |
|-------|---------------------------------|------------------------|------------------------|
| 1 | 2,6-lutidine | >95 | 48 |
| 2 | DABCO | >95 | 56 |
| 3 | DMAP | >95 | 59 |
| 4 | K ₂ CO ₃ | >95 | 47 |
| 5 | Cs ₂ CO ₃ | >95 | 12 |
| 6 | K ₂ HPO ₄ | >95 | 58 |

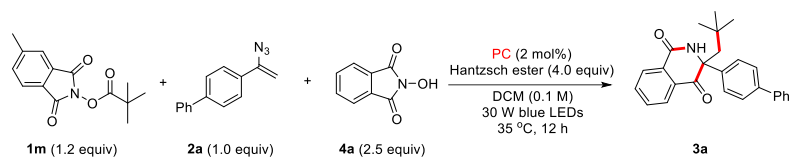
^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (0.25 mmol), Ir(ppy)₂(bpy)PF₆ (0.002 mmol), Hantzsch ester (0.40 mmol) and base (0.08 mmol) in DCM (1.8 mL) was irradiated with 30 W blue LEDs at room temperature under N₂ for 16 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

Table S6. Control experiment for two-component reaction.^a

| Entry | variation from standard conditions | Conv. of 2a (%) | Yield of 3a (%) |
|-------|---|------------------------|------------------------|
| 1 | w/o $\text{Ir(ppy)}_2(\text{bpy})\text{PF}_6$ | >95 | ND |
| 2 | w/o Light | <5 | ND |
| 3 | w/o Hantzsch | >95 | ND |
| 4 | w/o A1 | >95 | 11 |
| 5 | Air | >95 | 31 |

^a Reaction conditions: A mixture of **1a** (0.18 mmol), **2a** (0.10 mmol), **A1** (0.25 mmol), $\text{Ir(ppy)}_2(\text{bpy})\text{PF}_6$ (0.002 mmol), Hantzsch ester (0.40 mmol) and 2,4,6-collidine (0.08 mmol) in DCM (1.8 mL) was irradiated with 30 W blue LEDs at room temperature under N_2 for 16 h. Yield was determined by ^1H NMR of the crude mixture using mesitylene as internal standard. ND = not detected.

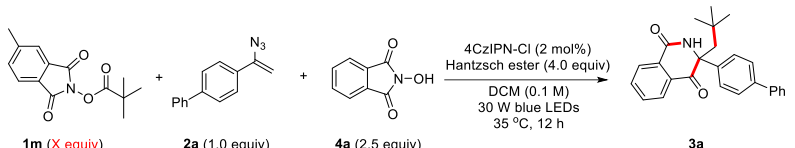
Table S7. Investigation of photocatalyst for three-component reaction.^a



| Entry | PC | Conv. of 2a (%) | Yield of 3a (%) |
|----------|---|------------------------|------------------------|
| 1 | 4CzIPN-Cl | >95 | 58 |
| 2 | Ir(ppy) ₂ (bpy)PF ₆ | >95 | 45 |
| 3 | 4CzIPN | >95 | 50 |

^a Reaction conditions: A mixture of **1m** (0.12 mmol), **2a** (0.10 mmol), **4a** (0.25 mmol), PC (0.002 mmol), Hantzsch ester (0.40 mmol) in DCM (1.0 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

Table S8. Investigation of amount of **1m** for three-component reaction.^a

|  | | | |
|--|---------------------|------------------------|------------------------|
| Entry | 1m (X equiv) | Conv. of 2a (%) | Yield of 3a (%) |
| 1 | 1.2 | >95 | 58 |
| 2 | 1.5 | >95 | 61 |
| 3 | 1.8 | >95 | 64 |
| 4 | 2.0 | >95 | 65 |
| 5 | 2.2 | >95 | 65 |
| 6 | 2.4 | >95 | 65 |
| 7 | 2.6 | >95 | 68 |
| 8 | 2.8 | >95 | 69 |
| 9 | 3.0 | >95 | 72 |
| 10 | 3.2 | >95 | 69 |
| 11 | 3.5 | >95 | 61 |

^a Reaction conditions: A mixture of **1m** (X equiv), **2a** (0.10 mmol), **4a** (0.25 mmol), 4CzIPN-Cl (0.002 mmol), Hantzsch ester (0.40 mmol) in DCM (1.0 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

Table S9. Investigation of amount of Hantzsch ester and concentration for three-component reaction.^a

$\text{1m (3.0 equiv)} + \text{2a (1.0 equiv)} + \text{4a (2.5 equiv)} \xrightarrow[\text{35 } ^\circ\text{C, 12 h}]{\text{4CzIPN-Cl (2 mol\%), Hantzsch ester (X equiv), DCM (Y mL), 30 W blue LEDs}} \text{3a}$

| Entry | HE (X equiv) | DCM (Y mL) | Conv. of 2a (%) | Yield of 3a (%) |
|-------|--------------|------------|------------------------|------------------------|
| 1 | 3.5 | 1.0 | >95 | 40 |
| 2 | 3.8 | 1.0 | >95 | 44 |
| 3 | 4.2 | 1.0 | >95 | 62 |
| 4 | 4.5 | 1.0 | >95 | 53 |
| 5 | 4.0 | 0.8 | >95 | 66 |
| 6 | 4.0 | 1.2 | >95 | 66 |
| 7 | 4.0 | 1.5 | >95 | 56 |

^a Reaction conditions: A mixture of **1m** (0.30 mmol), **2a** (0.10 mmol), **4a** (0.25 mmol), 4CzIPN-Cl (0.002 mmol), Hantzsch ester (X equiv) in DCM (Y mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

Table S10. Investigation of base for three-component reaction.^a

1m (3.0 equiv) + **2a** (1.0 equiv) + **4a** (2.5 equiv) $\xrightarrow[\text{35 } ^\circ\text{C, 12 h}]{\text{4CzIPN-Cl (2 mol\%), Hantzsch ester (4.0 equiv), base (X equiv), DCM (0.1 M)}}$ **3a**

| Entry | base (X equiv) | Conv. of 2a (%) | Yield of 3a (%) |
|-------|---|------------------------|------------------------|
| 1 | DMAP (1.0 equiv) | >95 | 38 |
| 2 | DABCO (1.0 equiv) | >95 | 45 |
| 3 | 2,4,6-collidine (1.0 equiv) | >95 | 61 |
| 4 | 2,6-lutidine (1.0 equiv) | >95 | 49 |
| 5 | Li ₂ CO ₃ (1.0 equiv) | >95 | 61 |
| 6 | K ₂ HPO ₄ (1.0 equiv) | >95 | 31 |
| 7 | KHCO ₃ (1.0 equiv) | >95 | 44 |
| 8 | NaOAc (1.0 equiv) | >95 | 24 |
| 9 | 2,4,6-collidine (0.8 equiv) | >95 | 60 |
| 10 | 2,4,6-collidine (0.6 equiv) | >95 | 61 |
| 11 | 2,4,6-collidine (0.4 equiv) | >95 | 61 |
| 12 | 2,4,6-collidine (0.2 equiv) | >95 | 62 |

^a Reaction conditions: A mixture of **1m** (0.30 mmol), **2a** (0.10 mmol), **4a** (0.25 mmol), 4CzIPN-Cl (0.002 mmol), Hantzsch ester (0.4 mmol) and base (X equiv) in DCM (1.0 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard.

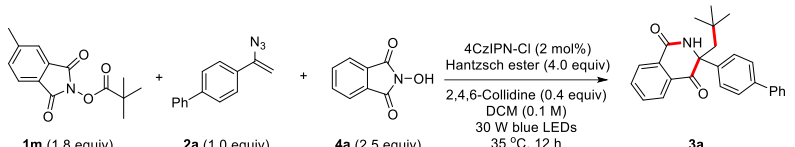
Table S11. Investigation of amount of **1m** for three-component reaction.^a

1m (X equiv) + **2a** (1.0 equiv) + **4a** (2.5 equiv) $\xrightarrow[\text{30 W blue LEDs, 35 } ^\circ\text{C, 12 h}]{\text{4CzIPN-Cl (2 mol\%), Hantzsch ester (4.0 equiv), 2,4,6-collidine (0.4 equiv), DCM (0.1 M)}}$ **3a**

| Entry | 1m (X equiv) | Conv. of 2a (%) | Yield of 3a (%) |
|-------|---------------------|------------------------|------------------------|
| 1 | 3.2 | >95 | 51 |
| 2 | 2.8 | >95 | 59 |
| 3 | 2.6 | >95 | 59 |
| 4 | 2.4 | >95 | 75 |
| 5 | 2.2 | >95 | 76 |
| 6 | 2.0 | >95 | 78 |
| 7 | 1.8 | >95 | 80 (78) |
| 8 | 1.6 | >95 | 72 |
| 9 | 1.4 | >95 | 66 |

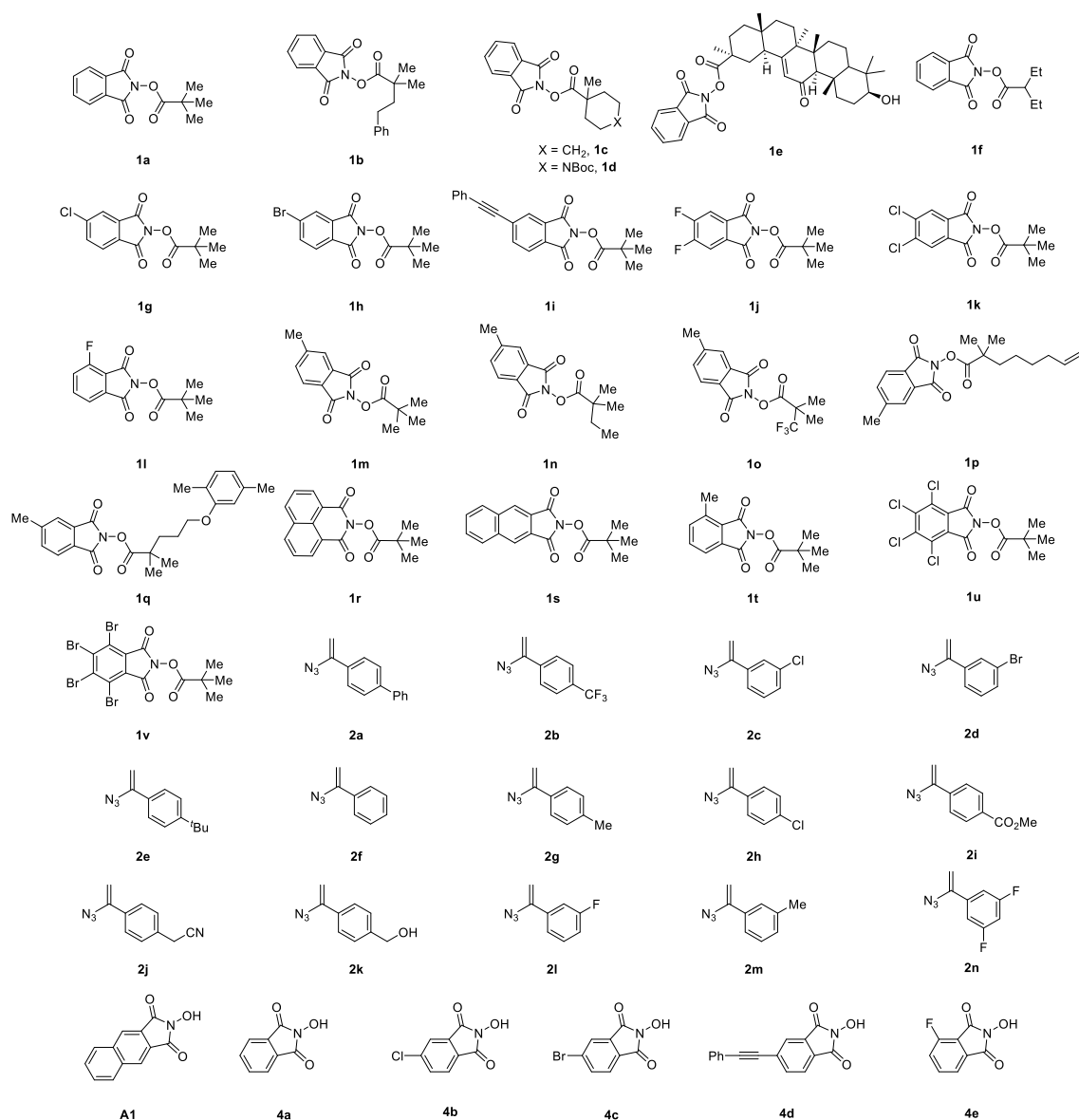
^a Reaction conditions: A mixture of **1m** (X equiv), **2a** (0.10 mmol), **4a** (0.25 mmol), 4CzIPN-Cl (0.002 mmol), Hantzsch ester (0.4 mmol) and 2,4,6-collidine (0.04 mmol) in DCM (1.0 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard. Isolated yield after flash chromatography is shown in the parentheses.

Table S12. Control experiment for three-component reaction.^a

|  1m (1.8 equiv) 2a (1.0 equiv) 4a (2.5 equiv) 3a | | | |
|---|------------------------------------|------------------------|------------------------|
| Entry | variation from standard conditions | Conv. of 2a (%) | Yield of 3a (%) |
| 1 | w/o PC | 55 | ND |
| 2 | w/o Hantzsch ester | >95 | ND |
| 3 | w/o light | <5 | ND |
| 4 | w/o 2,4,6-collidine | >95 | 56 |
| 5 | w/o 4a | >95 | ND |

^a Reaction conditions: a mixture of **1m** (0.18 mmol), **2a** (0.10 mmol), **4a** (0.25 mmol), 4CzIPN-Cl (0.002 mmol), Hantzsch ester (0.4 mmol) and 2,4,6-collidine (0.04 mmol) in DCM (1.0 mL) was irradiated with 30 W blue LEDs at 35 °C under N₂ for 12 h. Yield was determined by ¹H NMR of the crude mixture using mesitylene as internal standard. Isolated yield after flash chromatography is shown in the parentheses. ND = not detected. ND = not detected.

III. Substrate Source



Synthesis and characterization of substrates

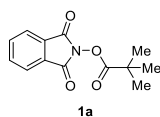
4a, **4f** were purchased and used directly without further purification.

The following substrates are known compounds:

1a¹, **1b**², **1c**³, **1d**⁴, **1e**⁵, **1f**¹, **1t**⁶, **2a**¹, **2b**¹, **2c**¹, **2d**¹, **2e**¹, **2f**¹, **2g**¹, **2h**⁷, **2i**⁷, **2j**⁸, **2k**¹, **2l**¹, **2m**⁷, **2n**⁹, **A1**¹⁰, **4b**¹¹, **4c**¹⁰, **4e**¹².

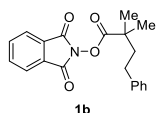
These compounds were prepared according to literature: **1g**¹, **1h**¹, **1i**¹, **1j**¹, **1k**¹, **1l**¹, **1m**¹, **1n**¹, **1o**¹, **1p**¹, **1q**¹, **1r**¹, **1s**¹, **1u**¹, **1v**¹, **4d**¹⁰.

1,3-Dioxoisindolin-2-yl pivalate (**1a**)



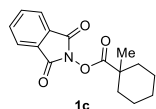
^1H NMR (400 MHz, Chloroform-*d*) δ 7.87 (dd, J = 5.4, 3.1 Hz, 2H), 7.78 (dd, J = 5.4, 3.1 Hz, 2H), 1.43 (s, 9H).

1,3-Dioxoisindolin-2-yl 2,2-dimethyl-4-phenylbutanoate (1b)



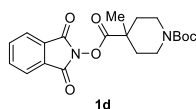
^1H NMR (600 MHz, Chloroform-*d*) δ 7.92 – 7.87 (m, 2H), 7.82 – 7.76 (m, 2H), 7.33 – 7.26 (m, 4H), 7.22 – 7.17 (m, 1H), 2.80 – 2.74 (m, 2H), 2.06 – 2.00 (m, 2H), 1.47 (s, 6H).

1,3-Dioxoisindolin-2-yl 1-methylcyclohexane-1-carboxylate (1c)



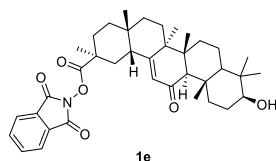
^1H NMR (600 MHz, Chloroform-*d*) δ 7.90 – 7.86 (m, 2H), 7.80 – 7.76 (m, 2H), 2.24 (d, J = 13.2 Hz, 2H), 1.70 – 1.62 (m, 3H), 1.61 – 1.55 (m, 2H), 1.43 (s, 3H), 1.42 – 1.35 (m, 2H), 1.31 – 1.25 (m, 1H).

1-(*tert*-Butyl) 4-(1,3-dioxoisindolin-2-yl) 4-methylpiperidine-1,4-dicarboxylate (1d)



^1H NMR (600 MHz, Chloroform-*d*) δ 7.90 – 7.87 (m, 2H), 7.81 – 7.78 (m, 2H), 4.05 – 3.85 (m, 2H), 3.18 – 3.08 (m, 2H), 2.28 – 2.22 (m, 2H), 1.56 – 1.50 (m, 2H), 1.48 (s, 3H), 1.46 (s, 9H).

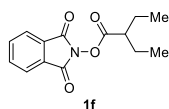
1,3-Dioxoisindolin-2-yl (2*S*,4*aS*,6*aS*,6*bR*,10*S*,12*aS*,12*bR*,14*bS*)-10-hydroxy-2,4*a*,6*a*,6*b*,9,9,12*a*-heptamethyl-13-oxo-1,2,3,4,4*a*,5,6,6*a*,6*b*,7,8,8*a*,9,10,11,12,12*a*,12*b*,13,14*b*-icosahydronicene-2-carboxylate (1e)



^1H NMR (400 MHz, Chloroform-*d*) δ 7.88 (dd, J = 5.5, 3.1 Hz, 2H), 7.79 (dd, J = 5.5, 3.1 Hz, 2H), 5.76 (s, 1H), 3.22 (dd, J = 10.8, 5.5 Hz, 1H), 2.79 (dt, J = 13.4, 3.6 Hz, 1H), 2.50 – 2.40 (m, 1H), 2.33 (s, 1H), 2.18 – 1.99 (m, 4H), 1.92 – 1.74 (m, 3H), 1.70 – 1.63 (m, 3H), 1.55 – 1.44 (m, 4H), 1.44 (s, 3H), 1.38 (s, 3H), 1.25 – 1.18 (m, 2H), 1.14 (s, 3H), 1.13 (s, 3H), 1.11 – 1.03 (m, 2H), 1.00 (s, 3H), 0.91 (s, 3H), 0.80 (s, 3H),

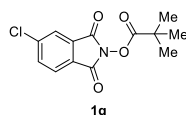
0.73 – 0.67 (m, 1H).

1,3-Dioxoisindolin-2-yl 2-ethylbutanoate (1f)



^1H NMR (600 MHz, Chloroform-*d*) δ 7.90 – 7.84 (m, 2H), 7.81 – 7.75 (m, 2H), 2.64 – 2.56 (m, 1H), 1.85 – 1.75 (m, 2H), 1.76 – 1.66 (m, 2H), 1.07 (t, J = 7.5 Hz, 6H).

5-Chloro-1,3-dioxoisindolin-2-yl pivalate (1g)

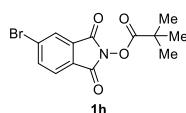


^1H NMR (400 MHz, Chloroform-*d*) δ 7.85 (d, J = 1.8 Hz, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.74 (dd, J = 8.0, 1.8 Hz, 1H), 1.43 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 161.3, 161.0, 141.6, 134.9, 130.8, 127.2, 125.3, 124.5, 38.6, 27.1.

HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_{12}\text{ClNNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 304.0347; found 304.0348.

5-Bromo-1,3-dioxoisindolin-2-yl pivalate (1h)

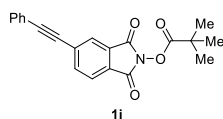


^1H NMR (400 MHz, Chloroform-*d*) δ 8.01 (d, J = 1.7 Hz, 1H), 7.92 (dd, J = 8.0, 1.7 Hz, 1H), 7.74 (d, J = 8.0 Hz, 1H), 1.43 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 161.5, 160.9, 137.8, 130.7, 129.8, 127.6, 127.3, 125.4, 38.6, 27.1.

HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_{12}\text{BrNNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 347.9842; found 347.9847.

1,3-Dioxo-5-(phenylethynyl)isindolin-2-yl pivalate (1i)

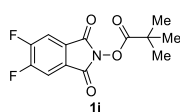


^1H NMR (400 MHz, Chloroform-*d*) δ 7.99 (s, 1H), 7.91 – 7.82 (m, 2H), 7.59 – 7.54 (m, 2H), 7.43 – 7.36 (m, 3H), 1.44 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 161.7, 161.6, 137.5, 132.0, 130.5, 129.50, 129.45, 128.7, 127.7, 126.7, 124.0, 122.1, 94.8, 87.6, 38.6, 27.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{21}\text{H}_{17}\text{NNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 370.1050; found 370.1050.

5,6-Difluoro-1,3-dioxoisindolin-2-yl pivalate (1j)



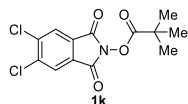
^1H NMR (400 MHz, Chloroform-*d*) δ 7.71 (t, J = 7.3 Hz, 2H), 1.42 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 160.4, 154.9 (dd, $J = 262.6, 14.9$ Hz), 126.2 (t, $J = 5.8$ Hz), 114.3 (dd, $J = 14.9, 7.7$ Hz), 38.6, 27.1.

^{19}F NMR (376 MHz, Chloroform-*d*) δ -123.93 (t, $J = 7.3$ Hz, 2F).

HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_{11}\text{F}_2\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 306.0548; found 306.0552.

5,6-Dichloro-1,3-dioxoisindolin-2-yl pivalate (1k)



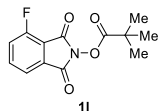
^1H NMR (400 MHz, Chloroform-*d*) δ 7.97 (s, 2H), 1.43 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.2, 160.4, 139.9, 128.2, 126.2, 38.6, 27.1.

^{13}C Quantitative-NMR (151 MHz, Chloroform-*d*) δ 174.3 (1C), 160.4 (2C), 139.9 (2C), 128.2 (2C), 126.2 (2C), 38.6 (1C), 27.1 (3C).

HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_{11}\text{Cl}_2\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 337.9957; found 337.9959.

4-Fluoro-1,3-dioxoisindolin-2-yl pivalate (1l)



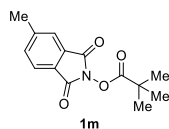
^1H NMR (400 MHz, Chloroform-*d*) δ 7.79 (ddd, $J = 8.4, 7.4, 4.4$ Hz, 1H), 7.71 (dd, $J = 7.4, 0.8$ Hz, 1H), 7.45 (td, $J = 8.4, 0.8$ Hz, 1H), 1.43 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.3, 161.1 (d, $J = 3.3$ Hz), 158.9 (d, $J = 0.9$ Hz), 157.7 (d, $J = 267.2$ Hz), 137.5 (d, $J = 7.9$ Hz), 131.0 (d, $J = 1.6$ Hz), 123.3 (d, $J = 19.7$ Hz), 120.2 (d, $J = 3.7$ Hz), 115.1 (d, $J = 12.8$ Hz), 38.5, 27.1.

^{19}F NMR (376 MHz, Chloroform-*d*) δ -111.05 (dd, $J = 8.7, 4.4$ Hz, 1F).

HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_{12}\text{FNNaO}_4^+ [\text{M}+\text{Na}]^+$ 288.0643; found 288.0642.

5-Methyl-1,3-dioxoisindolin-2-yl pivalate (1m)

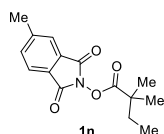


^1H NMR (400 MHz, Chloroform-*d*) δ 7.76 (d, $J = 7.6$ Hz, 1H), 7.68 (s, 1H), 7.56 (d, $J = 7.7$ Hz, 1H), 2.53 (s, 3H), 1.43 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 174.6, 162.5, 162.4, 146.3, 135.3, 129.5, 126.5, 124.5, 124.0, 38.5, 27.2, 22.3.

HR-MS (ESI-TOF) calcd for $\text{C}_{14}\text{H}_{15}\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 284.0893; found 284.0891.

5-Methyl-1,3-dioxoisindolin-2-yl 2,2-dimethylbutanoate (1n)

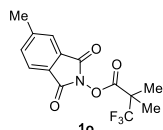


^1H NMR (400 MHz, Chloroform-*d*) δ 7.75 (d, J = 7.6 Hz, 1H), 7.67 (s, 1H), 7.56 (d, J = 7.6 Hz, 1H), 2.52 (s, 3H), 1.78 (q, J = 7.4 Hz, 2H), 1.38 (s, 6H), 1.04 (t, J = 7.4 Hz, 3H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 174.1, 162.6, 162.4, 146.2, 135.3, 129.5, 126.5, 124.5, 123.9, 42.6, 33.7, 24.8, 22.3, 9.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{15}\text{H}_{17}\text{NNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 298.1050; found 298.1049.

5-Methyl-1,3-dioxoisindolin-2-yl 3,3,3-trifluoro-2,2-dimethylpropanoate (1o)



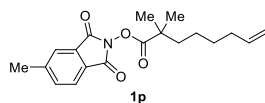
^1H NMR (400 MHz, Chloroform-*d*) δ 7.77 (d, J = 7.6 Hz, 1H), 7.70 (s, 1H), 7.59 (d, J = 7.2 Hz, 1H), 2.54 (s, 3H), 1.66 (s, 6H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 167.3, 162.8, 161.7, 146.6, 135.6, 129.2, 126.3, 125.6 (q, J = 283.8 Hz), 124.7, 124.2, 48.4 (q, J = 27.7 Hz), 22.3, 19.8 (q, J = 2.3 Hz).

^{19}F NMR (376 MHz, Chloroform-*d*) δ -74.94 (s, 3F).

HR-MS (ESI-TOF) calcd for $\text{C}_{14}\text{H}_{12}\text{F}_3\text{NNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 338.0611; found 338.0610.

5-Methyl-1,3-dioxoisindolin-2-yl 2,2-dimethyloct-7-enoate (1p)



^1H NMR (400 MHz, Chloroform-*d*) δ 7.74 (d, J = 7.6 Hz, 1H), 7.67 (s, 1H), 7.55 (d, J = 7.6 Hz, 1H), 5.89 – 5.77 (m, 1H), 5.05 – 4.88 (m, 2H), 2.52 (s, 3H), 2.14 – 2.06 (m, 2H), 1.75 – 1.67 (m, 2H), 1.49 – 1.42 (m, 4H), 1.38 (s, 6H).

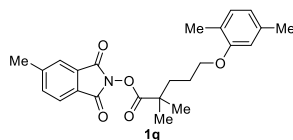
^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.0, 162.4, 162.3, 146.2, 138.9, 135.2, 129.4, 126.4, 124.4, 123.9, 114.5, 42.2, 40.6, 33.6, 29.3, 25.2, 24.3, 22.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{19}\text{H}_{23}\text{NNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 352.1519; found 352.1517.

5-Methyl-1,3-dioxoisindolin-2-yl

5-(2,5-dimethylphenoxy)-2,2-

dimethylpentanoate (1q)



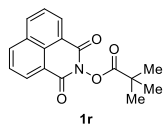
^1H NMR (400 MHz, Chloroform-*d*) δ 7.76 (d, J = 7.7 Hz, 1H), 7.68 (s, 1H), 7.57 (d, J = 7.7 Hz, 1H), 7.00 (d, J = 7.5 Hz, 1H), 6.68 – 6.63 (m, 2H), 4.01 (t, J = 3.4 Hz, 2H), 2.53 (s, 3H), 2.31 (s, 3H), 2.19 (s, 3H), 1.95 (d, J = 2.8 Hz, 4H), 1.44 (s, 6H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 173.9, 162.5, 162.4, 157.1, 146.3, 136.6, 135.3, 130.4, 129.4, 126.5, 124.5, 123.9, 123.7, 120.8, 112.1, 67.9, 42.1, 37.5, 25.2, 25.1, 22.3, 21.5, 15.9.

^{13}C Quantitative-NMR (151 MHz, Chloroform-*d*) δ 173.9 (1C), 162.5 (1C), 162.4 (1C), 157.1 (1C), 146.3 (1C), 136.6 (1C), 135.3 (1C), 130.4 (1C), 129.4 (1C), 126.4 (1C), 124.5 (1C), 123.9 (1C), 123.7 (1C), 120.8 (1C), 112.1(1C), 67.9 (1C), 42.1 (1C), 37.5 (1C), 25.2 (2C), 25.1 (1C), 22.3 (1C), 21.5 (1C), 15.9 (1C).

HR-MS (ESI-TOF) calcd for $\text{C}_{24}\text{H}_{27}\text{NNaO}_5^+ [\text{M}+\text{Na}]^+$ 432.1781; found 432.1782.

1,3-Dioxo-1*H*-benzo[*de*]isoquinolin-2(3*H*)-yl pivalate (1r)

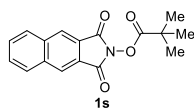


^1H NMR (400 MHz, Chloroform-*d*) δ 8.61 (dd, $J = 7.2, 1.2$ Hz, 2H), 8.27 (dd, $J = 8.4, 1.2$ Hz, 2H), 7.78 (ddd, $J = 8.4, 7.2, 1.2$ Hz, 2H), 1.51 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 159.8, 135.1, 132.0, 131.9, 127.7, 127.2, 122.5, 38.7, 27.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{17}\text{H}_{15}\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 320.0893; found 320.0894.

1,3-Dioxo-1,3-dihydro-2*H*-benzo[*f*]isoindol-2-yl pivalate (1s)

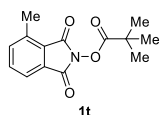


^1H NMR (400 MHz, Chloroform-*d*) δ 8.38 (s, 2H), 8.10 – 8.04 (m, 2H), 7.76 – 7.71 (m, 2H), 1.46 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.4, 161.8, 135.6, 130.5, 129.8, 125.8, 124.7, 38.6, 27.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{17}\text{H}_{15}\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 320.0893; found 320.0891.

4-Methyl-1,3-dioxoisindolin-2-yl pivalate (1t)

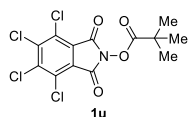


^1H NMR (400 MHz, Chloroform-*d*) δ 7.70 (d, $J = 7.6$ Hz, 1H), 7.63 (t, $J = 7.6$ Hz, 1H), 7.52 (d, $J = 7.6$ Hz, 1H), 2.69 (s, 3H), 1.43 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 174.6, 163.1, 162.3, 138.8, 137.2, 134.3, 129.6, 125.9, 121.7, 38.6, 27.2, 17.8.

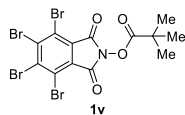
HR-MS (ESI-TOF) calcd for $\text{C}_{14}\text{H}_{15}\text{NNaO}_4^+ [\text{M}+\text{Na}]^+$ 284.0893; found 284.0895.

4,5,6,7-Tetrachloro-1,3-dioxoisindolin-2-yl pivalate (1u)



^1H NMR (400 MHz, Chloroform-*d*) δ 1.43 (s, 9H).

4,5,6,7-Tetrabromo-1,3-dioxoisindolin-2-yl pivalate (1v)

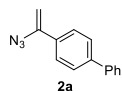


^1H NMR (400 MHz, Chloroform-*d*) δ 1.43 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 174.0, 158.3, 138.6, 128.0, 122.1, 38.6, 27.1.

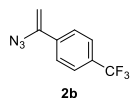
HR-MS (ESI-TOF) calcd for $\text{C}_{13}\text{H}_9\text{Br}_4\text{NNaO}_4^+$ $[\text{M}+\text{Na}]^+$ 581.7157; found 581.7163.

4-(1-Azidovinyl)-1,1'-biphenyl (2a)



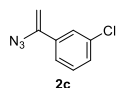
^1H NMR (600 MHz, Chloroform-*d*) δ 7.66 – 7.62 (m, 2H), 7.62 – 7.58 (m, 4H), 7.47 – 7.42 (m, 2H), 7.37 (td, J = 7.2, 1.2 Hz, 1H), 5.49 (d, J = 2.5 Hz, 1H), 4.99 (d, J = 2.5 Hz, 1H).

1-(1-Azidovinyl)-4-(trifluoromethyl)benzene (2b)



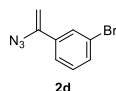
^1H NMR (600 MHz, Chloroform-*d*) δ 7.68 (d, J = 8.2 Hz, 2H), 7.61 (d, J = 8.2 Hz, 2H), 5.54 (d, J = 2.7 Hz, 1H), 5.07 (d, J = 2.7 Hz, 1H).

1-(1-Azidovinyl)-3-chlorobenzene (2c)



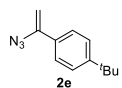
^1H NMR (600 MHz, Chloroform-*d*) δ 7.52 (s, 1H), 7.41 (d, J = 7.6 Hz, 1H), 7.30 – 7.24 (m, 2H), 5.44 – 5.42 (m, 1H), 4.97 – 4.95 (m, 1H).

1-(1-Azidovinyl)-3-bromobenzene (2d)



^1H NMR (400 MHz, Chloroform-*d*) δ 7.72 (t, J = 1.9 Hz, 1H), 7.48 (tdd, J = 7.9, 1.9, 1.0 Hz, 2H), 7.22 (t, J = 7.9 Hz, 1H), 5.46 (d, J = 2.7 Hz, 1H), 4.99 (d, J = 2.7 Hz, 1H).

1-(1-Azidovinyl)-4-(*tert*-butyl)benzene (2e)



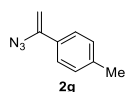
^1H NMR (400 MHz, Chloroform-*d*) δ 7.58 – 7.51 (m, 2H), 7.46 – 7.39 (m, 2H), 5.44 (d, J = 2.3 Hz, 1H), 4.96 (d, J = 2.3 Hz, 1H), 1.37 (s, 9H).

(1-Azidovinyl)benzene (2f)



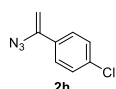
^1H NMR (400 MHz, Chloroform-*d*) δ 7.60 – 7.55 (m, 2H), 7.43 – 7.33 (m, 3H), 5.45 (d, J = 2.4 Hz, 1H), 4.97 (d, J = 2.4 Hz, 1H).

1-(1-Azidovinyl)-4-methylbenzene (2g)



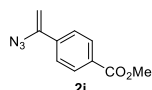
^1H NMR (400 MHz, Chloroform-*d*) δ 7.53 – 7.47 (m, 2H), 7.22 – 7.18 (m, 2H), 5.43 (d, J = 2.3 Hz, 1H), 4.95 (d, J = 2.3 Hz, 1H), 2.40 (s, 3H).

1-(1-Azidovinyl)-4-chlorobenzene (2h)



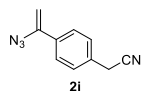
^1H NMR (600 MHz, Chloroform-*d*) δ 7.52 – 7.47 (m, 2H), 7.35 – 7.31 (m, 2H), 5.44 – 5.43 (m, 1H), 5.00 – 4.96 (m, 1H).

Methyl 4-(1-azidovinyl)benzoate (2i)



^1H NMR (400 MHz, Chloroform-*d*) δ 8.05 – 7.98 (m, 2H), 7.66 – 7.59 (m, 2H), 5.57 (d, J = 2.7 Hz, 1H), 5.07 (d, J = 2.7 Hz, 1H), 3.92 (s, 3H).

2-(4-(1-Azidovinyl)phenyl)acetonitrile (2j)

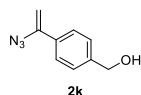


^1H NMR (400 MHz, Chloroform-*d*) δ 7.57 (d, J = 8.0 Hz, 2H), 7.32 (d, J = 8.0 Hz, 2H), 5.46 (d, J = 2.6 Hz, 1H), 4.99 (d, J = 2.6 Hz, 1H), 3.76 (s, 2H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 144.4, 134.3, 130.9, 128.2, 126.4, 117.6, 98.5, 23.5.

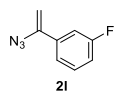
HR-MS (ESI-TOF) calcd for $\text{C}_{10}\text{H}_7\text{N}_4^-$ [$\text{M}-\text{H}$] $^-$ 183.0676; found 183.0669.

(4-(1-azidovinyl)phenyl)methanol (2k)



^1H NMR (400 MHz, Chloroform-*d*) δ 7.55 (d, J = 8.4 Hz, 2H), 7.35 (d, J = 8.4 Hz, 2H), 5.43 (d, J = 2.4 Hz, 1H), 4.96 (d, J = 2.4 Hz, 1H), 4.70 (d, J = 4.0 Hz, 2H).

1-(1-Azidovinyl)-3-fluorobenzene (2l)

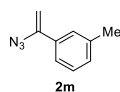


^1H NMR (400 MHz, Chloroform-*d*) δ 7.38 – 7.24 (m, 3H), 7.04 (tdd, J = 8.1, 2.6, 1.4

Hz, 1H), 5.47 (d, $J = 2.6$ Hz, 1H), 5.00 (d, $J = 2.6$ Hz, 1H).

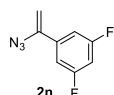
^{19}F NMR (376 MHz, Chloroform- d) δ -112.76 – -112.87 (m, 1F).

1-(1-Azidovinyl)-3-methylbenzene (2m)



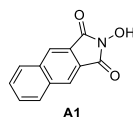
^1H NMR (400 MHz, Chloroform- d) δ 7.42 – 7.37 (m, 2H), 7.29 (t, $J = 3.8$ Hz, 1H), 7.19 (d, $J = 7.6$ Hz, 1H), 5.44 (d, $J = 2.4$ Hz, 1H), 4.97 (d, $J = 2.4$ Hz, 1H), 2.40 (s, 3H).

1-(1-Azidovinyl)-3,5-difluorobenzene (2n)



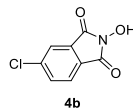
^1H NMR (400 MHz, Chloroform- d) δ 7.13 – 7.06 (m, 2H), 6.79 (tt, $J = 8.7, 2.3$ Hz, 1H), 5.49 (d, $J = 2.9$ Hz, 1H), 5.04 (d, $J = 2.9$ Hz, 1H).

2-Hydroxy-1H-benzo[f]isoindole-1,3(2H)-dione (A1)



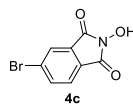
^1H NMR (600 MHz, DMSO- d_6) δ 10.97 (brs, 1H), 8.47 (s, 2H), 8.27 – 8.22 (m, 2H), 7.79 – 7.75 (m, 2H).

5-Chloro-2-hydroxyisoindoline-1,3-dione (4b)



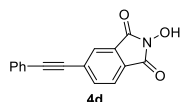
^1H NMR (400 MHz, DMSO- d_6) δ 10.94 (brs, 1H), 7.94 – 7.79 (m, 3H).

5-Bromo-2-hydroxyisoindoline-1,3-dione (4c)



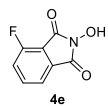
^1H NMR (400 MHz, DMSO- d_6) δ 10.92 (brs, 1H), 8.05 – 8.01 (m, 2H), 7.76 (d, $J = 8.3$ Hz, 1H).

2-Hydroxy-5-(phenylethynyl)isoindoline-1,3-dione (4d)



^1H NMR (400 MHz, DMSO- d_6) δ 10.91 (brs, 1H), 7.99 – 7.94 (m, 2H), 7.89 – 7.84 (m, 1H), 7.66 – 7.60 (m, 2H), 7.50 – 7.45 (m, 3H).

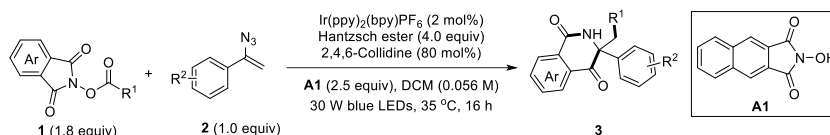
4-Fluoro-2-hydroxyisoindoline-1,3-dione (4e)



^1H NMR (400 MHz, DMSO- d_6) δ 10.91 (brs, 1H), 7.90 – 7.84 (m, 1H), 7.70 – 7.62 (m, 2H).

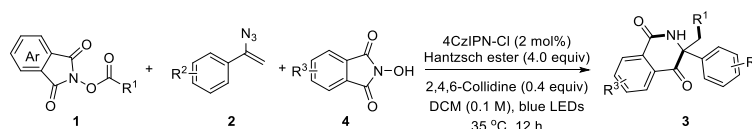
IV. General Procedures

Typical Procedure A



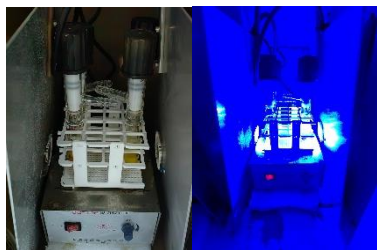
To a Schlenk tube were added **1** (0.18 mmol, 1.8 equiv), **2** (if solid) (0.10 mmol, 1.0 equiv), **A1** (53.4 mg, 0.25 mmol, 2.5 equiv), Ir(ppy)₂(bpy)PF₆ (1.6 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.8 mL), 2,4,6-collidine (9.7 mg, 0.08 mmol, 80 mol%) and **2** (if liquid) (0.10 mmol, 1.0 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring until TLC indicated the complete consumption of **2** (typically 16 h). The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography to afford the pure product (**3a-3p**).

Typical Procedure B



To a Schlenk tube were added **1** (0.18 mmol, 1.8 equiv), **2** (if solid) (0.10 mmol, 1.0 equiv), **4** (0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μL, 0.04 mmol, 40 mol%) and **2** (if liquid) (0.10 mmol, 1.0 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring until TLC indicated the complete consumption of **2** (typically 12 h). The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product (**3**).

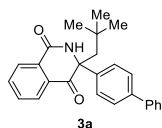
Reaction Setup



The reaction was carried out using the above setup with two 30W of blue LEDs (LED DRIVER YJ-30W, $\lambda = 460\text{-}470\text{ nm}$). There is 2.5 cm distance between the reactor and LEDs at 35 °C, which is monitored by a thermometer.

V. Characterization of Products

3-([1,1'-Biphenyl]-4-yl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3a**)



Following typical procedure A, the reaction of **1a** (44.5 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3a** as white solid (24.1 mg, 63% yield).

Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3a** as white solid (30.0 mg, 78% yield).

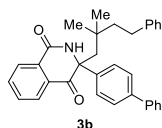
^1H NMR (400 MHz, Chloroform-*d*) δ 8.32 (dd, $J = 7.8, 1.3\text{ Hz}$, 1H), 8.03 (dd, $J = 7.8, 1.3\text{ Hz}$, 1H), 7.79 (td, $J = 7.6, 1.3\text{ Hz}$, 1H), 7.68 (td, $J = 7.6, 1.3\text{ Hz}$, 1H), 7.61 – 7.55 (m, 2H), 7.55 – 7.47 (m, 4H), 7.43 – 7.37 (m, 2H), 7.36 – 7.29 (m, 1H), 6.58 (brs, 1H), 3.13 (d, $J = 14.9\text{ Hz}$, 1H), 1.88 (d, $J = 14.9\text{ Hz}$, 1H), 1.01 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 192.5, 162.7, 141.3, 141.1, 140.2, 135.0, 133.3, 131.1, 131.0, 128.9, 128.5, 127.70, 127.68, 127.4, 127.2, 126.1, 70.9, 52.8, 32.0, 31.7.

^{13}C Quantitative-NMR (151 MHz, Chloroform-*d*) δ 192.4 (1C), 162.6 (1C), 141.3 (1C), 141.2 (1C), 140.2 (1C), 135.0 (1C), 133.4 (1C), 131.1 (1C), 131.0 (1C), 128.9 (2C), 128.5 (1C), 127.72 (1C), 127.69 (2C), 127.4 (1C), 127.2 (2C), 126.0 (2C), 70.9 (1C), 52.8 (1C), 32.0 (1C), 31.7 (3C).

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{25}\text{NNaO}_2^+$ $[\text{M}+\text{Na}]^+$ 406.1778 found 406.1780.

3-([1,1'-Biphenyl]-4-yl)-3-(2,2-dimethyl-4-phenylbutyl)-2,3-dihydroisoquinoline-1,4-dione (**3b**)



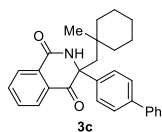
Following typical procedure A, the reaction of **1b** (60.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3b** as white solid (24.1 mg, 51% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.32 (dd, *J* = 7.8, 1.3 Hz, 1H), 8.03 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.79 (td, *J* = 7.6, 1.4 Hz, 1H), 7.68 (td, *J* = 7.6, 1.4 Hz, 1H), 7.61 – 7.56 (m, 2H), 7.55 – 7.49 (m, 4H), 7.44 – 7.38 (m, 2H), 7.36 – 7.31 (m, 1H), 7.28 – 7.22 (m, 2H), 7.19 – 7.12 (m, 3H), 6.57 (brs, 1H), 3.19 (d, *J* = 14.9 Hz, 1H), 2.68 – 2.59 (m, 2H), 1.97 (d, *J* = 14.9 Hz, 1H), 1.66 – 1.59 (m, 2H), 1.13 (s, 3H), 1.00 (s, 3H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 192.4, 162.7, 142.8, 141.3, 141.2, 140.2, 135.1, 133.4, 131.0, 130.9, 129.0, 128.5, 128.4, 127.7, 127.4, 127.2, 126.1, 125.8, 70.8, 50.9, 47.1, 34.6, 30.7, 29.4, 28.4.

¹³C Quantitative-NMR (151 MHz, Chloroform-*d*) δ 192.4 (1C), 162.6 (1C), 142.8 (1C), 141.3 (1C), 141.2 (1C), 140.2 (1C), 135.1 (1C), 133.4 (1C), 131.0 (1C), 130.9 (1C), 129.0 (2C), 128.5 (3C), 128.4 (2C), 127.7 (3C), 127.4 (1C), 127.2 (2C), 126.0 (2C), 125.8 (1C), 70.8 (1C), 50.9 (1C), 47.1 (1C), 34.6 (1C), 30.7 (1C), 29.4 (1C), 28.4 (1C). HR-MS (ESI-TOF) calcd for C₃₃H₃₁NNaO₂⁺ [M+Na]⁺ 496.2247; found 496.2249.

3-([1,1'-Biphenyl]-4-yl)-3-((1-methylcyclohexyl)methyl)-2,3-dihydroisoquinoline-1,4-dione (3c)



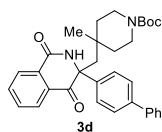
Following typical procedure A, the reaction of **1c** (51.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3c** as white solid (22.2 mg, 52% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.32 (dd, *J* = 7.8, 1.3 Hz, 1H), 8.04 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.78 (td, *J* = 7.6, 1.4 Hz, 1H), 7.68 (td, *J* = 7.6, 1.4 Hz, 1H), 7.63 – 7.57 (m, 2H), 7.55 – 7.47 (m, 4H), 7.44 – 7.36 (m, 2H), 7.36 – 7.28 (m, 1H), 6.84 (brs, 1H), 3.06 (d, *J* = 14.9 Hz, 1H), 1.96 (d, *J* = 14.9 Hz, 1H), 1.53 – 1.60 (m, 1H), 1.48 – 1.35 (m, 6H), 1.31 – 1.19 (m, 3H), 0.96 (s, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 192.5, 162.6, 141.5, 141.1, 140.2, 135.0, 133.4, 131.0, 130.9, 128.9, 128.5, 127.71, 127.68, 127.4, 127.2, 126.1, 70.8, 53.3, 40.0, 39.7, 34.6, 26.2, 24.9, 22.1, 21.9.

HR-MS (ESI-TOF) calcd for C₂₉H₂₉NNaO₂⁺ [M+Na]⁺ 446.2091; found 446.2090.

***tert*-Butyl 4-((3-([1,1'-biphenyl]-4-yl)-1,4-dioxo-1,2,3,4-tetrahydroisoquinolin-3-yl)methyl)-4-methylpiperidine-1-carboxylate (3d)**



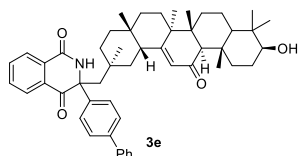
Following typical procedure A, the reaction of **1d** (69.8 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3d** as white solid (26.0 mg, 50% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.31 (dd, *J* = 7.8, 1.3 Hz, 1H), 8.05 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.80 (td, *J* = 7.6, 1.4 Hz, 1H), 7.69 (td, *J* = 7.6, 1.4 Hz, 1H), 7.61 – 7.56 (m, 2H), 7.55 – 7.47 (m, 4H), 7.43 – 7.37 (m, 2H), 7.36 – 7.29 (m, 1H), 7.00 (brs, 1H), 3.79 – 3.64 (m, 2H), 3.11 (d, *J* = 14.9 Hz, 1H), 3.05 – 2.94 (m, 2H), 1.94 (d, *J* = 14.9 Hz, 1H), 1.65 – 1.51 (m, 3H) 1.42 (s, 9H), 1.29 – 1.23 (m, 1H), 1.02 (s, 3H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 192.3, 162.6, 155.0, 141.4, 141.1, 140.1, 135.2, 133.6, 130.9, 130.8, 129.0, 128.6, 127.8, 127.5, 127.2, 126.0, 79.6, 70.6, 53.0, 38.8, 38.6, 33.1, 28.6, 23.2.

HR-MS (ESI-TOF) calcd for C₃₃H₃₆N₂NaO₄⁺ [M+Na]⁺ 547.2567; found 547.2569.

3-([1,1'-Biphenyl]-4-yl)-3-(((2*S*,4*aR*,6*aS*,6*bR*,10*S*,12*aS*,12*bR*,14*bS*)-10-hydroxy-2,4*a*,6*a*,6*b*,9,9,12*a*-heptamethyl-13-oxo-1,2,3,4,4*a*,5,6,6*a*,6*b*,7,8,8*a*,9,10,11,12,12*a*,12*b*,13,14*b*-icosahydricen-2-yl)methyl)-2,3-dihydroisoquinoline-1,4-dione (3e**)**



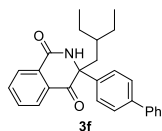
Following typical procedure A, the reaction of **1e** (110.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3e** as white solid (32.2 mg, 44% yield, dr = 1:1:1:2).

¹H NMR (600 MHz, Chloroform-*d*) δ 8.31 (dd, *J* = 7.8, 1.2 Hz, 1H), 8.03 (ddd, *J* = 7.8, 5.7, 1.2 Hz, 1H), 7.81 – 7.75 (m, 1H), 7.68 (dtd, *J* = 12.4, 7.6, 1.3 Hz, 1H), 7.60 – 7.56 (m, 2H), 7.53 – 7.46 (m, 4H), 7.39 (td, *J* = 7.8, 2.7 Hz, 2H), 7.34 – 7.30 (m, 1H), 6.84 (brs, 1H), 5.47 (s, 1H), 3.22 (dt, *J* = 10.5, 4.7 Hz, 1H), 3.07 (d, *J* = 14.9 Hz, 1H), 2.78 – 2.74 (m, 1H), 2.25 (s, 1H), 2.15 – 2.08 (m, 1H), 2.03 – 1.93 (m, 1H), 1.85 (d, *J* = 14.9 Hz, 1H), 1.67 – 1.53 (m, 4H), 1.47 – 1.35 (m, 3H), 1.29 – 1.18 (m, 4H), 1.11 (d, *J* = 2.1 Hz, 6H), 1.08 (s, 6H), 1.00 (s, 3H), 0.97 (s, 3H), 0.96 – 0.85 (m, 3H), 0.81 (s, 6H), 0.70 – 0.61 (m, 1H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 200.2, 192.2, 169.6, 162.6, 141.3, 141.1, 140.1, 135.2, 133.4, 130.9, 129.0, 128.6, 128.5, 128.3, 127.7, 127.4, 127.1, 126.0, 78.9, 70.6, 61.9, 55.3, 55.0, 47.1, 45.6, 44.6, 43.4, 39.2, 37.3, 35.9, 35.5, 34.4, 33.6, 32.8, 32.3, 28.7, 28.2, 27.4, 26.42, 26.36, 23.7, 23.2, 22.5, 18.8, 17.6, 16.4, 15.7.

HR-MS (ESI-TOF) calcd for C₅₁H₆₁NNaO₄⁺ [M+Na]⁺ 774.4493; found 774.4496.

3-([1,1'-Biphenyl]-4-yl)-3-(2-ethylbutyl)-2,3-dihydroisoquinoline-1,4-dione (3f**)**



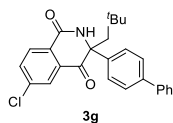
Following typical procedure A, the reaction of **1f** (47.0 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), 4CzIPN (1.6 mg, 0.002 mmol), DIPEA (10.4 mg, 0.08 mmol) in DCM (1.0 mL) for 16 h afforded **3f** as white solid (8.0 mg, 20% yield).

^1H NMR (600 MHz, Chloroform-*d*) δ 8.30 (d, J = 7.8 Hz, 1H), 8.01 (d, J = 7.8 Hz, 1H), 7.78 (t, J = 7.6 Hz, 1H), 7.68 (t, J = 7.6 Hz, 1H), 7.61 – 7.56 (m, 2H), 7.55 – 7.49 (m, 4H), 7.40 (t, J = 7.6 Hz, 2H), 7.33 (t, J = 7.4 Hz, 1H), 6.63 (brs, 1H), 2.79 (dd, J = 14.6, 7.0 Hz, 1H), 2.00 (dd, J = 14.6, 5.0 Hz, 1H), 1.56 – 1.48 (m, 1H), 1.36 – 1.30 (m, 2H), 1.25 – 1.19 (m, 2H), 0.86 – 0.78 (m, 6H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 192.8, 163.2, 141.2, 140.5, 140.2, 135.1, 133.4, 131.12, 131.10, 129.0, 128.5, 127.74, 127.69, 127.3, 127.2, 126.2, 70.7, 43.9, 36.8, 26.2, 26.0, 10.7, 10.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{27}\text{H}_{28}\text{NO}_2^+ [\text{M}+\text{H}]^+$ 398.2115; found 398.2114.

3-([1,1'-Biphenyl]-4-yl)-6-chloro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3g**)



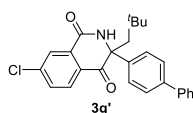
Following typical procedure A, the reaction of **1g** (50.6 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3g** and **3g'** as white solid (28.8 mg, 69% yield, rr = 1:1).

^1H NMR (400 MHz, Chloroform-*d*) δ 8.26 (d, J = 8.4 Hz, 1H), 7.98 (d, J = 2.2 Hz, 1H), 7.72 (dd, J = 8.4, 2.2 Hz, 1H), 7.58 – 7.48 (m, 6H), 7.41 (t, J = 7.6 Hz, 2H), 7.37 – 7.31 (m, 1H), 6.74 (brs, 1H), 3.10 (d, J = 14.9 Hz, 1H), 1.87 (d, J = 14.9 Hz, 1H), 1.00 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 191.4, 161.9, 141.4, 140.7, 140.3, 140.1, 135.1, 132.1, 130.4, 129.3, 129.0, 127.8, 127.3, 127.2, 126.0, 71.3, 52.6, 31.9, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{25}\text{ClNO}_2^+ [\text{M}+\text{H}]^+$ 418.1568; found 418.1569.

3-([1,1'-Biphenyl]-4-yl)-7-chloro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3g'**)

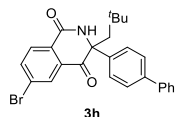


^1H NMR (400 MHz, Chloroform-*d*) δ 8.29 (d, J = 2.1 Hz, 1H), 7.97 (d, J = 8.4 Hz, 1H), 7.62 (dd, J = 8.4, 2.1 Hz, 1H), 7.59 – 7.48 (m, 6H), 7.44 – 7.37 (m, 2H), 7.37 – 7.30 (m, 1H), 6.77 (brs, 1H), 3.11 (d, J = 14.9 Hz, 1H), 1.90 (d, J = 14.9 Hz, 1H), 1.01 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 191.4, 161.5, 142.1, 141.4, 140.9, 140.1, 133.7, 132.5, 129.24, 129.19, 129.0, 128.6, 127.8, 127.2, 126.0, 71.0, 52.6, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{25}\text{ClNO}_2^+ [\text{M}+\text{H}]^+$ 418.1568; found 418.1570.

3-([1,1'-Biphenyl]-4-yl)-6-bromo-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3h)



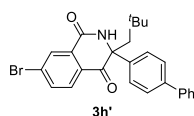
Following typical procedure A, the reaction of **1h** (58.5 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3h** and **3h'** as white solid (33.2 mg, 72% yield, rr = 1:1).

^1H NMR (600 MHz, Chloroform-*d*) δ 8.18 (d, J = 8.3 Hz, 1H), 8.14 (d, J = 2.0 Hz, 1H), 7.89 (dd, J = 8.3, 2.0 Hz, 1H), 7.57 – 7.48 (m, 6H), 7.41 (t, J = 7.6 Hz, 2H), 7.33 (t, J = 7.4 Hz, 1H), 6.55 (brs, 1H), 3.11 (d, J = 14.9 Hz, 1H), 1.89 (d, J = 14.9 Hz, 1H), 1.01 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 191.2, 161.9, 141.5, 140.7, 140.1, 138.1, 132.1, 130.4, 130.3, 129.7, 129.0, 128.7, 127.84, 127.82, 127.2, 126.0, 71.3, 52.6, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{24}\text{BrNNaO}_2^+ [\text{M}+\text{Na}]^+$ 484.0883; found 484.0884.

3-([1,1'-Biphenyl]-4-yl)-7-bromo-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3h')

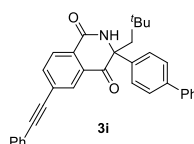


^1H NMR (400 MHz, Chloroform-*d*) δ 8.47 (d, J = 2.0 Hz, 1H), 7.88 (d, J = 8.3 Hz, 1H), 7.80 (dd, J = 8.3, 2.0 Hz, 1H), 7.58 – 7.48 (m, 6H), 7.44 – 7.38 (m, 2H), 7.37 – 7.31 (m, 1H), 6.58 (brs, 1H), 3.11 (d, J = 14.9 Hz, 1H), 1.89 (d, J = 14.9 Hz, 1H), 1.01 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 191.6, 161.3, 141.4, 140.8, 140.1, 136.7, 132.3, 131.6, 130.8, 129.6, 129.1, 129.0, 127.8, 127.2, 126.0, 71.0, 52.6, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{24}\text{BrNNaO}_2^+ [\text{M}+\text{Na}]^+$ 484.0883; found 484.0884.

3-([1,1'-Biphenyl]-4-yl)-3-neopentyl-6-(phenylethynyl)-2,3-dihydroisoquinoline-1,4-dione (3i)



Following typical procedure A, the reaction of **1i** (62.5 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3i** and **3i'** as white solid (32.0 mg, 66% yield, rr = 1:1).

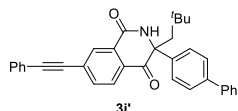
^1H NMR (600 MHz, Chloroform-*d*) δ 8.30 (d, J = 8.0 Hz, 1H), 8.16 (d, J = 1.8 Hz, 1H), 7.88 (dd, J = 8.0, 1.8 Hz, 1H), 7.60 – 7.56 (m, 2H), 7.55 – 7.48 (m, 6H), 7.43 – 7.38

(m, 2H), 7.39 – 7.30 (m, 4H), 6.66 (brs, 1H), 3.13 (d, $J = 14.9$ Hz, 1H), 1.90 (d, $J = 14.9$ Hz, 1H), 1.03 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 191.8, 162.2, 141.3, 141.0, 140.2, 137.4, 132.0, 131.0, 130.4, 129.9, 129.2, 128.98, 128.95, 128.7, 128.6, 127.78, 127.76, 127.2, 126.0, 122.4, 93.8, 87.7, 71.1, 52.6, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{34}\text{H}_{30}\text{NO}_2^+ [\text{M}+\text{H}]^+$ 484.2271; found 484.2271.

3-([1,1'-Biphenyl]-4-yl)-3-neopentyl-7-(phenylethynyl)-2,3-dihydroisoquinoline-1,4-dione (3i')

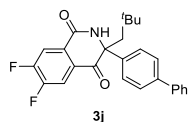


^1H NMR (600 MHz, Chloroform- d) δ 8.46 (s, 1H), 8.02 (d, $J = 8.0$ Hz, 1H), 7.77 (d, $J = 8.0$ Hz, 1H), 7.64 – 7.60 (m, 2H), 7.58 – 7.49 (m, 6H), 7.43 – 7.35 (m, 5H), 7.33 (t, $J = 7.4$ Hz, 1H), 7.20 (brs, 1H), 3.14 (d, $J = 14.8$ Hz, 1H), 1.96 (d, $J = 14.8$ Hz, 1H), 1.04 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 191.8, 162.2, 141.2, 141.2, 140.2, 135.8, 132.0, 131.5, 131.2, 130.6, 129.8, 129.3, 128.9, 128.6, 127.7, 127.5, 127.2, 126.1, 122.4, 94.9, 88.1, 70.9, 52.8, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{34}\text{H}_{29}\text{NNaO}_2^+ [\text{M}+\text{Na}]^+$ 506.2091; found 506.2092.

3-([1,1'-Biphenyl]-4-yl)-6,7-difluoro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3j)



Following typical procedure A, the reaction of **1j** (50.9 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3j** as white solid (28.9 mg, 69% yield).

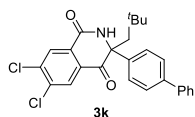
^1H NMR (400 MHz, Chloroform- d) δ 8.10 (dd, $J = 9.9, 7.3$ Hz, 1H), 7.81 (dd, $J = 9.6, 7.4$ Hz, 1H), 7.57 – 7.49 (m, 6H), 7.45 – 7.37 (m, 2H), 7.37 – 7.32 (m, 1H), 6.73 (brs, 1H), 3.10 (d, $J = 14.9$ Hz, 1H), 1.90 (d, $J = 14.9$ Hz, 1H), 1.01 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 190.4, 160.8, 154.9 (dd, $J = 261.1, 13.2$ Hz), 153.9 (dd, $J = 259.5, 13.5$ Hz), 141.5, 140.6, 140.0, 129.3 (dd, $J = 6.7, 3.5$ Hz), 129.0, 128.8 (dd, $J = 5.5, 3.6$ Hz), 127.9, 127.2, 125.9, 118.0 (d, $J = 19.7$ Hz), 116.7 (d, $J = 18.9$ Hz), 71.3, 52.7, 32.0, 31.7.

^{19}F NMR (376 MHz, Chloroform- d) δ -124.7 (ddd, $J = 20.9, 10.0, 7.4$ Hz, 1F), -127.7 (ddd, $J = 20.8, 9.7, 7.2$ Hz, 1F).

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{24}\text{F}_2\text{NO}_2^+ [\text{M}+\text{H}]^+$ 420.1770; found 420.1769.

3-([1,1'-Biphenyl]-4-yl)-6,7-dichloro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3k)

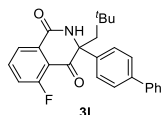


Following typical procedure A, the reaction of **1k** (56.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3k** as white solid (27.5 mg, 61% yield).

^1H NMR (600 MHz, Chloroform-*d*) δ 8.38 (s, 1H), 8.08 (s, 1H), 7.56 – 7.49 (m, 6H), 7.41 (t, J = 7.6 Hz, 2H), 7.34 (t, J = 7.3 Hz, 1H), 6.88 (brs, 1H), 3.10 (d, J = 14.9 Hz, 1H), 1.91 (d, J = 14.9 Hz, 1H), 1.01 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 190.6, 161.0, 141.5, 140.5, 140.3, 140.0, 138.7, 130.7, 130.3, 130.0, 129.4, 129.0, 127.88, 127.85, 127.2, 125.9, 71.4, 52.6, 32.0, 31.7. HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{24}\text{Cl}_2\text{NO}_2^+ [\text{M}+\text{H}]^+$ 452.1179; found 452.1180.

3-([1,1'-Biphenyl]-4-yl)-5-fluoro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3l**)**



Following typical procedure A, the reaction of **1l** (47.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3l** and **3l'** as white solid (18.0 mg, 45% yield, rr = 1:1).

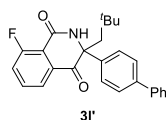
^1H NMR (600 MHz, Chloroform-*d*) δ 7.87 (d, J = 7.8 Hz, 1H), 7.67 – 7.60 (m, 1H), 7.60 – 7.55 (m, 2H), 7.55 – 7.49 (m, 4H), 7.49 – 7.42 (m, 1H), 7.43 – 7.38 (m, 2H), 7.36 – 7.30 (m, 1H), 6.44 (brs, 1H), 3.05 (d, J = 14.9 Hz, 1H), 1.88 (d, J = 14.9 Hz, 1H), 1.02 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 191.5 (d, J = 3.4 Hz), 161.9 (d, J = 266.6 Hz), 160.2 (d, J = 3.5 Hz), 141.4, 140.7, 140.1, 134.7 (d, J = 9.5 Hz), 132.8, 129.0, 127.80, 127.78, 127.2, 126.0, 123.9 (d, J = 5.6 Hz), 123.8 (d, J = 13.0 Hz), 118.4 (d, J = 4.4 Hz), 70.8, 52.6, 31.9, 31.8.

^{19}F NMR (565 MHz, Chloroform-*d*) δ -109.86 – -109.92 (m, 1F).

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{25}\text{FNO}_2^+ [\text{M}+\text{H}]^+$ 402.1864; found 402.1862.

3-([1,1'-Biphenyl]-4-yl)-8-fluoro-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3l'**)**



^1H NMR (600 MHz, Chloroform-*d*) δ 8.15 (d, J = 7.8 Hz, 1H), 7.73 (td, J = 8.0, 4.4 Hz, 1H), 7.57 – 7.49 (m, 6H), 7.43 – 7.38 (m, 2H), 7.38 – 7.31 (m, 2H), 6.68 (brs, 1H), 3.07 (d, J = 14.9 Hz, 1H), 1.87 (d, J = 14.9 Hz, 1H), 1.03 (s, 9H).

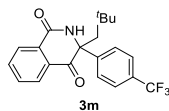
^{13}C NMR (151 MHz, Chloroform-*d*) δ 189.3 (d, J = 1.3 Hz), 161.6 (d, J = 3.2 Hz), 161.3 (d, J = 269.3 Hz), 141.4, 140.6, 140.1, 136.3 (d, J = 9.8 Hz), 132.7, 129.0, 127.81,

127.79, 127.2, 126.0, 124.7 (d, $J = 3.7$ Hz), 122.0 (d, $J = 21.4$ Hz), 119.2 (d, $J = 5.4$ Hz), 71.2, 52.0, 31.9, 31.8.

^{19}F NMR (565 MHz, Chloroform- d) δ -110.87 – -110.92 (m, 1F)

HR-MS (ESI-TOF) calcd for $\text{C}_{26}\text{H}_{25}\text{FNO}_2^+ [\text{M}+\text{H}]^+ 402.1864$; found 402.1862.

3-Neopentyl-3-(4-(trifluoromethyl)phenyl)-2,3-dihydroisoquinoline-1,4-dione (3m)



Following typical procedure A, the reaction of **1a** (44.5 mg, 0.18 mmol), **2b** (21.3 mg, 0.10 mmol) for 16 h afforded **3m** as white solid (19.0 mg, 51% yield).

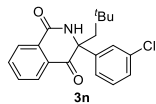
^1H NMR (400 MHz, Chloroform- d) δ 8.29 (d, $J = 7.8$ Hz, 1H), 8.01 (d, $J = 7.8$ Hz, 1H), 7.79 (tt, $J = 7.6, 1.4$ Hz, 1H), 7.72 – 7.64 (m, 3H), 7.58 – 7.53 (m, 2H), 7.10 (brs, 1H), 3.07 (d, $J = 14.8$ Hz, 1H), 1.87 (d, $J = 14.8$ Hz, 1H), 1.00 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 192.1, 162.6, 146.3, 135.3, 133.6, 131.0, 130.8, 130.5 (q, $J = 32.7$ Hz), 128.6, 127.4, 126.1, 126.0 (q, $J = 3.7$ Hz), 123.9 (q, $J = 272.4$ Hz), 70.9, 52.9, 32.1, 31.7.

^{19}F NMR (376 MHz, Chloroform- d) δ -62.73 (s, 3F).

HR-MS (ESI-TOF) calcd for $\text{C}_{21}\text{H}_{21}\text{F}_3\text{NO}_2^+ [\text{M}+\text{H}]^+ 376.1519$; found 376.1521.

3-(3-Chlorophenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3n)



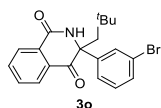
Following typical procedure A, the reaction of **1a** (44.5 mg, 0.18 mmol), **2c** (17.9 mg, 0.10 mmol) for 16 h afforded **3n** as white solid (17.7 mg, 52% yield).

^1H NMR (400 MHz, Chloroform- d) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.02 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.79 (td, $J = 7.6, 1.4$ Hz, 1H), 7.69 (td, $J = 7.6, 1.4$ Hz, 1H), 7.54 – 7.51 (m, 1H), 7.45 – 7.38 (m, 1H), 7.25 – 7.19 (m, 2H), 6.74 (brs, 1H), 3.03 (d, $J = 14.8$ Hz, 1H), 1.83 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (101 MHz, Chloroform- d) δ 192.0, 162.4, 144.4, 135.2, 135.1, 133.5, 130.9, 130.8, 130.2, 128.6, 128.5, 127.4, 126.0, 123.8, 70.6, 52.9, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{20}\text{H}_{20}\text{ClNNaO}_2^+ [\text{M}+\text{Na}]^+ 364.1075$; found 364.1075.

3-(3-Bromophenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3o)



Following typical procedure A, the reaction of **1a** (44.5 mg, 0.18 mmol), **2d** (22.3 mg, 0.10 mmol) for 16 h afforded **3o** as white solid (20.0 mg, 52% yield).

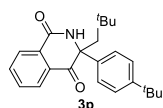
^1H NMR (400 MHz, Chloroform- d) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.02 (dd, $J = 7.8,$

1.3 Hz, 1H), 7.80 (td, $J = 7.6, 1.4$ Hz, 1H), 7.72 – 7.65 (m, 2H), 7.46 (ddd, $J = 8.1, 1.9, 1.0$ Hz, 1H), 7.38 (ddd, $J = 8.1, 1.9, 1.0$ Hz, 1H), 7.17 (t, $J = 8.0$ Hz, 1H), 6.65 (brs, 1H), 3.02 (d, $J = 14.8$ Hz, 1H), 1.82 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (101 MHz, Chloroform- d) δ 192.0, 162.3, 144.6, 135.2, 133.5, 131.4, 130.9, 130.8, 130.5, 128.8, 128.6, 127.4, 124.2, 123.3, 70.6, 53.0, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{20}\text{H}_{20}\text{BrNNaO}_2^+ [\text{M}+\text{Na}]^+$ 408.0570; found 408.0572.

3-(4-(*tert*-Butyl)phenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (3p)



Following typical procedure A, the reaction of **1a** (44.5 mg, 0.18 mmol), **2e** (20.1 mg, 0.10 mmol), HOAc (4.8 mg, 0.08 mmol) for 16 h afforded **3p** as white solid (13.1 mg, 36% yield).

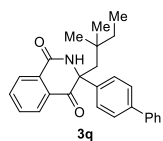
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2e** (20.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3p** as white solid (15.2 mg, 42% yield).

^1H NMR (600 MHz, Chloroform- d) δ 8.30 (d, $J = 7.8$ Hz, 1H), 8.01 (d, $J = 7.8$ Hz, 1H), 7.76 (t, $J = 7.6$ Hz, 1H), 7.65 (t, $J = 7.6$ Hz, 1H), 7.47 – 7.42 (m, 2H), 7.32 – 7.28 (m, 2H), 6.92 (brs, 1H), 3.10 (d, $J = 14.8$ Hz, 1H), 1.84 (d, $J = 14.8$ Hz, 1H), 1.24 (s, 9H), 0.99 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 192.6, 162.6, 151.2, 139.4, 134.9, 133.2, 131.2, 131.0, 128.5, 127.3, 126.0, 125.2, 70.8, 52.8, 34.5, 31.9, 31.7, 31.3.

HR-MS (ESI-TOF) calcd for $\text{C}_{24}\text{H}_{29}\text{NNaO}_2^+ [\text{M}+\text{Na}]^+$ 386.2091; found 386.2093.

3-([1,1'-Biphenyl]-4-yl)-3-(2,2-dimethylbutyl)-2,3-dihydroisoquinoline-1,4-dione (3q)



Following typical procedure B, the reaction of **1n** (49.5 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3q** as white solid (27.0 mg, 68% yield).

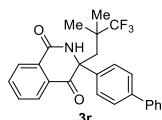
^1H NMR (600 MHz, Chloroform- d) δ 8.32 (d, $J = 7.8$ Hz, 1H), 8.04 (d, $J = 7.8$ Hz, 1H), 7.78 (t, $J = 7.6$ Hz, 1H), 7.68 (t, $J = 7.6$ Hz, 1H), 7.60 (d, $J = 8.7$ Hz, 2H), 7.55 – 7.49 (m, 4H), 7.40 (t, $J = 7.6$ Hz, 2H), 7.35 – 7.30 (m, 1H), 6.80 (brs, 1H), 3.09 (d, $J = 15.0$ Hz, 1H), 1.91 (d, $J = 15.0$ Hz, 1H), 1.36 (q, $J = 7.4$ Hz, 2H), 1.01 (s, 3H), 0.91 (s, 3H), 0.88 (t, $J = 7.4$ Hz, 3H).

^{13}C NMR (151 MHz, Chloroform- d) δ 192.5, 162.6, 141.4, 141.2, 140.2, 135.0, 133.4,

131.1, 131.0, 128.9, 128.5, 127.71, 126.69, 127.4, 127.2, 126.1, 70.8, 50.7, 37.1, 34.5, 28.9, 27.9, 8.6.

HR-MS (ESI-TOF) calcd for $C_{27}H_{27}NNaO_2^+$ $[M+Na]^+$ 420.1934; found 420.1938.

3-([1,1'-Biphenyl]-4-yl)-3-(3,3,3-trifluoro-2,2-dimethylpropyl)-2,3-dihydroisoquinoline-1,4-dione (3r)



Following typical procedure B, the reaction of **1o** (56.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3r** as white solid (21.9 mg, 50% yield).

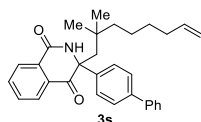
1H NMR (600 MHz, Chloroform-*d*) δ 8.31 (d, J = 7.8 Hz, 1H), 8.05 (d, J = 7.8 Hz, 1H), 7.80 (t, J = 7.6 Hz, 1H), 7.70 (t, J = 7.6 Hz, 1H), 7.58 (d, J = 8.4 Hz, 2H), 7.54 (d, J = 8.5 Hz, 2H), 7.50 (d, J = 7.8 Hz, 2H), 7.41 (t, J = 7.6 Hz, 2H), 7.34 (t, J = 7.3 Hz, 1H), 6.77 (brs, 1H), 3.41 (d, J = 15.3 Hz, 1H), 2.11 (d, J = 15.3 Hz, 1H), 1.32 (s, 3H), 1.13 (s, 3H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 191.7, 162.8, 141.7, 140.2, 140.0, 135.4, 133.7, 130.8, 130.7, 129.04 (q, J = 283.3 Hz), 129.00, 128.7, 128.0, 127.9, 127.6, 127.2, 126.0, 69.9, 43.8, 41.2 (q, J = 24.5 Hz), 22.8 (q, J = 2.0 Hz), 21.9 (q, J = 2.0 Hz).

^{19}F NMR (376 MHz, Chloroform-*d*) δ -79.12 (s, 3F).

HR-MS (ESI-TOF) calcd for $C_{26}H_{22}F_3NNaO_2^+$ $[M+Na]^+$ 460.1495; found 460.1496.

3-([1,1'-Biphenyl]-4-yl)-3-(2,2-dimethyloct-7-en-1-yl)-2,3-dihydroisoquinoline-1,4-dione (3s)



Following typical procedure B, the reaction of **1p** (59.3 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3s** as white solid (27.1 mg, 60% yield).

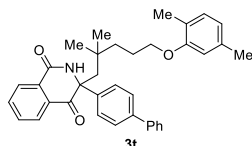
1H NMR (600 MHz, Chloroform-*d*) δ 8.32 (d, J = 7.8 Hz, 1H), 8.03 (d, J = 7.8 Hz, 1H), 7.79 (t, J = 7.6 Hz, 1H), 7.68 (t, J = 7.6 Hz, 1H), 7.59 (d, J = 8.2 Hz, 2H), 7.55 – 7.47 (m, 4H), 7.40 (t, J = 7.4 Hz, 2H), 7.33 (t, J = 7.4 Hz, 1H), 6.53 (brs, 1H), 5.78 (ddt, J = 17.0, 10.2, 6.7 Hz, 1H), 4.98 (d, J = 17.0 Hz, 1H), 4.93 (d, J = 10.2 Hz, 1H), 3.09 (d, J = 14.9 Hz, 1H), 2.03 (q, J = 7.5 Hz, 2H), 1.90 (d, J = 14.9 Hz, 1H), 1.34 – 1.27 (m, 6H), 1.02 (s, 3H), 0.91 (s, 3H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 192.3, 162.4, 141.2, 141.1, 140.1, 138.9, 134.9, 133.3, 130.9, 130.8, 128.8, 128.4, 127.61, 127.59, 127.3, 127.1, 125.9, 114.4, 70.7, 50.9,

44.7, 34.3, 33.7, 29.6, 29.3, 28.4, 23.5.

HR-MS (ESI-TOF) calcd for $C_{31}H_{33}NNaO_2^+$ $[M+Na]^+$ 474.2404; found 474.2406.

3-([1,1'-Biphenyl]-4-yl)-3-(5-(2,5-dimethylphenoxy)-2,2-dimethylpentyl)-2,3-dihydroisoquinoline-1,4-dione (3t)



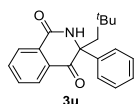
Following typical procedure B, the reaction of **1q** (73.7 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3t** as white solid (31.3 mg, 59% yield).

1H NMR (400 MHz, Chloroform-*d*) δ 8.32 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.05 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.76 (td, $J = 7.6, 1.4$ Hz, 1H), 7.70 – 7.61 (m, 3H), 7.56 – 7.48 (m, 4H), 7.44 – 7.38 (m, 2H), 7.37 – 7.30 (m, 1H), 7.24 (s, 1H), 7.00 (d, $J = 7.5$ Hz, 1H), 6.66 (t, $J = 7.5$ Hz, 1H), 6.58 (brs, 1H), 3.89 (t, $J = 6.4$ Hz, 2H), 3.17 (d, $J = 14.9$ Hz, 1H), 2.29 (s, 3H), 2.16 (s, 3H), 2.00 (d, $J = 14.9$ Hz, 1H), 1.90 – 1.81 (m, 2H), 1.56 – 1.47 (m, 2H), 1.12 (s, 3H), 0.99 (s, 3H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 192.5, 162.8, 157.1, 141.4, 141.2, 140.2, 136.5, 135.0, 133.4, 131.1, 130.9, 130.4, 128.9, 128.5, 127.7, 127.4, 127.2, 126.1, 123.7, 120.8, 112.1, 70.8, 68.4, 50.8, 41.3, 34.3, 28.3, 27.0, 24.5, 21.5, 15.9.

HR-MS (ESI-TOF) calcd for $C_{36}H_{37}NNaO_3^+$ $[M+Na]^+$ 554.2666; found 554.2669.

3-Neopentyl-3-phenyl-2,3-dihydroisoquinoline-1,4-dione (3u)



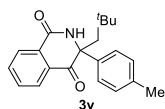
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2f** (14.5 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3u** as white solid (15.4 mg, 50% yield).

1H NMR (400 MHz, Chloroform-*d*) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.00 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.77 (td, $J = 7.6, 1.4$ Hz, 1H), 7.66 (td, $J = 7.6, 1.4$ Hz, 1H), 7.55 – 7.49 (m, 2H), 7.33 – 7.27 (m, 2H), 7.26 – 7.21 (m, 1H), 6.52 (brs, 1H), 3.09 (d, $J = 14.8$ Hz, 1H), 1.84 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (101 MHz, Chloroform-*d*) δ 192.4, 162.6, 142.3, 135.0, 133.4, 131.0, 130.9, 129.0, 128.5, 128.3, 127.4, 125.5, 71.0, 52.7, 31.9, 31.7.

HR-MS (ESI-TOF) calcd for $C_{20}H_{21}NNaO_2^+$ $[M+Na]^+$ 330.1465; found 330.1465.

3-Neopentyl-3-(*p*-tolyl)-2,3-dihydroisoquinoline-1,4-dione (3v)



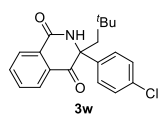
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2g** (15.9 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3v** as white solid (14.0 mg, 44% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.29 (dd, *J* = 7.8, 1.3 Hz, 1H), 8.00 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.76 (td, *J* = 7.6, 1.4 Hz, 1H), 7.66 (td, *J* = 7.6, 1.4 Hz, 1H), 7.43 – 7.34 (m, 2H), 7.13 – 7.08 (m, 2H), 6.50 (brs, 1H), 3.07 (d, *J* = 14.8 Hz, 1H), 2.27 (s, 3H), 1.81 (d, *J* = 14.8 Hz, 1H), 0.99 (s, 9H).

¹³C NMR (101 MHz, Chloroform-*d*) δ 192.5, 162.6, 139.4, 138.2, 134.9, 133.3, 131.02, 130.95, 129.7, 128.4, 127.4, 125.4, 70.8, 52.6, 31.9, 31.7, 21.0.

HR-MS (ESI-TOF) calcd for C₂₁H₂₃NNaO₂⁺ [M+Na]⁺ 344.1621; found 344.1622.

3-(4-Chlorophenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3w**)



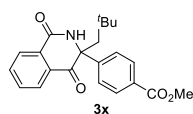
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2h** (17.9 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3w** as white solid (22.9 mg, 67% yield).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.30 (dd, *J* = 7.8, 1.3 Hz, 1H), 8.01 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.79 (td, *J* = 7.6, 1.4 Hz, 1H), 7.69 (td, *J* = 7.6, 1.4 Hz, 1H), 7.51 – 7.44 (m, 2H), 7.29 – 7.25 (m, 2H), 6.79 (s, 1H), 3.04 (d, *J* = 14.8 Hz, 1H), 1.84 (d, *J* = 14.8 Hz, 1H), 1.00 (s, 9H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 192.2, 162.5, 140.9, 135.2, 134.5, 133.5, 131.0, 130.8, 129.1, 128.6, 127.4, 127.1, 70.6, 52.7, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for C₂₀H₂₁ClNO₂⁺ [M+H]⁺ 342.1255; found 342.1256.

Methyl 4-(3-neopentyl-1,4-dioxo-1,2,3,4-tetrahydroisoquinolin-3-yl)benzoate (**3x**)



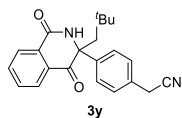
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2i** (20.3 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3x** as white solid (17.2 mg, 47% yield).

¹H NMR (600 MHz, Chloroform-*d*) δ 8.30 (d, *J* = 7.8 Hz, 1H), 8.00 (d, *J* = 7.8 Hz, 1H), 7.95 (d, *J* = 8.3 Hz, 2H), 7.79 (t, *J* = 7.6 Hz, 1H), 7.67 (t, *J* = 7.6 Hz, 1H), 7.61 (d, *J* = 8.3 Hz, 2H), 6.68 (brs, 1H), 3.87 (s, 3H), 3.07 (d, *J* = 14.8 Hz, 1H), 1.87 (d, *J* = 14.8 Hz, 1H), 1.00 (s, 9H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 192.0, 166.6, 162.5, 147.1, 135.2, 133.5, 131.0, 130.8, 130.2, 130.1, 128.6, 127.4, 125.7, 71.1, 52.6, 52.4, 32.0, 31.7.

HR-MS (ESI-TOF) calcd for C₂₂H₂₄NO₄⁺ [M+H]⁺ 366.1700; found 366.1690.

2-(4-(3-Neopentyl-1,4-dioxo-1,2,3,4-tetrahydroisoquinolin-3-yl)phenyl)acetonitrile (**3y**)



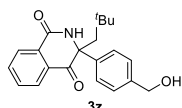
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2j** (18.4 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3y** as white solid (18.7 mg, 54% yield).

^1H NMR (400 MHz, Chloroform-*d*) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.00 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.79 (td, $J = 7.6, 1.4$ Hz, 1H), 7.68 (td, $J = 7.6, 1.4$ Hz, 1H), 7.57 – 7.51 (m, 2H), 7.29 – 7.25 (m, 2H), 6.76 (brs, 1H), 3.68 (s, 2H), 3.06 (d, $J = 14.8$ Hz, 1H), 1.84 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 192.3, 162.5, 142.5, 135.2, 133.5, 131.0, 130.8, 130.1, 128.6, 128.6, 127.4, 126.5, 117.5, 70.8, 52.7, 32.0, 31.7, 23.2.

HR-MS (ESI-TOF) calcd for $\text{C}_{22}\text{H}_{22}\text{N}_2\text{NaO}_2^+ [\text{M}+\text{Na}]^+$ 369.1573; found 369.1574.

3-(4-(Hydroxymethyl)phenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3z**)



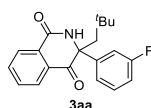
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2k** (17.5 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3z** as white solid (17.8 mg, 53% yield).

^1H NMR (400 MHz, Chloroform-*d*) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.00 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.78 (td, $J = 7.6, 1.4$ Hz, 1H), 7.67 (td, $J = 7.6, 1.4$ Hz, 1H), 7.53 – 7.48 (m, 2H), 7.32 – 7.28 (m, 2H), 6.54 (s, 1H), 4.63 (s, 2H), 3.08 (d, $J = 14.8$ Hz, 1H), 1.83 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (151 MHz, Chloroform-*d*) δ 192.4, 162.8, 141.5, 141.2, 135.0, 133.4, 131.0, 130.9, 128.5, 127.5, 127.3, 125.8, 70.8, 64.5, 52.7, 31.9, 31.7.

HR-MS (ESI-TOF) calcd for $\text{C}_{21}\text{H}_{24}\text{NO}_3^+ [\text{M}+\text{H}]^+$ 338.1751; found 338.1744.

3-(3-Fluorophenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3aa**)



Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2l** (16.3 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3aa** as white solid (18.5 mg, 57% yield).

^1H NMR (400 MHz, Chloroform-*d*) δ 8.30 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.02 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.79 (td, $J = 7.6, 1.4$ Hz, 1H), 7.68 (td, $J = 7.6, 1.4$ Hz, 1H), 7.33 – 7.28

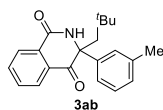
(m, 1H), 7.28 – 7.26 (m, 1H), 7.26 – 7.22 (m, 1H), 6.93 (tt, $J = 8.3, 1.3$ Hz, 1H), 6.80 (brs, 1H), 3.03 (d, $J = 14.8$ Hz, 1H), 1.86 (d, $J = 14.8$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 192.1, 163.1 (d, $J = 247.2$ Hz), 162.4, 145.0 (d, $J = 6.5$ Hz), 135.2, 133.5, 131.0, 130.8, 130.4 (d, $J = 8.2$ Hz), 128.6, 127.4, 121.1 (d, $J = 2.8$ Hz), 115.3 (d, $J = 21.2$ Hz), 113.2 (d, $J = 23.5$ Hz), 70.7 (d, $J = 1.6$ Hz), 52.8, 32.0, 31.7.

^{19}F NMR (376 MHz, Chloroform- d) δ -111.11 – -111.25 (m, 1F).

HR-MS (ESI-TOF) calcd for $\text{C}_{20}\text{H}_{21}\text{FNO}_2^+ [\text{M}+\text{H}]^+$ 326.1551; found 326.1551.

3-Neopentyl-3-(*m*-tolyl)-2,3-dihydroisoquinoline-1,4-dione (**3ab**)



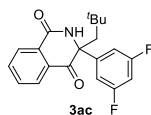
Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2m** (15.9 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3ab** as white solid (13.2 mg, 41% yield).

^1H NMR (600 MHz, Chloroform- d) δ 8.30 (d, $J = 7.7$ Hz, 1H), 8.01 (d, $J = 7.7$ Hz, 1H), 7.77 (t, $J = 7.6$ Hz, 1H), 7.66 (t, $J = 7.6$ Hz, 1H), 7.33 – 7.29 (m, 2H), 7.18 (t, $J = 8.0$ Hz, 1H), 7.05 (d, $J = 7.5$ Hz, 1H), 6.65 (brs, 1H), 3.06 (d, $J = 14.9$ Hz, 1H), 2.30 (s, 3H), 1.78 (d, $J = 14.9$ Hz, 1H), 0.97 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 192.5, 162.6, 142.3, 138.8, 134.9, 133.3, 131.1, 131.0, 129.0, 128.9, 128.5, 127.4, 126.1, 122.6, 71.0, 52.8, 31.9, 31.7, 21.8.

HR-MS (ESI-TOF) calcd for $\text{C}_{21}\text{H}_{24}\text{NO}_2^+ [\text{M}+\text{H}]^+$ 322.1802; found 322.1802.

3-(3,5-Difluorophenyl)-3-neopentyl-2,3-dihydroisoquinoline-1,4-dione (**3ac**)



Following typical procedure B, the reaction of **1m** (47.0 mg, 0.18 mmol), **2n** (18.1 mg, 0.10 mmol), **4a** (40.8 mg, 0.25 mmol) for 12 h afforded **3ac** as white solid (22.3 mg, 65% yield).

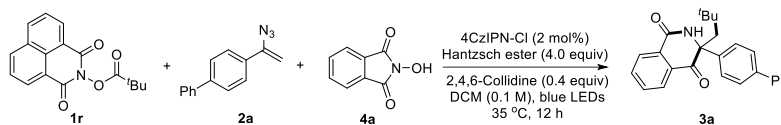
^1H NMR (600 MHz, Chloroform- d) δ 8.31 (dd, $J = 7.8, 1.3$ Hz, 1H), 8.03 (dd, $J = 7.8, 1.3$ Hz, 1H), 7.81 (td, $J = 7.6, 1.3$ Hz, 1H), 7.71 (td, $J = 7.6, 1.3$ Hz, 1H), 7.10 – 7.05 (m, 2H), 6.69 (tt, $J = 8.4, 2.3$ Hz, 1H), 6.63 (brs, 1H), 2.98 (d, $J = 14.9$ Hz, 1H), 1.83 (d, $J = 14.9$ Hz, 1H), 0.99 (s, 9H).

^{13}C NMR (151 MHz, Chloroform- d) δ 191.8, 163.24 (d, $J = 249.6$ Hz), 163.15 (d, $J = 249.6$ Hz), 162.5, 146.7 (t, $J = 7.6$ Hz), 135.3, 133.5, 131.0, 130.8, 128.6, 127.4, 109.2 (d, $J = 6.0$ Hz), 109.1 (d, $J = 6.0$ Hz), 103.7 (t, $J = 25.6$ Hz), 70.5 (t, $J = 2.2$ Hz), 52.8 (d, $J = 13.7$ Hz), 32.0, 31.6.

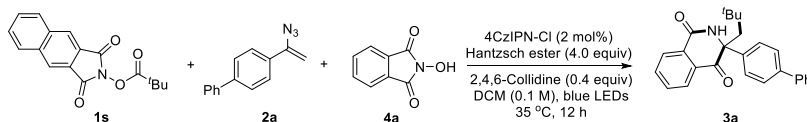
^{19}F NMR (376 MHz, Chloroform- d) δ -107.68 – -107.79 (m, 2F).

HR-MS (ESI-TOF) calcd for $C_{20}H_{20}F_2NO_2^+$ $[M+H]^+$ 344.1457; found 344.1458.

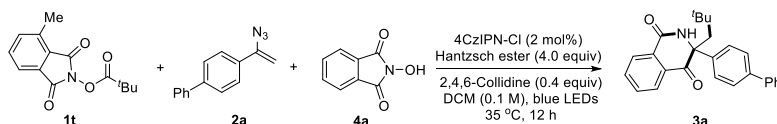
Convergent synthesis of **3a** from different NHPIs



To a Schlenk tube were added **1r** (53.5 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4a** (32.6 mg, 0.20 mmol, 2.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μ L, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** in 51% (19.5 mg) yield.

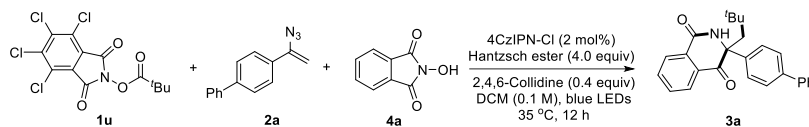


To a Schlenk tube were added **1s** (53.5 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μ L, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** in 69% (26.4 mg) yield.

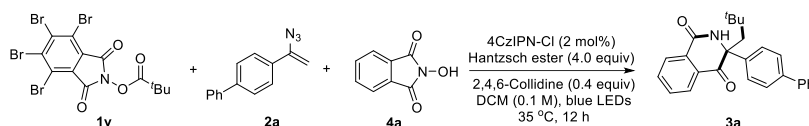


To a Schlenk tube were added **1t** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μ L, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed

to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** in 66% (25.3 mg) yield.

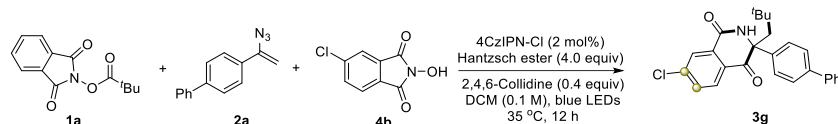


To a Schlenk tube were added **1u** (45.9 mg, 0.12 mmol, 1.2 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μL, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** in 52% (19.9 mg) yield.

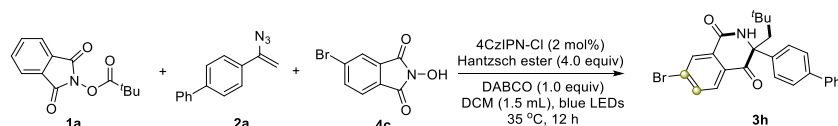


To a Schlenk tube were added **1v** (89.4 mg, 0.16 mmol, 1.6 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μL, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** in 45% (17.3 mg) yield.

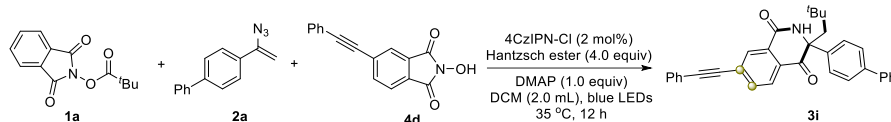
Divergent synthesis of **3** from **1a**



To a Schlenk tube were added **1a** (29.6 mg, 0.12 mmol, 1.2 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4b** (23.6 mg, 0.12 mmol, 1.2 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) 2,4,6-collidine (4.8 μ L, 0.04 mmol, 40 mol%). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3g** in 45% (18.8 mg, rr = 1:1) yield.

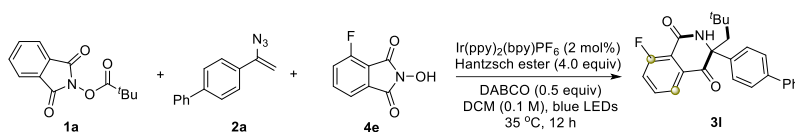


To a Schlenk tube were added **1a** (44.5 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4c** (43.2 mg, 0.18 mmol, 1.8 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv), DABCO (11.2 mg, 0.1 mmol, 1.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.5 mL). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3h** in 46% (21.2 mg, rr = 1:1) yield.



To a Schlenk tube were added **1a** (44.5 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4d** (47.3 mg, 0.18 mmol, 1.8 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv), DMAP (12.1 mg, 0.1 mmol, 1.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (2.0 mL). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL).

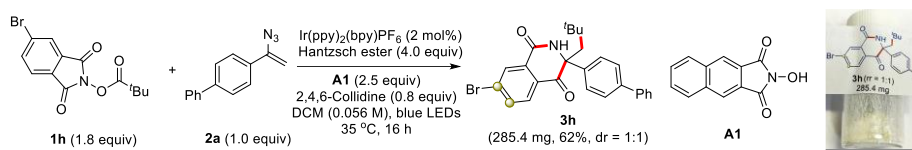
The organic phases were dried over anhydrous sodium sulfate, filtered, and evacuated under vacuum. The residue was purified directly by column chromatography to afford the pure product **3i** in 48% (23.2 mg, rr = 1:1) yield.



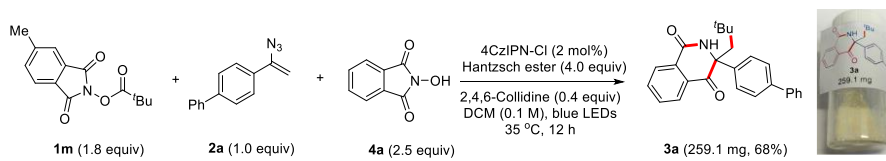
To a Schlenk tube were added **1a** (44.5 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.10 mmol, 1.0 equiv), **4e** (32.6 mg, 0.18 mmol, 1.8 equiv), Ir(ppy)₂(bpy)PF₆ (1.6 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv), DABCO (5.6 mg, 0.05 mmol, 50 mol%). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3i** in 48% (19.2 mg, rr = 1:1) yield.

VI. Mechanistic Study

Scale-up reaction

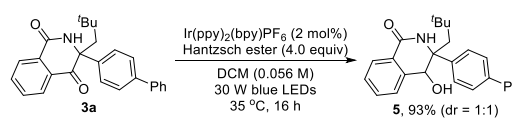


To a Schlenk tube were added **1h** (0.59 g, 1.8 mmol, 1.8 equiv), **2a** (0.22 g, 1.0 mmol, 1.0 equiv), **A1** (0.44 g, 2.5 mmol, 2.5 equiv), $\text{Ir(ppy)}_2(\text{bpy})\text{PF}_6$ (16.0 mg, 0.02 mmol, 2.0 mol%), Hantzsch ester (1.01 g, 4.0 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (18.0 mL) and 2,4,6-collidine (96.0 μL , 0.8 mmol, 0.8 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H_2O (20.0 mL) and extracted with DCM (3 \times 10.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3h** (285.4 mg, 62%, dr = 1:1).



To a Schlenk tube were added **1m** (0.47 g, 1.8 mmol, 1.8 equiv), **2a** (0.22 g, 1.0 mmol, 1.0 equiv), **4a** (0.41 g, 2.5 mmol, 2.5 equiv), 4CzIPN-Cl (21.0 mg, 0.02 mmol, 2.0 mol%), Hantzsch ester (1.01 g, 4.0 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (10.0 mL) and 2,4,6-collidine (48.0 μL , 0.4 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with stirring 12 h. The reaction was diluted with H_2O (20.0 mL) and extracted with DCM (3 \times 10.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **3a** (259.1 mg, 68%).

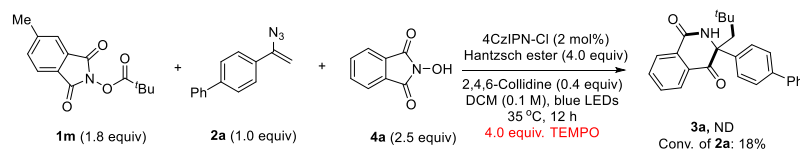
Selective reduction of **3a**



To a Schlenk tube were added **3a** (38.3 mg, 0.10 mmol, 1.0 equiv), $\text{Ir(ppy)}_2(\text{bpy})\text{PF}_6$

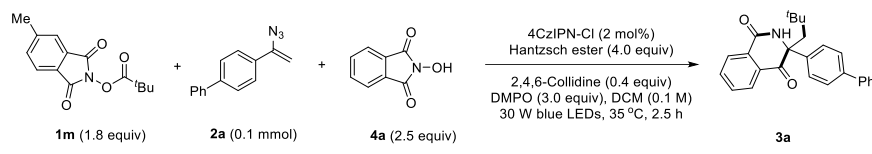
(1.6 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.40 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.8 mL). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C with 16 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **5** (35.8 mg, 93% yield). ¹H NMR (600 MHz, Chloroform-*d*) δ 8.14 (d, *J* = 7.6 Hz, 1H), 7.64 (d, *J* = 8.1 Hz, 2H), 7.62 – 7.58 (m, 4H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.49 – 7.41 (m, 4H), 7.35 (t, *J* = 7.4 Hz, 1H), 6.48 (brs, 1H), 4.77 (s, 1H), 2.23 (d, *J* = 14.9 Hz, 1H), 2.19 (brs, 1H), 1.85 (d, *J* = 14.9 Hz, 1H), 0.78 (s, 9H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 165.1, 140.6, 140.1, 139.0, 138.9, 133.2, 129.05, 128.96, 128.3, 127.9, 127.7, 127.5, 127.1, 127.0, 126.7, 75.0, 65.5, 49.1, 32.0, 31.8. HR-MS (ESI-TOF) calcd for C₂₆H₂₈NO₂⁺ [*M*+*H*]⁺ 386.2115 found 386.2102.

Radical intermediate quench reactions



To a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), TEMPO (62.4 mg, 0.40 mmol, 4.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μL, 0.04 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The conversion of **2a** (18%) and yield of **3a** (0%) were determined by ¹H NMR of the crude reaction mixture using mesitylene as internal standard.

EPR Experiment



To a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), DMPO (33.9 mg, 0.30 mmol,

3.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μ L, 0.04 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 $^{\circ}$ C for 2.5 h, then detected the EPR signal.

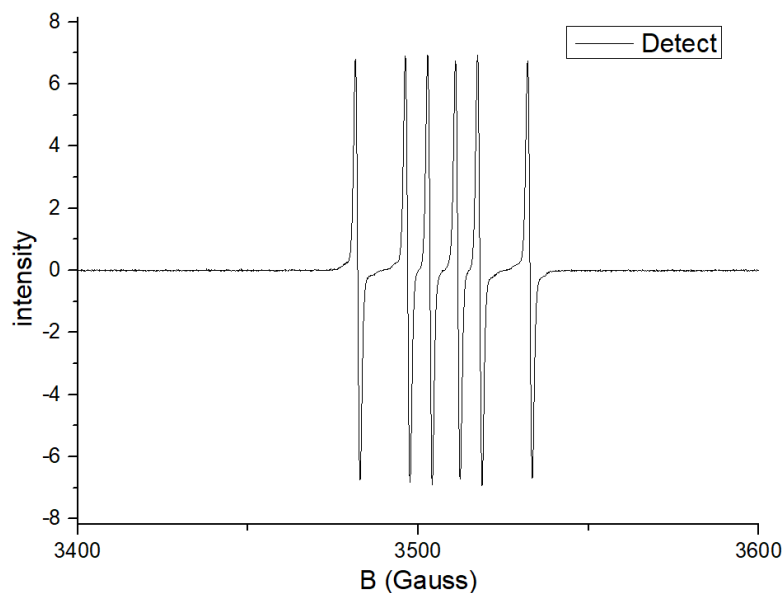
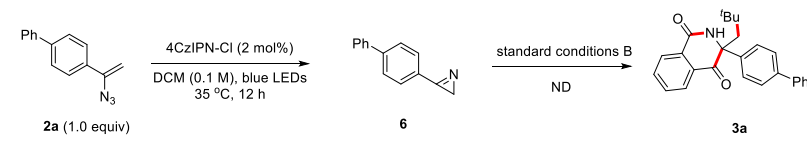


Figure S1. EPR spectra of the reaction mixture.

Excluding 2*H*-azirine as an intermediate

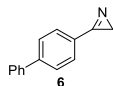


To a Schlenk tube were added **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 $^{\circ}$ C for 12 h. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 \times 5.0 mL). The combined organic phases were dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was purified directly by column chromatography to afford the pure product **6**.

Then, to a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **6** (19.3 mg, 0.1 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μ L, 0.04 mmol, 0.4 equiv). Then the tube was sealed

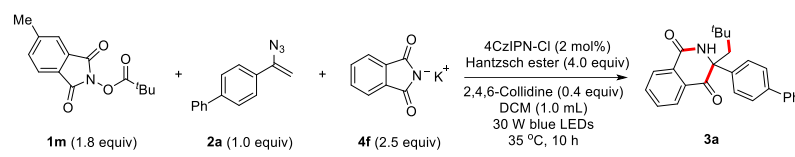
and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 12 h. The crude reaction mixture was analyzed by ¹H NMR spectroscopy and desired product **3a** was not detected.

3-([1,1'-biphenyl]-4-yl)-2*H*-azirine (**6**)¹



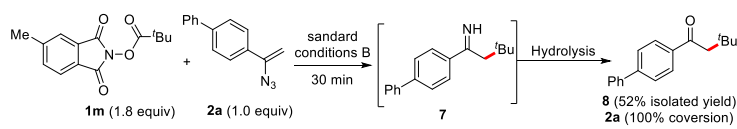
¹H NMR (600 MHz, Chloroform-*d*) δ 8.00 – 7.97 (m, 2H), 7.81 – 7.78 (m, 2H), 7.67 – 7.65 (m, 2H), 7.51 – 7.47 (m, 2H), 7.44 – 7.40 (m, 1H), 1.82 (s, 2H).

Excluding phthalimide anion as intermediate for the reaction



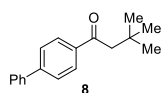
To a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), **4f** (46.3 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μL, 0.04 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 10 h. The crude reaction mixture was analyzed by ¹H NMR spectroscopy and the desired product **3a** was not detected.

Identifying ketimine as an intermediate

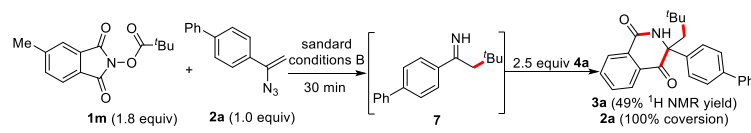


To a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μL, 0.04 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 30 minutes. The reaction was diluted with H₂O (5.0 mL) and extracted with DCM (3 × 5.0 mL). The organic phases were dried over Na₂SO₄ and concentrated under reduced pressure after filtration. The residue was purified directly by column chromatography to afford the pure product **8** (13.0 mg) in 52% yield.

1-([1,1'-Biphenyl]-4-yl)-3,3-dimethylbutan-1-one (**8**)¹

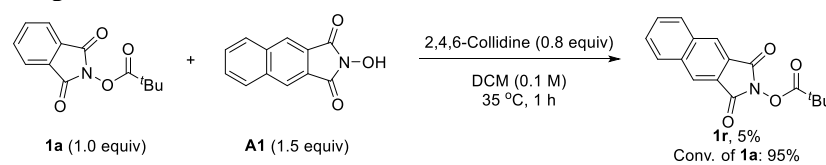


¹H NMR (400 MHz, Chloroform-*d*) δ 8.04 – 7.99 (m, 2H), 7.70 – 7.61 (m, 4H), 7.50 – 7.44 (m, 2H), 7.42 – 7.37 (m, 1H), 2.89 (s, 2H), 1.09 (s, 9H).



To a Schlenk tube were added **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv). The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (4.8 μ L, 0.04 mmol, 0.4 equiv). Then the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 30 minutes. Then **4a** (40.7 mg, 0.25 mmol, 2.5 equiv) was added under N₂, and the tube was sealed and exposed to blue LEDs (30 W LED light bulbs 2.5 cm away from the vial) at 35 °C for 12 h. The reaction mixture was diluted with ethyl acetate and poured into a separatory funnel, washed with brine. The combined organic layers were dried over Na₂SO₄ and concentrated under reduced pressure after filtration. The conversion of **2a** (100%) and yield of **3a** (49%) were determined by ¹H NMR of the crude reaction mixture using mesitylene as internal standard.

Cross-over experiment



To a Schlenk tube were added **1a** (24.7 mg, 0.10 mmol, 1.0 equiv), **A1** (32.0 mg, 0.15 mmol, 1.5 equiv), The tube was evacuated and back-filled with nitrogen (3 cycles), followed by the addition of DCM (1.0 mL) and 2,4,6-collidine (9.6 μ L, 0.08 mmol, 0.8 equiv). Then the tube was sealed and stirred at 35 °C for 1 h. The reaction mixture was concentrated under reduced pressure. The yield of **1r** (5%) and conv. of **1a** (95%) was determined by ¹H NMR of the crude reaction mixture using mesitylene as internal standard.

Quantum yield measurement

Blue LED (λ_{max} = 440 nm) was used for measurement of quantum yield.

Determination of the light intensity at 440 nm

The photon flux of the blue LED light was determined by standard ferrioxalate actinometry,¹³⁻¹⁵ the photon flux of the LED ($\lambda_{\text{max}} = 440$ nm) was determined by standard ferrioxalate actinometry. A 0.15 M solution of ferrioxalate was prepared by dissolving potassium ferrioxalate hydrate (2.21 g) in H₂SO₄ (30 mL of a 0.05 M solution). A buffered solution of 1,10-phenanthroline was prepared by dissolving 1,10-phenanthroline (50 mg) and sodium acetate (11.25g) in H₂SO₄ (50 mL of a 0.5 M solution). Both solutions were stored in the dark. To determine the photon flux of the LED, the ferrioxalate solution (2.0 mL) was placed in a cuvette and irradiated for 45 seconds at $\lambda_{\text{max}} = 440$ nm. After irradiation, the phenanthroline solution (0.35 mL) was added to the cuvette and the mixture was allowed to stand in the dark for 1 h to allow the ferrous ions to completely coordinate to the phenanthroline. The absorbance of the solution was measured at 510 nm. A nonirradiated sample was also prepared and the absorbance at 510 nm was measured. Conversion was calculated using eq 1.

$$\text{Mol of Fe}^{2+} = \frac{V \cdot \Delta A_{510\text{nm}}}{l \cdot \epsilon} = \frac{(0.00235 \text{ L}) \cdot (1.655)}{(1.0 \text{ cm}) \cdot 11100 \frac{\text{L}}{\text{mol} \cdot \text{cm}}} = 3.50 \times 10^{-7} \text{ mol} \quad (1)$$

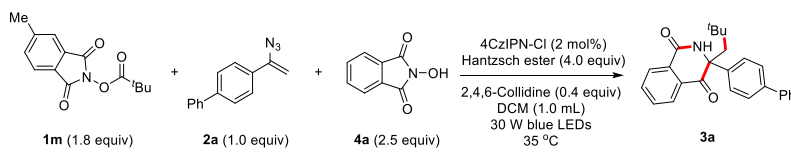
V is the total volume (0.00235 L) of the solution after addition of phenanthroline, ΔA (1.425) is the difference in absorbance at 510 nm between the irradiated and non-irradiated solutions (**Figure S2**), l is the path length (1.00 cm), and ϵ is the molar absorptivity of the ferrioxalate actinometer at 510 nm (11,100 Lmol⁻¹ cm⁻¹). The photon flux can be calculated using eq 2.

$$\text{Photon flux} = \frac{\text{mol of Fe}^{2+}}{\Phi \cdot t \cdot f} = \frac{3.50 \times 10^{-7}}{(1.01) \cdot (45 \text{ s}) \cdot (0.996)} = 7.73 \times 10^{-9} \text{ einstein/s} \quad (2)$$

Where Φ is the quantum yield for the ferrioxalate actinometer (1.01 at $\lambda = 440$ nm), t is the irradiation time (45 s), and f is the fraction of light absorbed at 440 nm by the ferrioxalate actinometer. This value is calculated using eq 3 where $A_{440 \text{ nm}}$ is the absorbance of the ferrioxalate solution at 440 nm.

$$f = 1 - 10^{-A_{440\text{nm}}} = 1 - 10^{-2.354} = 0.996 \quad (3)$$

The photon flux was thus calculated to be 7.73×10^{-9} einstein/s



A cuvette sealed with a rubber stopper was charged with **1m** (47.0 mg, 0.18 mmol, 1.8 equiv), **2a** (22.1 mg, 0.1 mmol, 1.0 equiv), **4a** (40.7 mg, 0.25 mmol, 2.5 equiv), 4CzIPN-Cl (2.1 mg, 0.002 mmol, 2.0 mol%), Hantzsch ester (101.3 mg, 0.4 mmol, 4.0 equiv), 2,4,6-collidine (4.8 μ L, 0.04 mmol, 0.4 equiv) in 1 mL DCM under argon atmosphere. The sample was stirred and irradiated ($\lambda = 440$ nm) for 1800 s (0.5 h) at room temperature. After irradiation, the solvent was removed. The yield of product

formed was determined as 23% yield (2.3×10^{-5} mol of **3a**) by crude ^1H NMR based on a mesitylene standard. The reaction quantum yield (Φ) was determined using eq 5 where the photon flux is 7.73×10^{-9} einstein/s (determined by actinometry as described above), t is the reaction time (1800 s) and f is the fraction of incident light absorbed by the photosensitizer, 4CzIPN-Cl, determined using eq 4. An absorption spectrum of the catalyst (1.25×10^{-4} M) gave an absorbance value of 0.752 at 440 nm (**Figure S3**), indicating that the fraction of light absorbed by the photocatalyst (f) is 0.823.

$$f = 1 - 10^{-A_{440\text{nm}}} = 1 - 10^{-0.752} = 0.823 \quad (4)$$

$$\Phi = \frac{\text{mol of product}}{\text{flux} \times t \times f} = \frac{2.30 \times 10^{-5} \text{ mol}}{7.73 \times 10^{-9} \text{ einstein/s} \times 1800 \text{ s} \times 0.823} = 2.01 \quad (5)$$

The reaction quantum yield (Φ) was calculated to be 2.01.

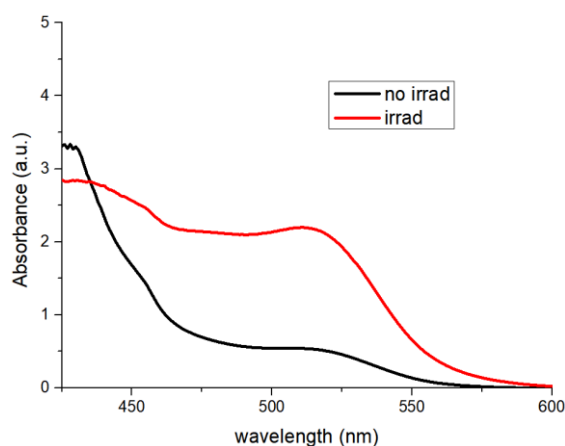


Figure S2. The irradiation experiment and non-irradiation experiment absorption spectra.

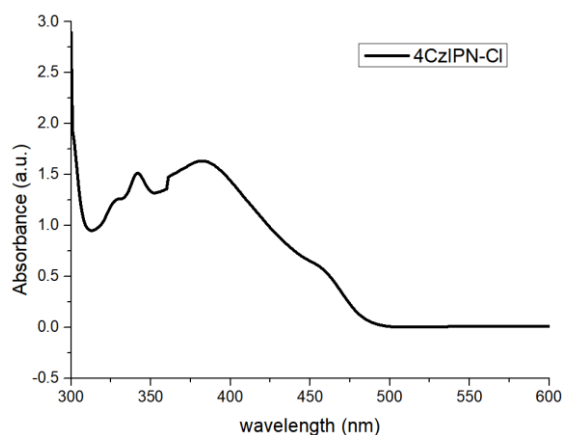
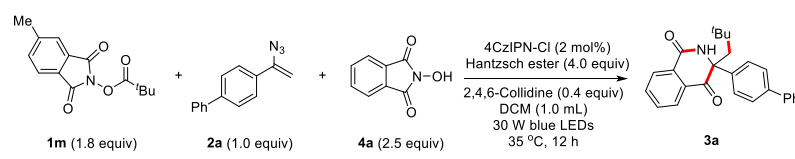


Figure S3. Absorption spectra of 4CzIPN-Cl.

The luminescence quenching experiments



Emission intensities were recorded using FS-5 fluorescence spectrometer (Edinburgh instrument) for all experiments. All 4CzIPN-Cl solutions were excited at 454 nm and the emission intensity was collected at 425-750 nm. In a typical experiment, the DCM as solution of 4CzIPN-Cl (0.5 μ M) was added the appropriate amount of quencher in a 4.5 cm quartz cuvette. After degassing with nitrogen for 10 min, the emission spectra of the samples were collected. The results showed that Hantzsch could quench the photoexcited 4CzIPN-Cl effectively, while **1m**, **2a**, **4a** and 2,4,6-collidine were less effective. The emission intensity at 542 nm was observed.

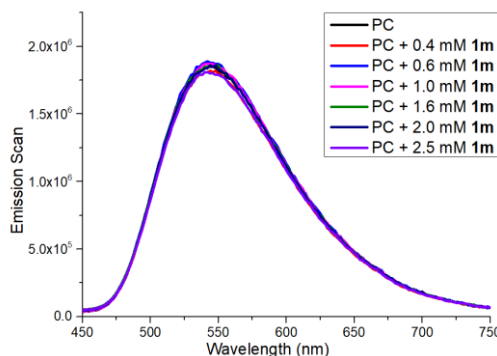


Figure S4. The luminescence quenching of **1m**.

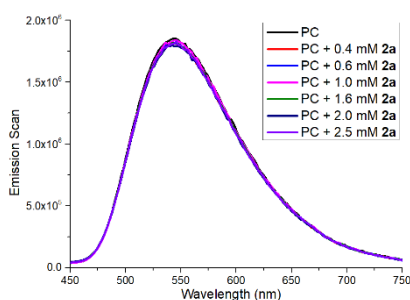


Figure S5. The luminescence quenching of **2a**.

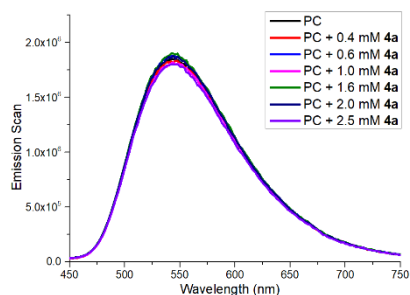


Figure S6. The luminescence quenching of **4a**.

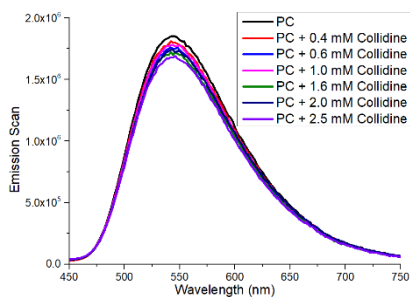


Figure S7. The luminescence quenching of 2,4,6-collidine.

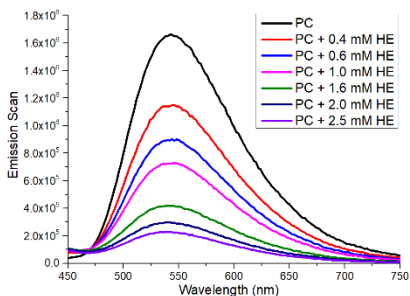


Figure S8. The luminescence quenching of Hantzsch ester.

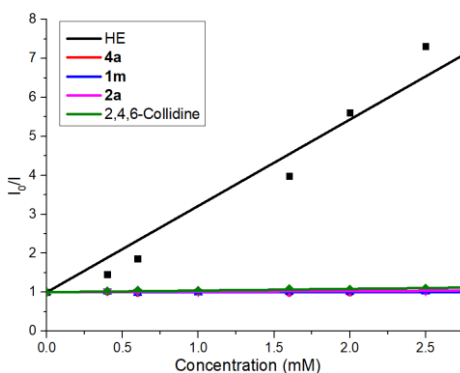


Figure S9. The lines of luminescence quenching experiments.

The Cyclic Voltammetry Experiments

Cyclic voltammetry was carried out in a gas-tight glass cell with CHI 660Epotentiostat in a nitrogen atmosphere. A glassy carbon disk electrode (diameter is 1.0 mm.PTFE shroud) was used as a working electrode. A platinum foil was used as a counter electrode. Ag/AgNO₃ electrode was used as a reference electrode, which was calibrated with Fc/Fc* redoxcouple, and a scan rate of 1 V/s. Samples were prepared with 0.05 mmol of substrate in 10.0 mL of 0.1 M tetrabutylammonium hexafluorophosphate in dry, degassed acetonitrile.

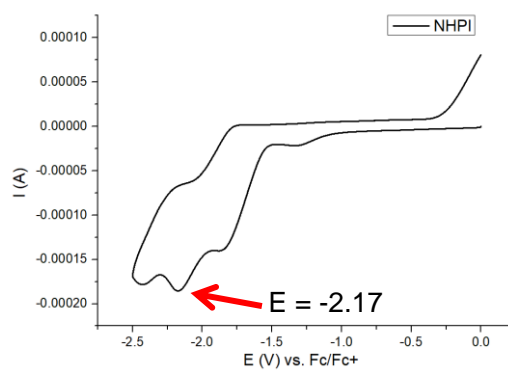


Figure S10. Cyclic voltammogram of NHPI (4a).

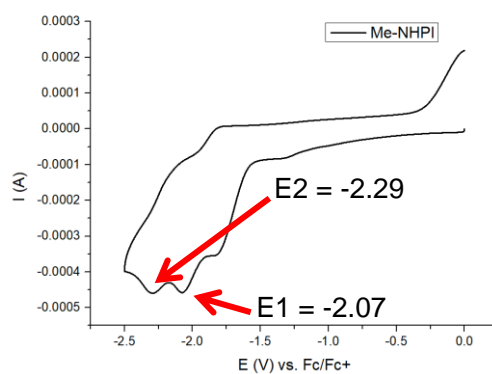
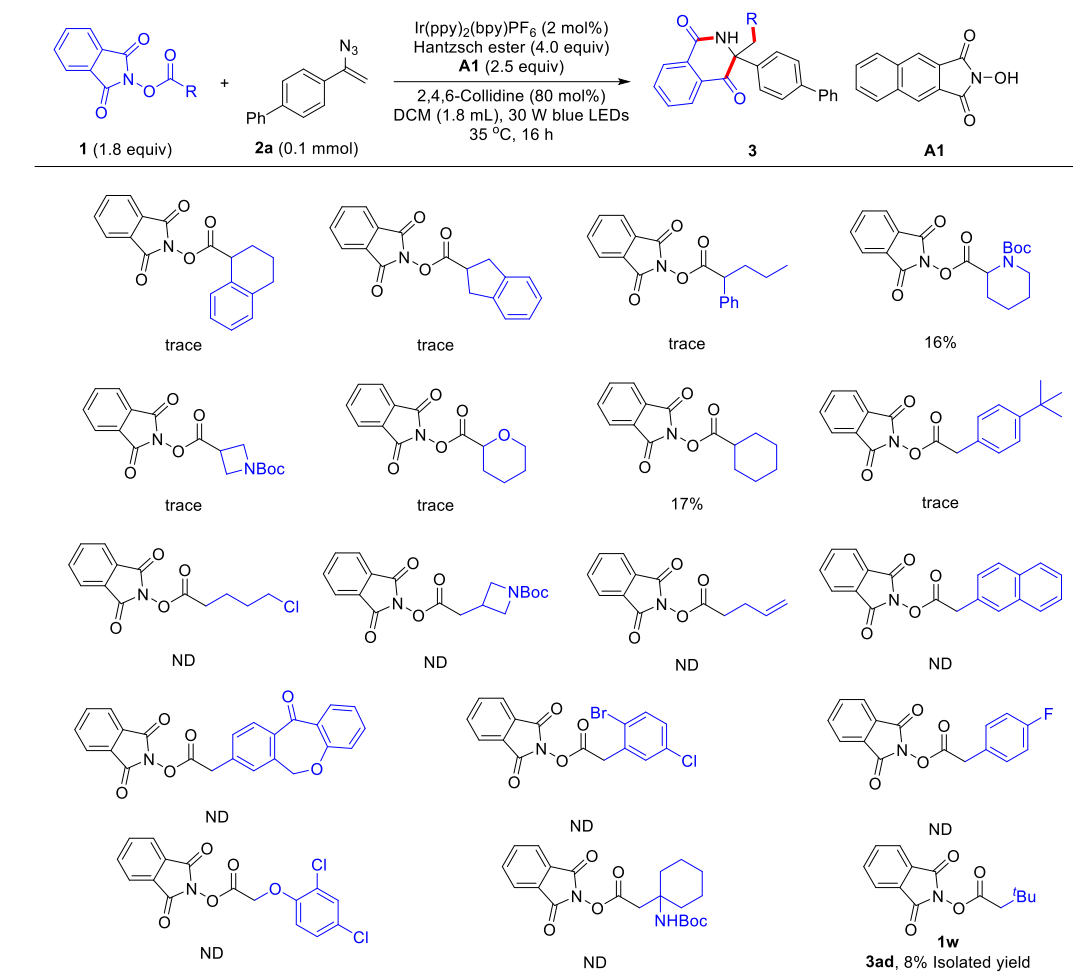
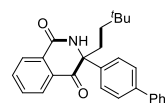


Figure S11. Cyclic voltammogram of Me-NHPI.



Yield was determined by ^1H NMR of the crude mixture using mesitylene internal standard.

**3-([1,1'-biphenyl]-4-yl)-3-(3,3-dimethylbutyl)-2,3-dihydroisoquinoline-1,4-dione
(3ad)**

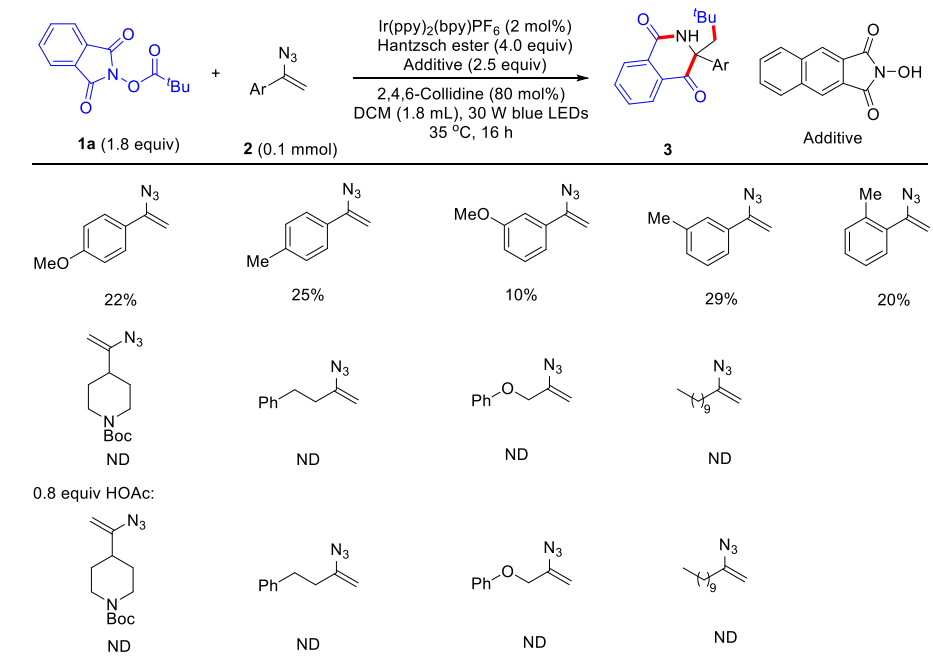


Following typical procedure A, the reaction of **1w** (47.0 mg, 0.18 mmol), **2a** (22.1 mg, 0.10 mmol) for 16 h afforded **3ad** as white solid (3.0 mg, 8% yield).

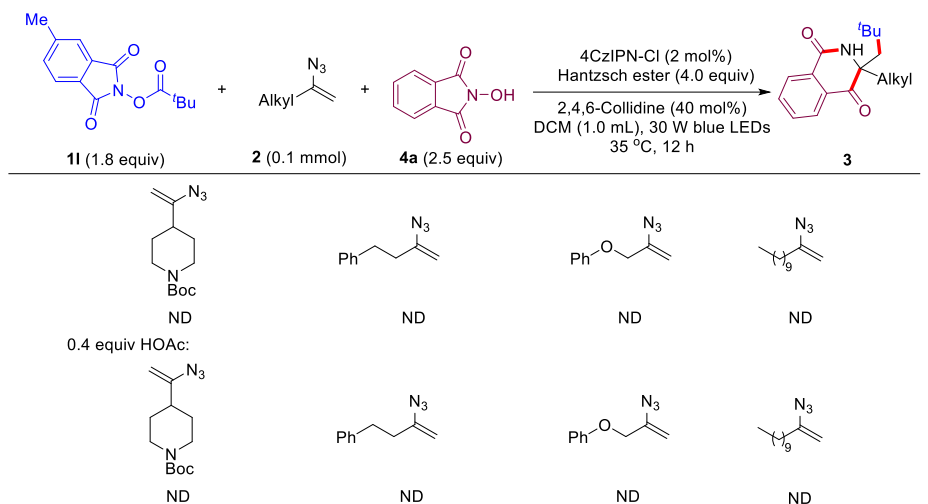
¹H NMR (600 MHz, Chloroform-*d*) δ 8.29 (d, *J* = 7.8 Hz, 1H), 8.00 (d, *J* = 7.5 Hz, 1H), 7.78 (t, *J* = 7.5 Hz, 1H), 7.68 (t, *J* = 7.3 Hz, 1H), 7.60 – 7.50 (m, 6H), 7.41 (t, *J* = 7.2 Hz, 2H), 7.36 – 7.31 (m, 1H), 6.54 (brs, 1H), 2.63 (tt, *J* = 13.3, 3.2 Hz, 1H), 2.07 (tt, *J* = 13.3, 3.2 Hz, 1H), 1.34 – 1.27 (m, 2H), 0.91 (s, 9H).

¹³C NMR (151 MHz, Chloroform-*d*) δ 193.1, 163.4, 141.3, 140.2, 139.4, 135.1, 133.4, 131.3, 131.2, 129.0, 128.5, 127.8, 127.20, 127.18, 126.3, 70.8, 37.8, 35.8, 30.4, 29.4.

HR-MS (ESI-TOF) calcd for $\text{C}_{27}\text{H}_{28}\text{NO}_2^+$ $[\text{M}+\text{H}]^+$ 398.2115; found 398.2113.



Yield was determined by ^1H NMR of the crude mixture using mesitylene internal standard.



Yield was determined by ^1H NMR of the crude mixture using mesitylene internal standard.

VIII. X-Ray Diffraction Data

X-Ray Diffraction Data of 3a (CCDC 2361327)

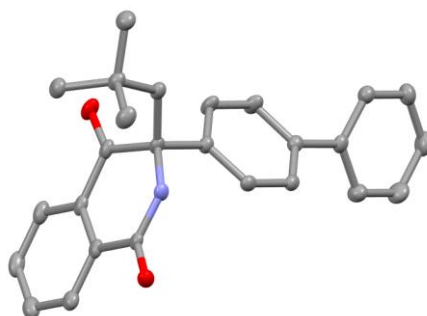


Table S13. Crystal data and structure refinement for **3a** (dhw512602_0m)

| | |
|--|--|
| Identification code | dhw512602_0m |
| Empirical formula | C ₂₆ H ₂₅ NO ₂ |
| Formula weight | 383.47 |
| Temperature/K | 100.0(2) |
| Crystal system | triclinic |
| Space group | P-1 |
| a/Å | 13.8148(7) |
| b/Å | 17.2854(8) |
| c/Å | 19.6680(9) |
| α /° | 79.694(2) |
| β /° | 74.868(2) |
| γ /° | 68.162(2) |
| Volume/Å ³ | 4190.8(4) |
| Z | 8 |
| $\rho_{\text{calc}}/\text{cm}^3$ | 1.216 |
| μ/mm^{-1} | 0.598 |
| F(000) | 1632.0 |
| Crystal size/mm ³ | 0.03 × 0.02 × 0.02 |
| Radiation | CuK α (λ = 1.54178) |
| 2 Θ range for data collection/° | 4.674 to 144.762 |
| Index ranges | -17 ≤ h ≤ 17, -21 ≤ k ≤ 21, -24 ≤ l ≤ 22 |
| Reflections collected | 79252 |
| Independent reflections | 16346 [R _{int} = 0.0603, R _{sigma} = 0.0467] |
| Data/restraints/parameters | 16346/0/1057 |

| | |
|--|----------------------------------|
| Goodness-of-fit on F^2 | 1.047 |
| Final R indexes [$I \geq 2\sigma(I)$] | $R_1 = 0.0581$, $wR_2 = 0.1555$ |
| Final R indexes [all data] | $R_1 = 0.0764$, $wR_2 = 0.1726$ |
| Largest diff. peak/hole / $e \text{ \AA}^{-3}$ | 1.17/-0.50 |

X-Ray Diffraction Data of **3h** (CCDC 2361326)

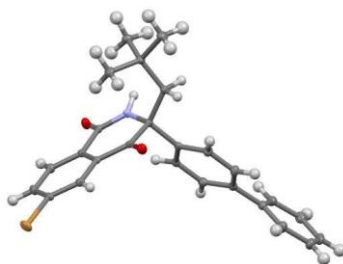


Table S14. Crystal data and structure refinement for **3h** (DHW202403215Q_0m)

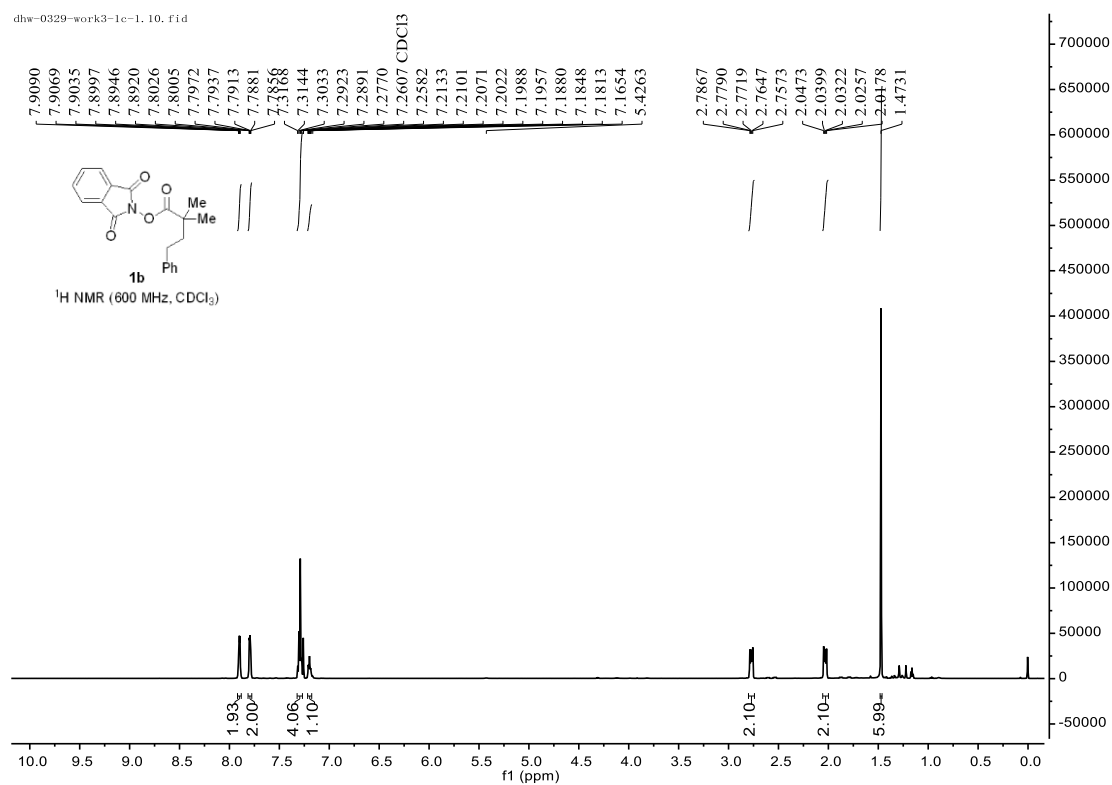
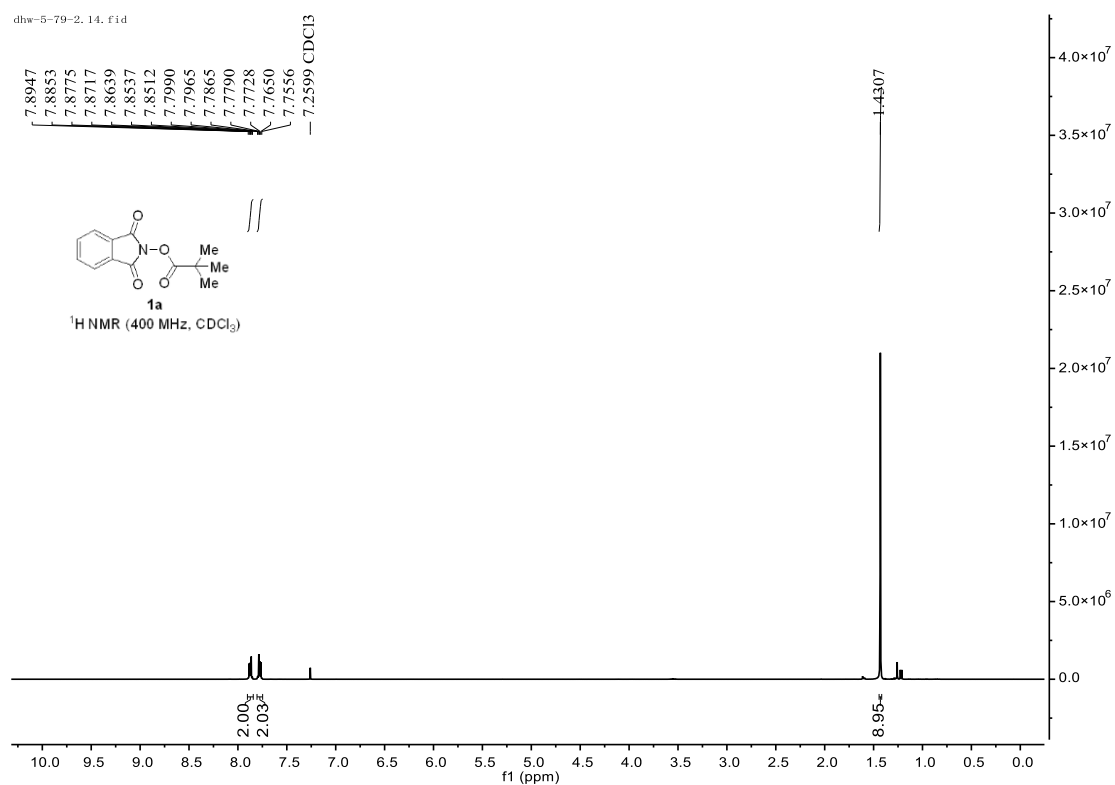
| | |
|---|--|
| Identification code | DHW202403215Q_0m |
| Empirical formula | $C_{26}H_{24}BrNO_2$ |
| Formula weight | 462.37 |
| Temperature/K | 100.0 |
| Crystal system | monoclinic |
| Space group | $P2_1/c$ |
| $a/\text{\AA}$ | 12.1999(2) |
| $b/\text{\AA}$ | 8.1197(2) |
| $c/\text{\AA}$ | 22.7110(5) |
| $\alpha/^\circ$ | 90 |
| $\beta/^\circ$ | 103.9270(10) |
| $\gamma/^\circ$ | 90 |
| Volume/ \AA^3 | 2183.61(8) |
| Z | 4 |
| $\rho_{\text{calc}}/\text{cm}^3$ | 1.406 |
| μ/mm^{-1} | 2.736 |
| $F(000)$ | 952.0 |
| Crystal size/ mm^3 | $0.03 \times 0.02 \times 0.02$ |
| Radiation | $\text{CuK}\alpha$ ($\lambda = 1.54178$) |
| 2θ range for data collection/ $^\circ$ | 7.466 to 145.394 |
| Index ranges | $-15 \leq h \leq 15$, $-8 \leq k \leq 9$, $-27 \leq l \leq 28$ |

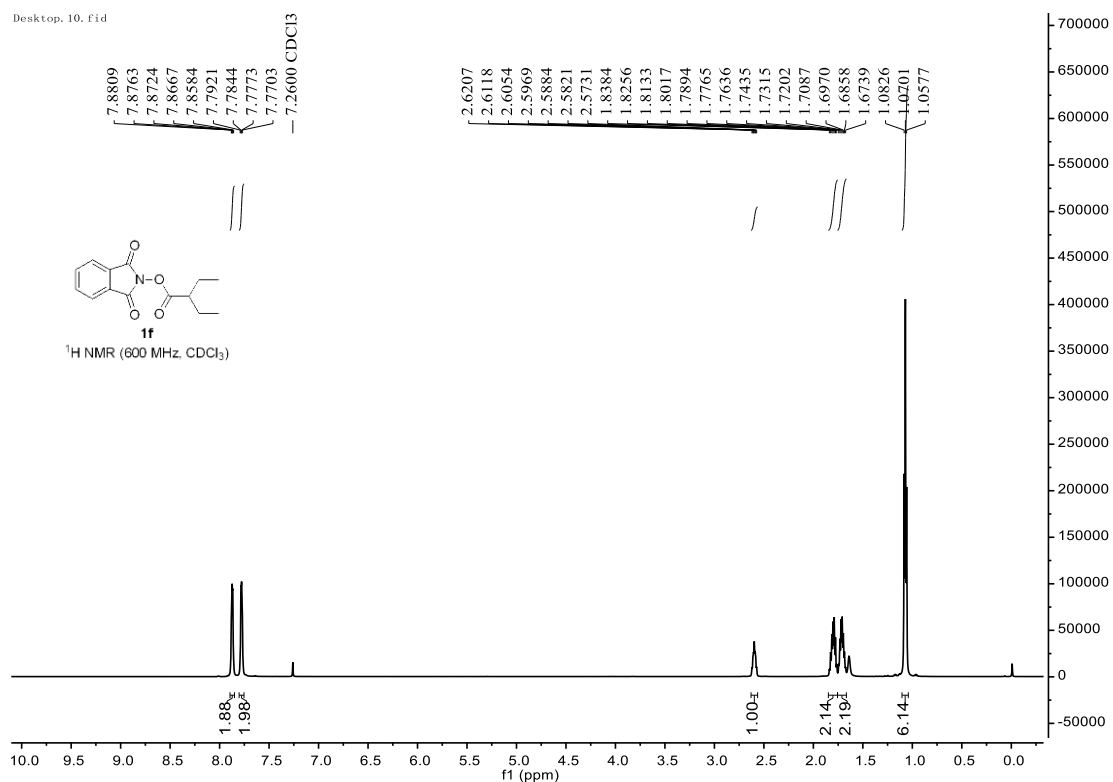
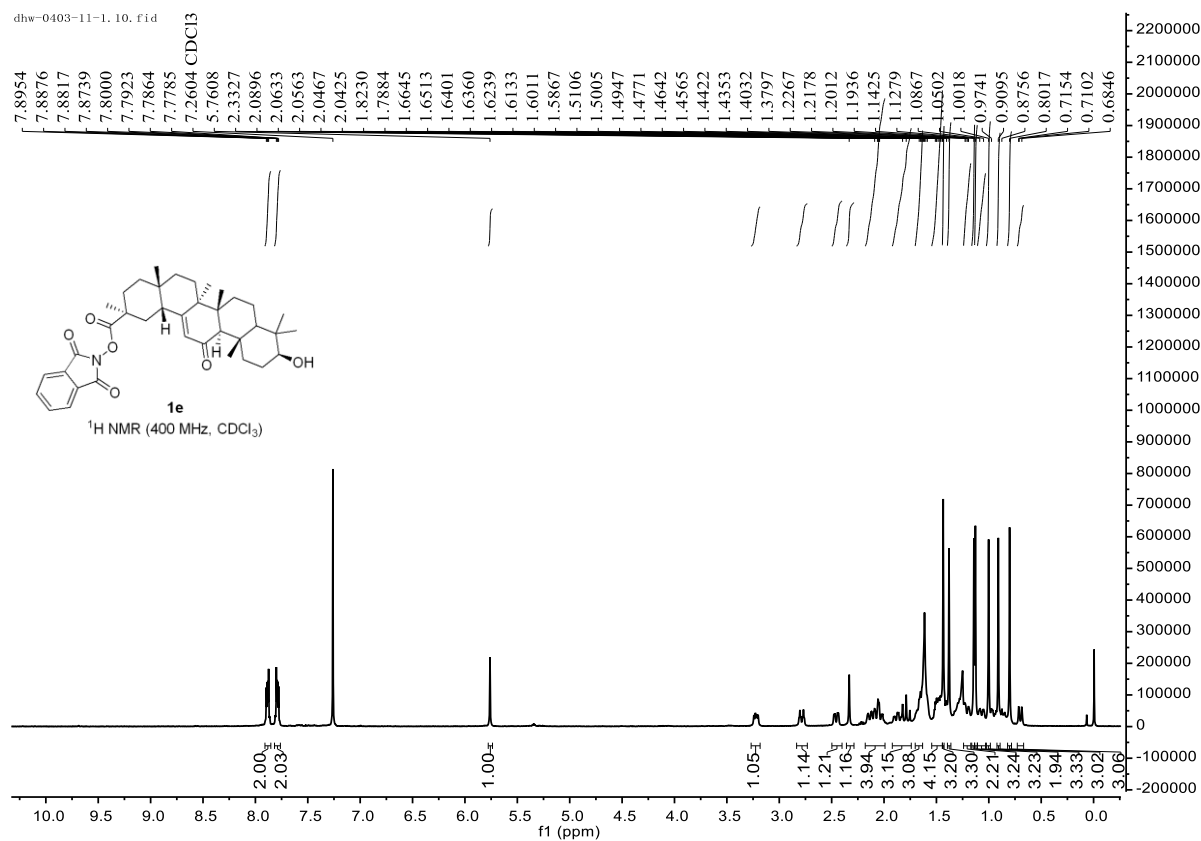
| | |
|--|--|
| Reflections collected | 27244 |
| Independent reflections | 4281 [$R_{\text{int}} = 0.0209$, $R_{\text{sigma}} = 0.0131$] |
| Data/restraints/parameters | 4281/0/274 |
| Goodness-of-fit on F^2 | 1.070 |
| Final R indexes [$I \geq 2\sigma(I)$] | $R_1 = 0.0231$, $wR_2 = 0.0601$ |
| Final R indexes [all data] | $R_1 = 0.0235$, $wR_2 = 0.0603$ |
| Largest diff. peak/hole / $e \text{ \AA}^{-3}$ | 0.37/-0.47 |

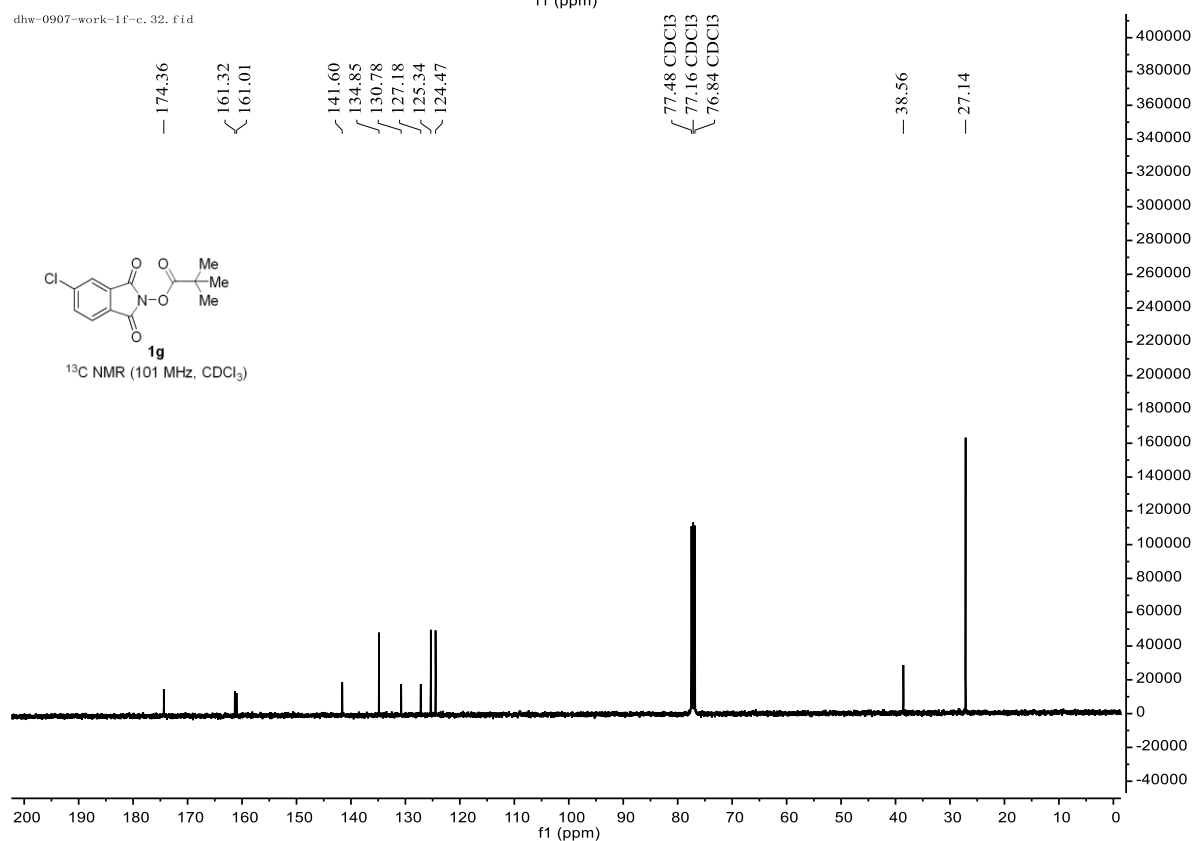
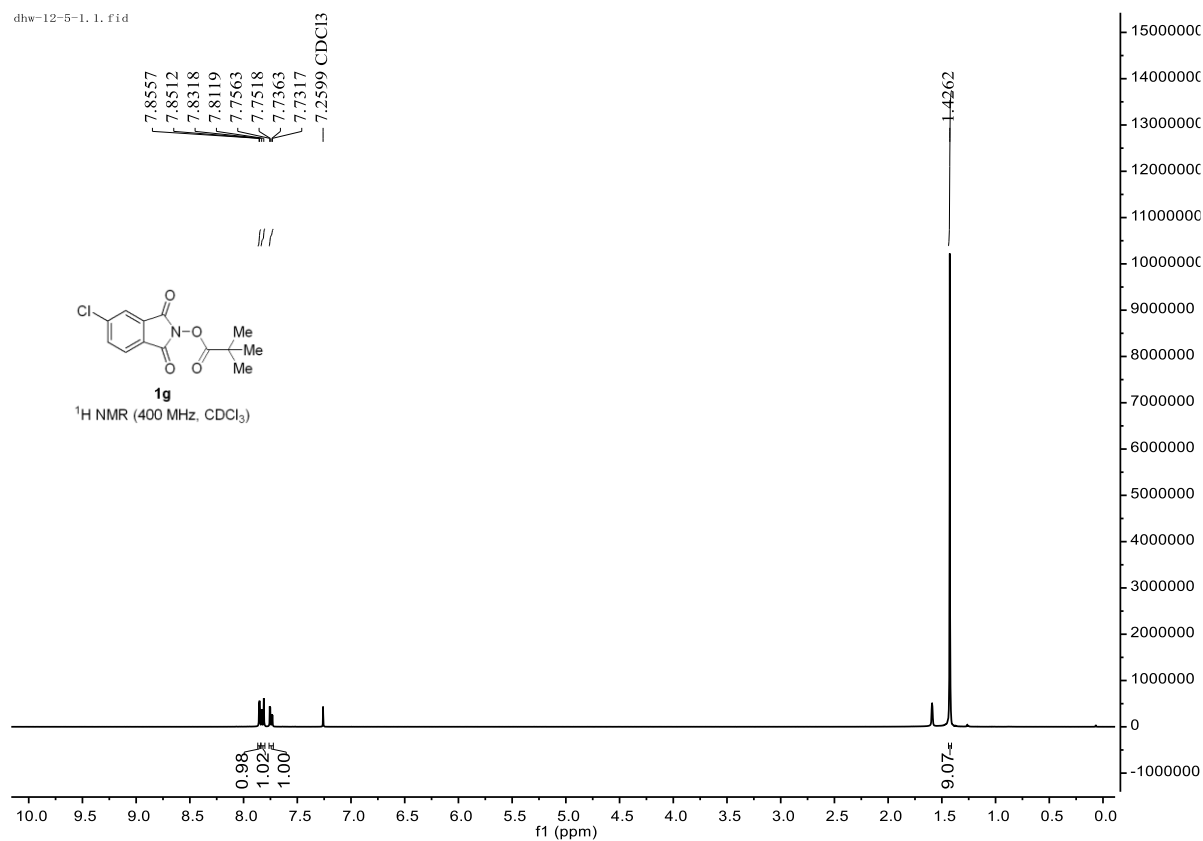
IX. References

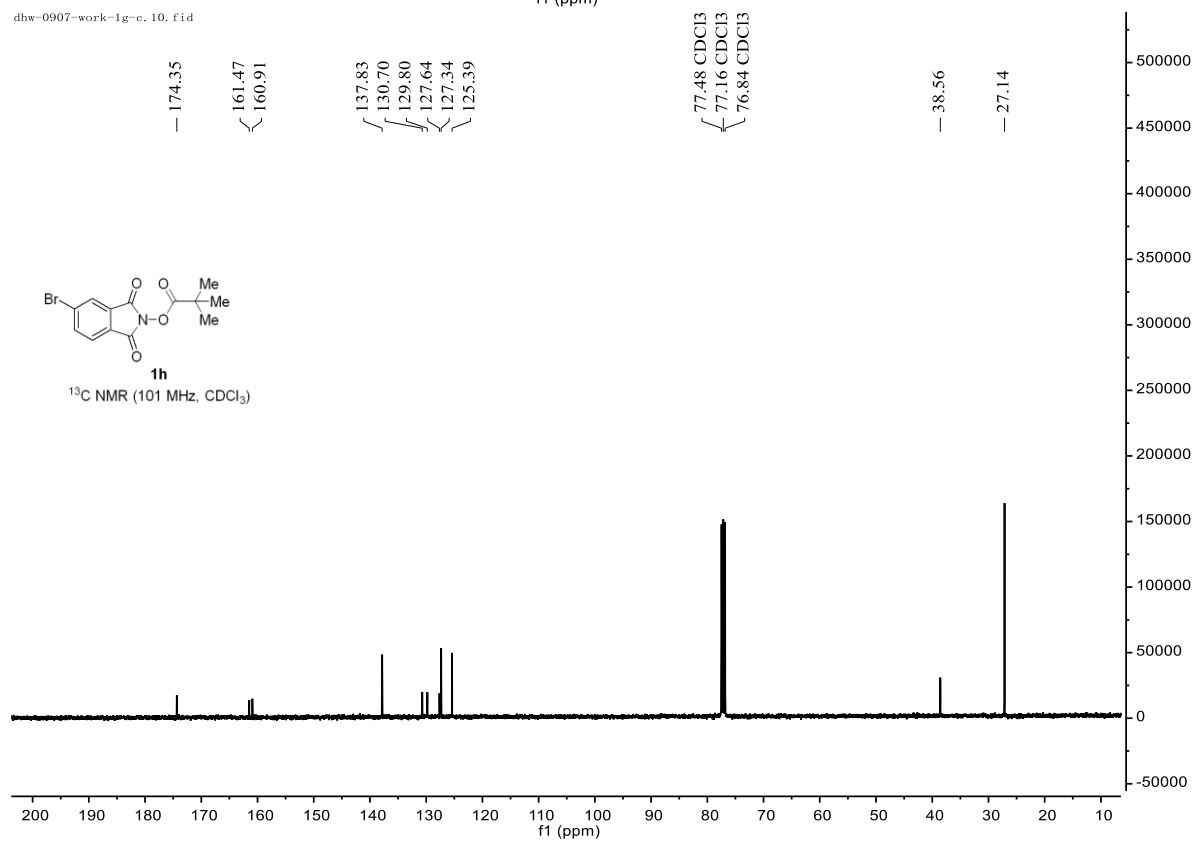
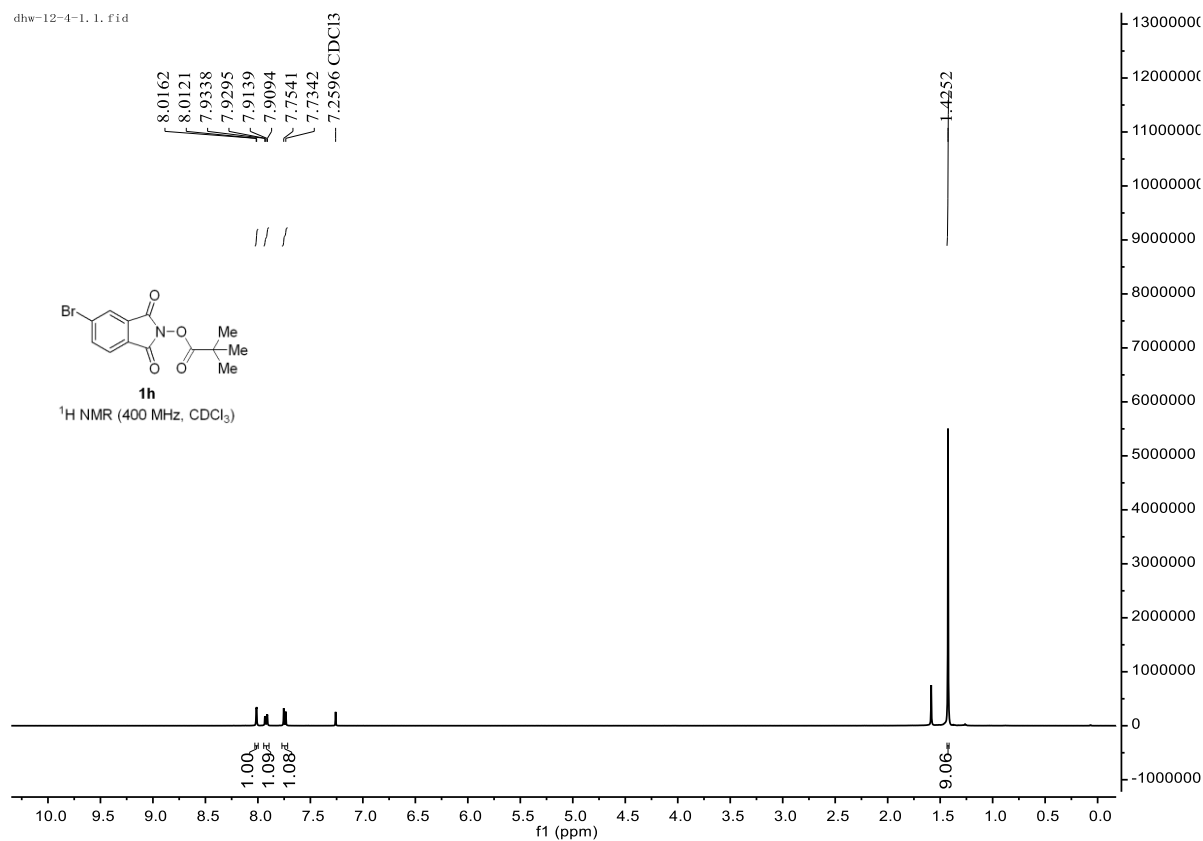
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X. ^1H , ^{13}C and ^{19}F Spectra of Compounds

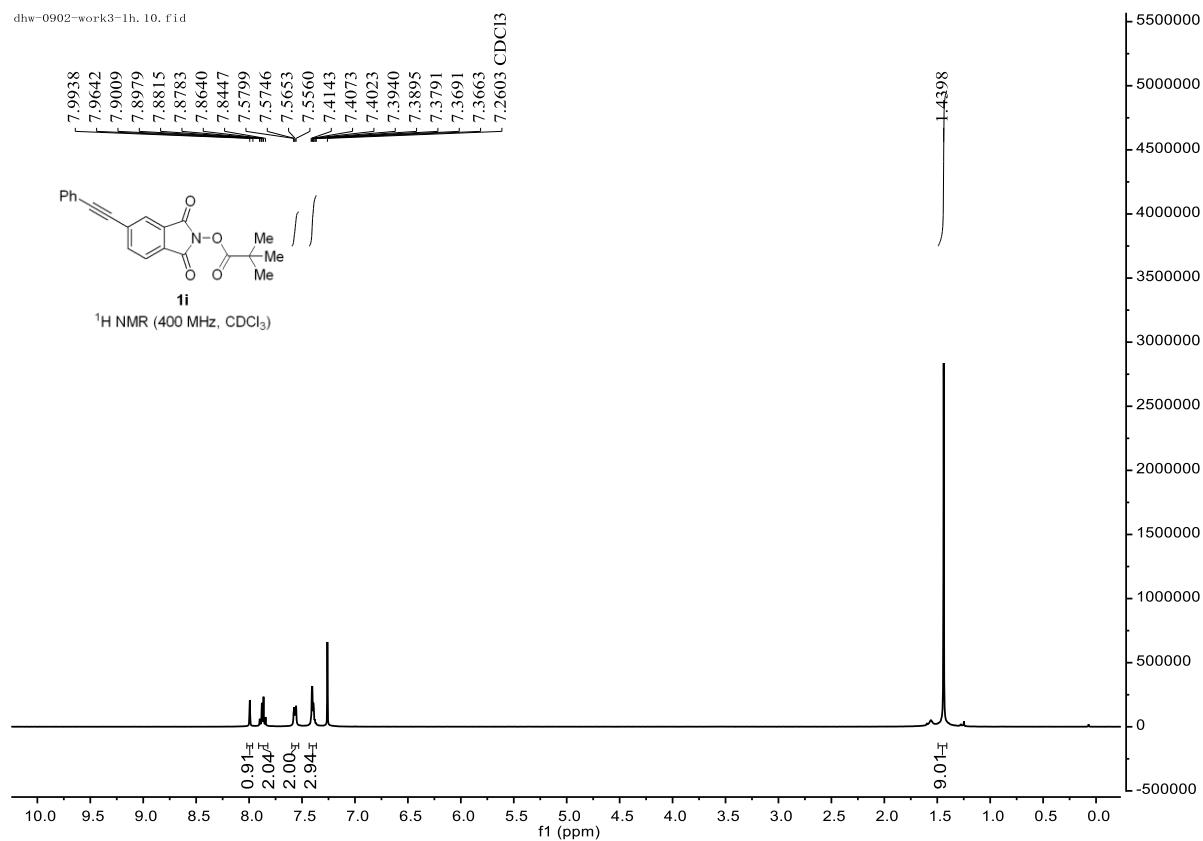




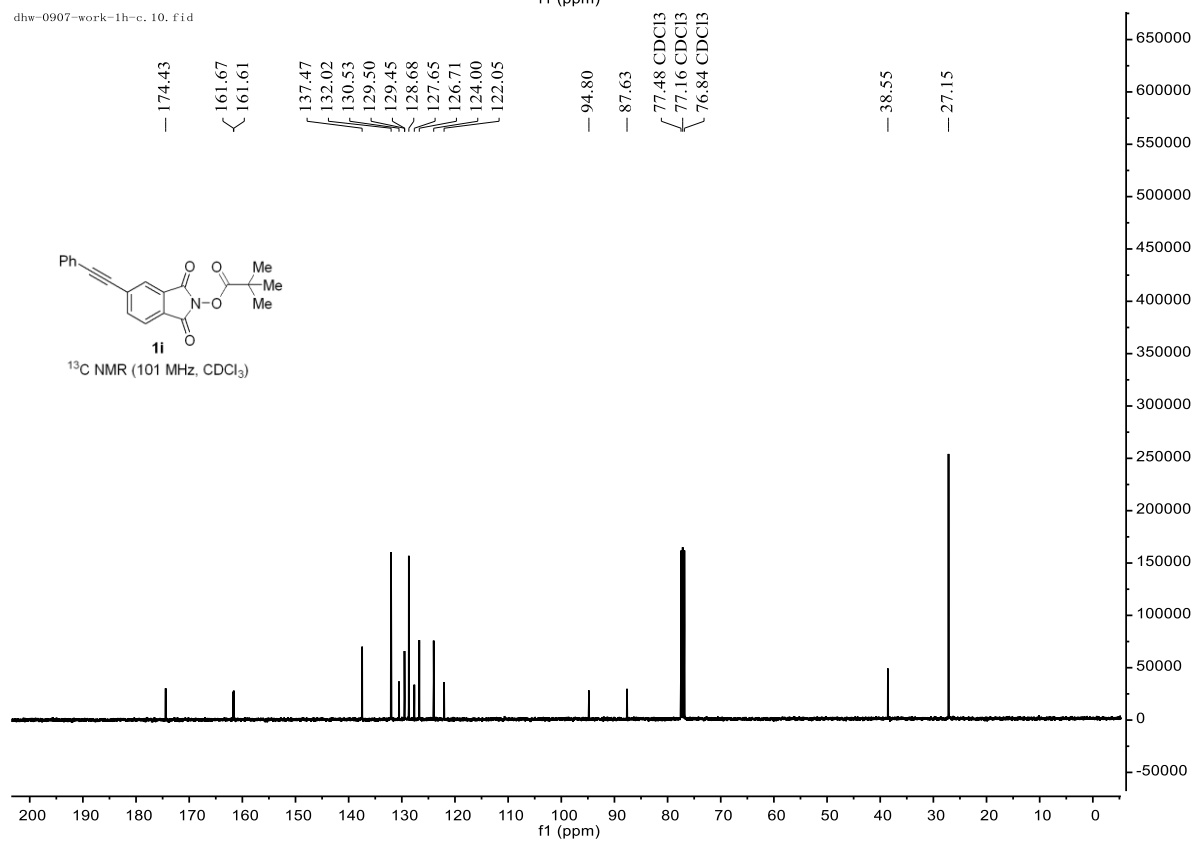




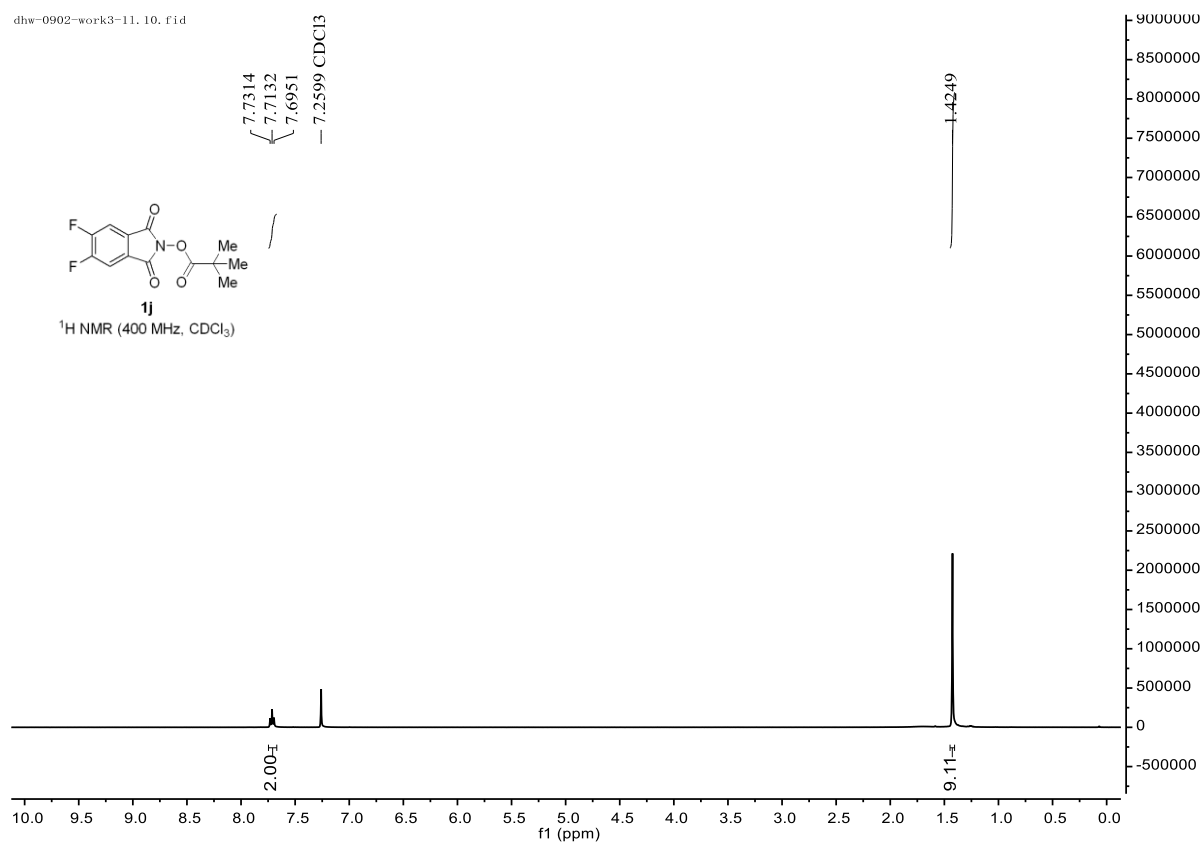
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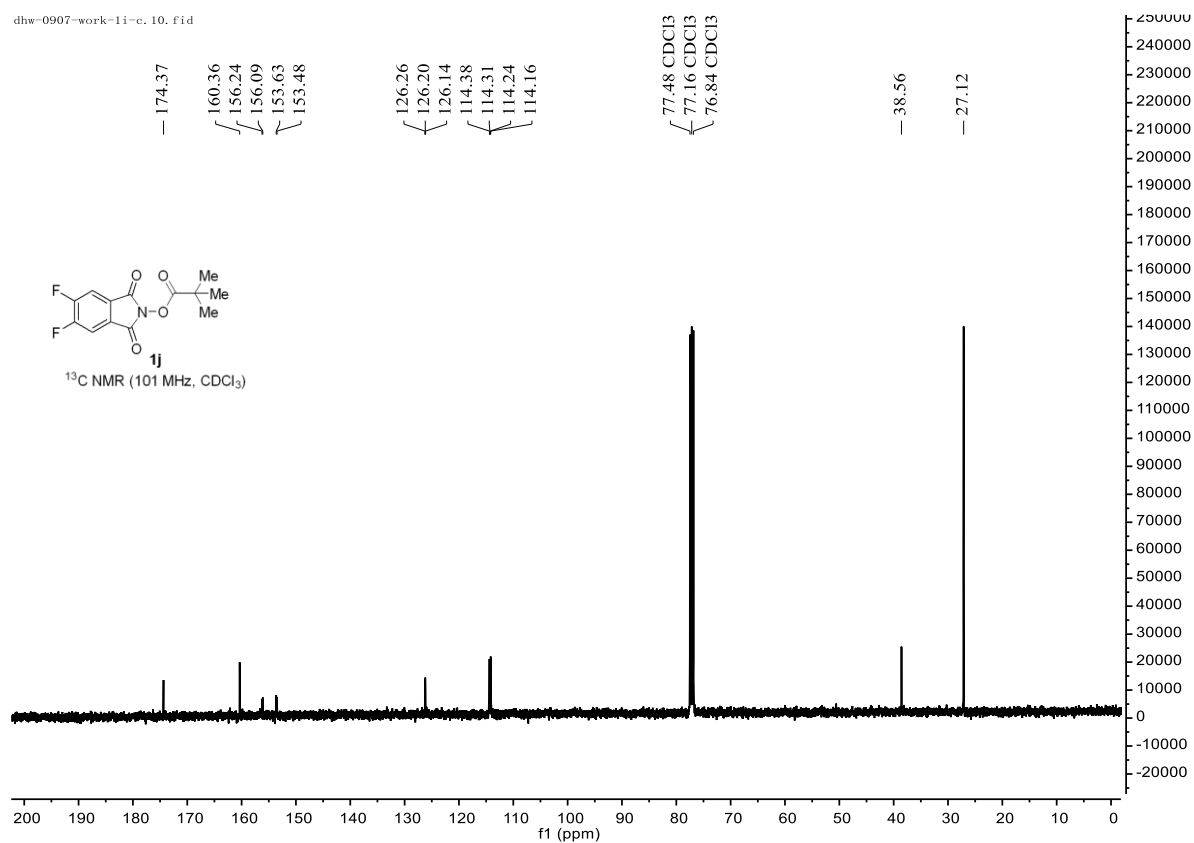
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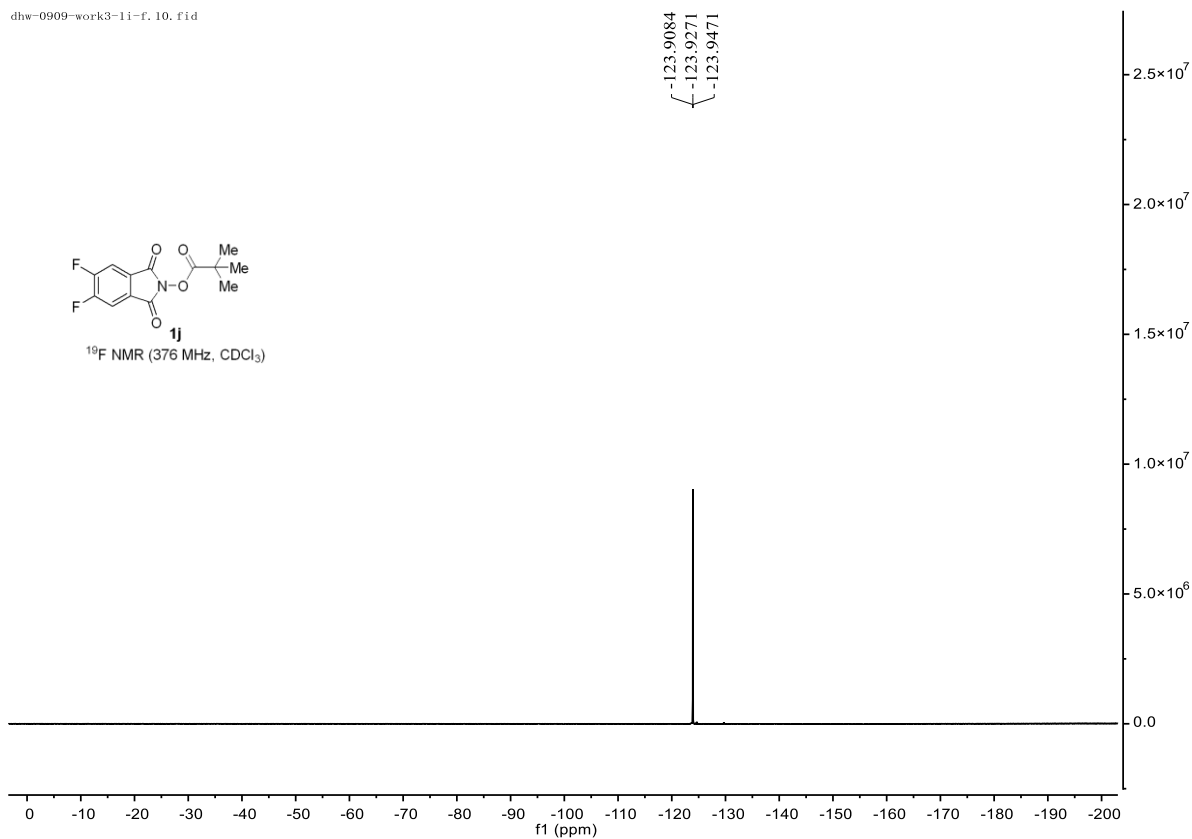
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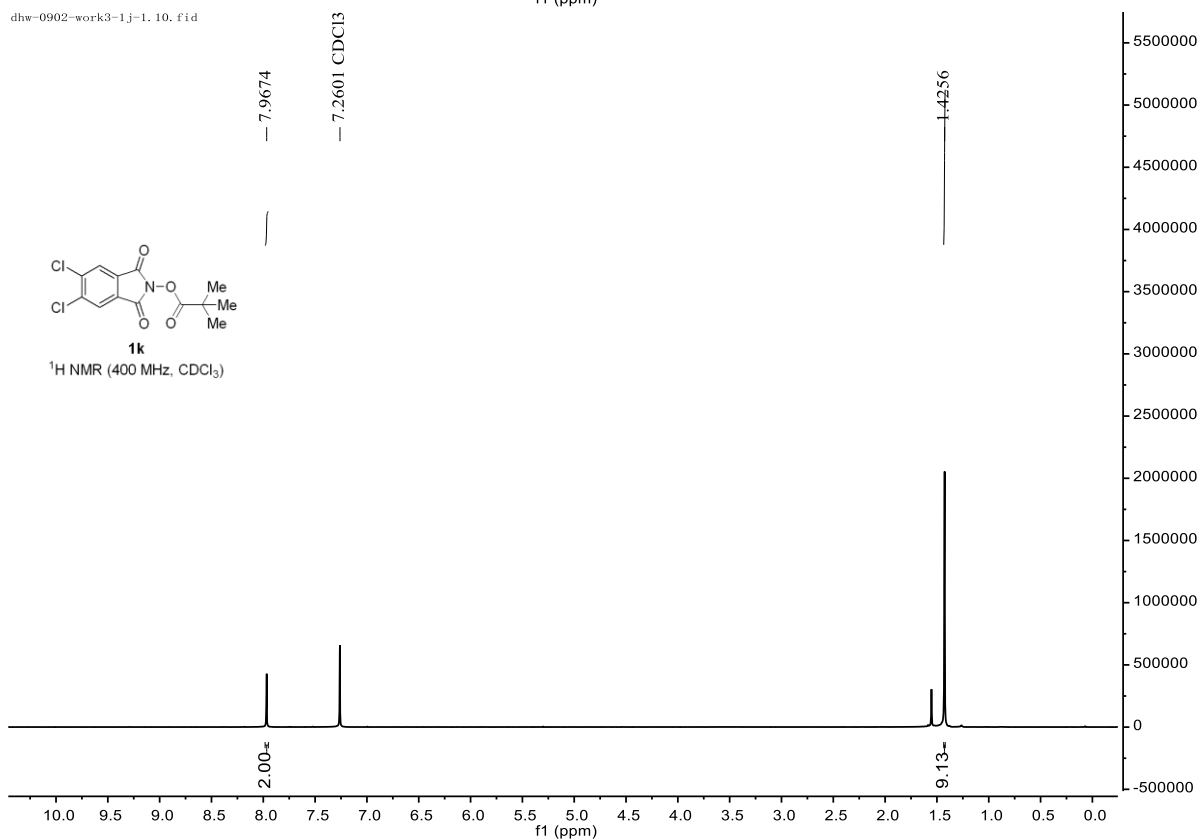
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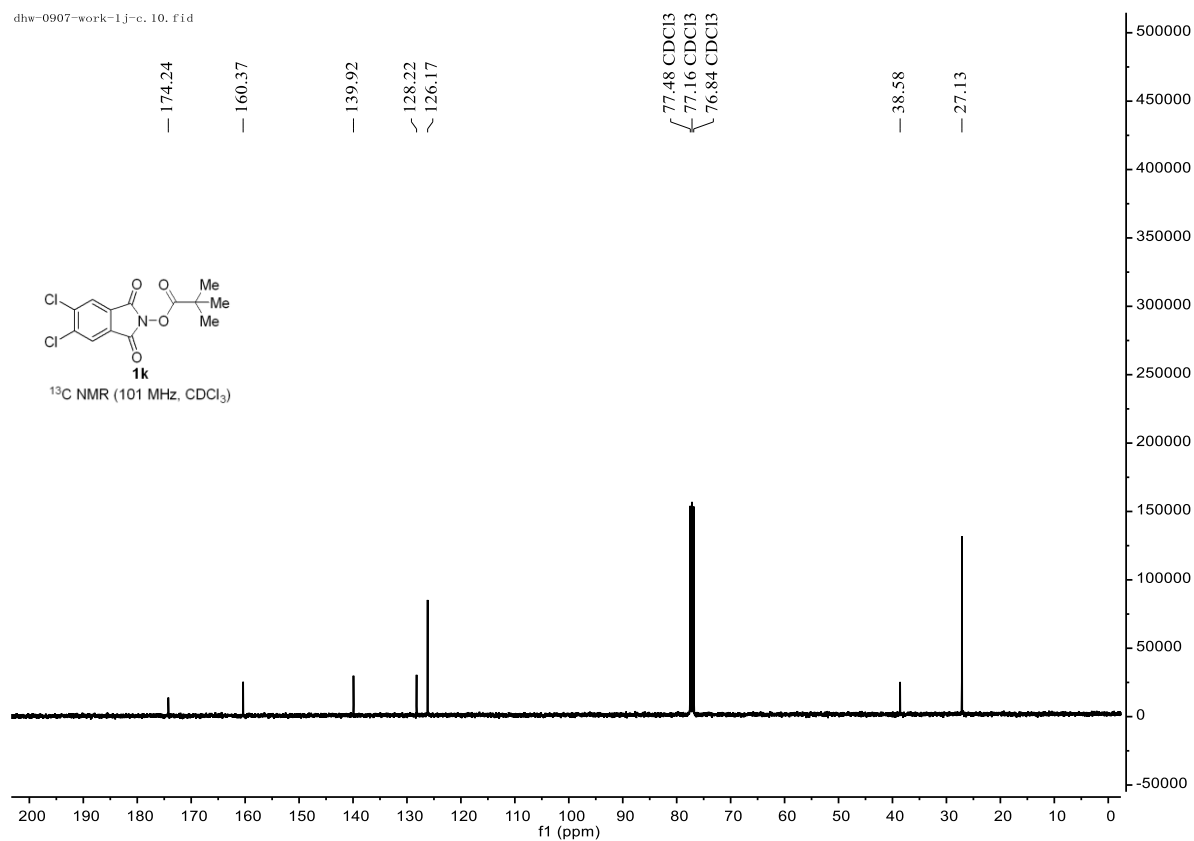
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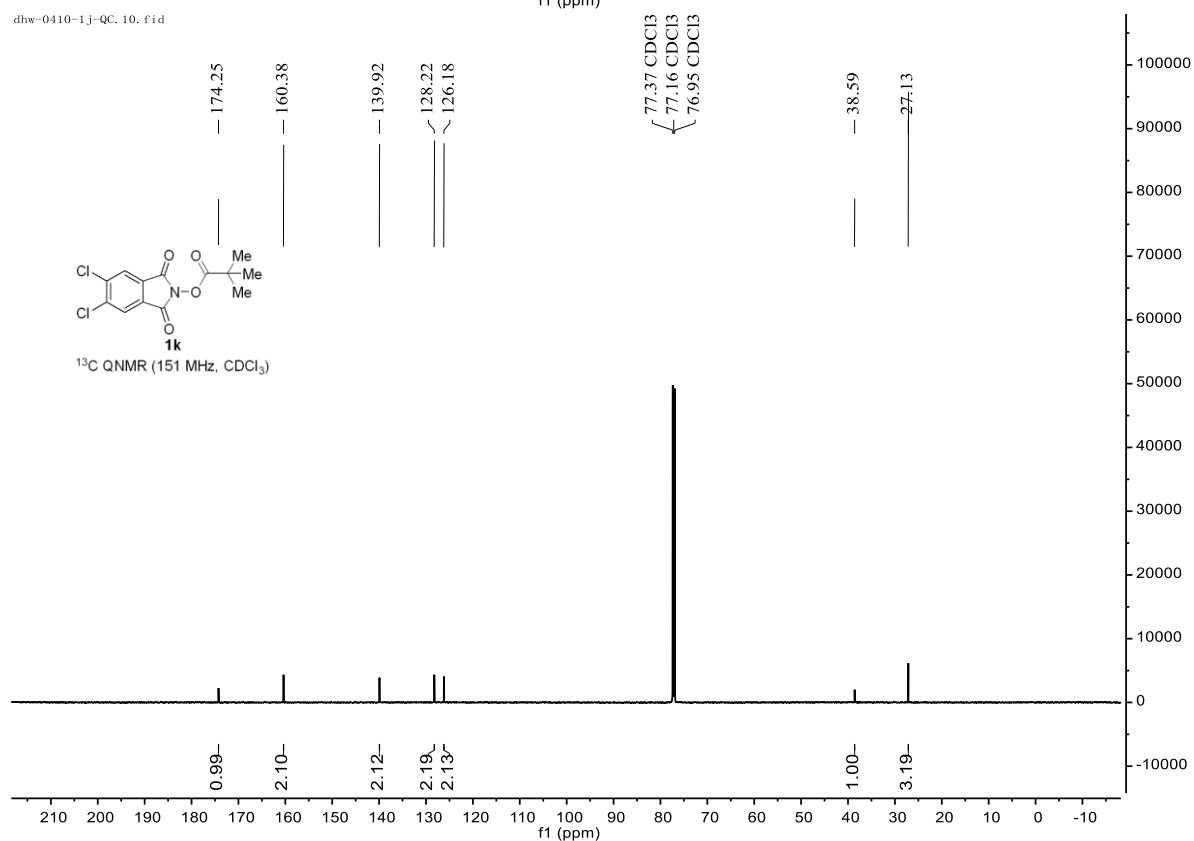
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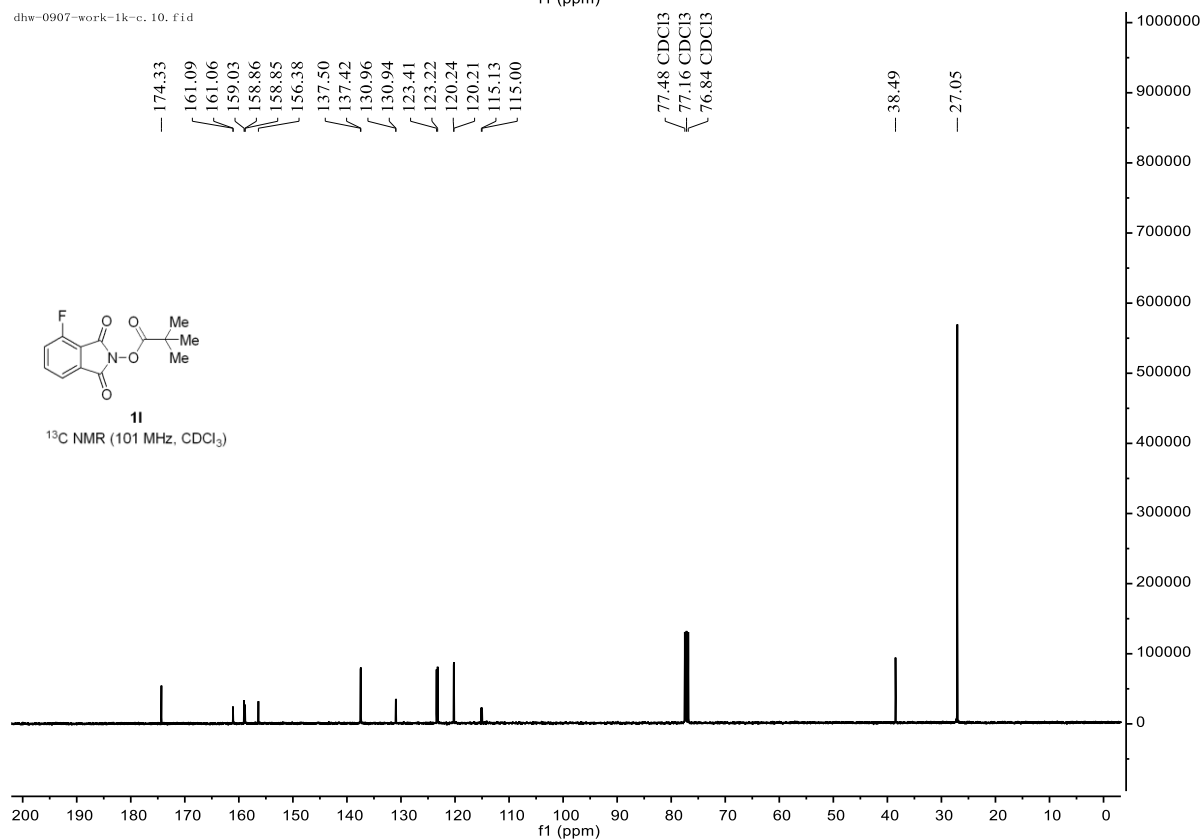
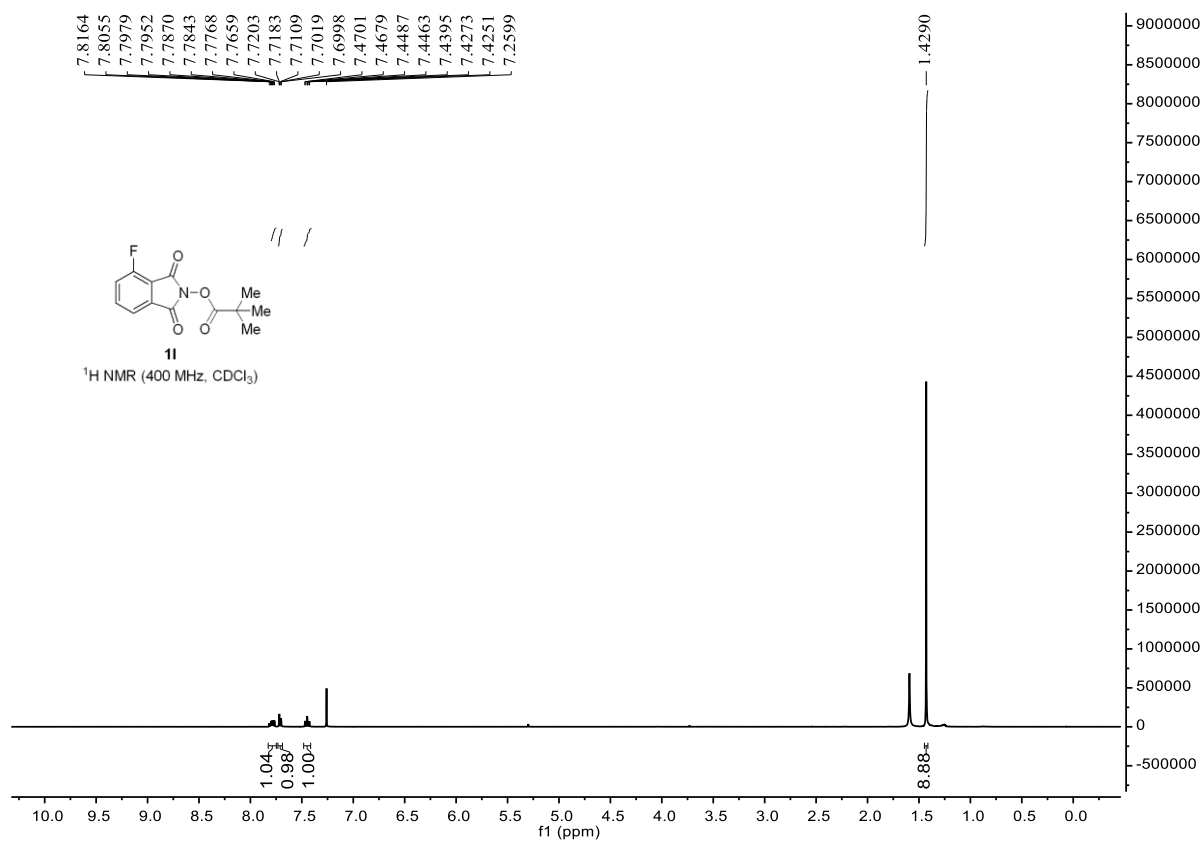


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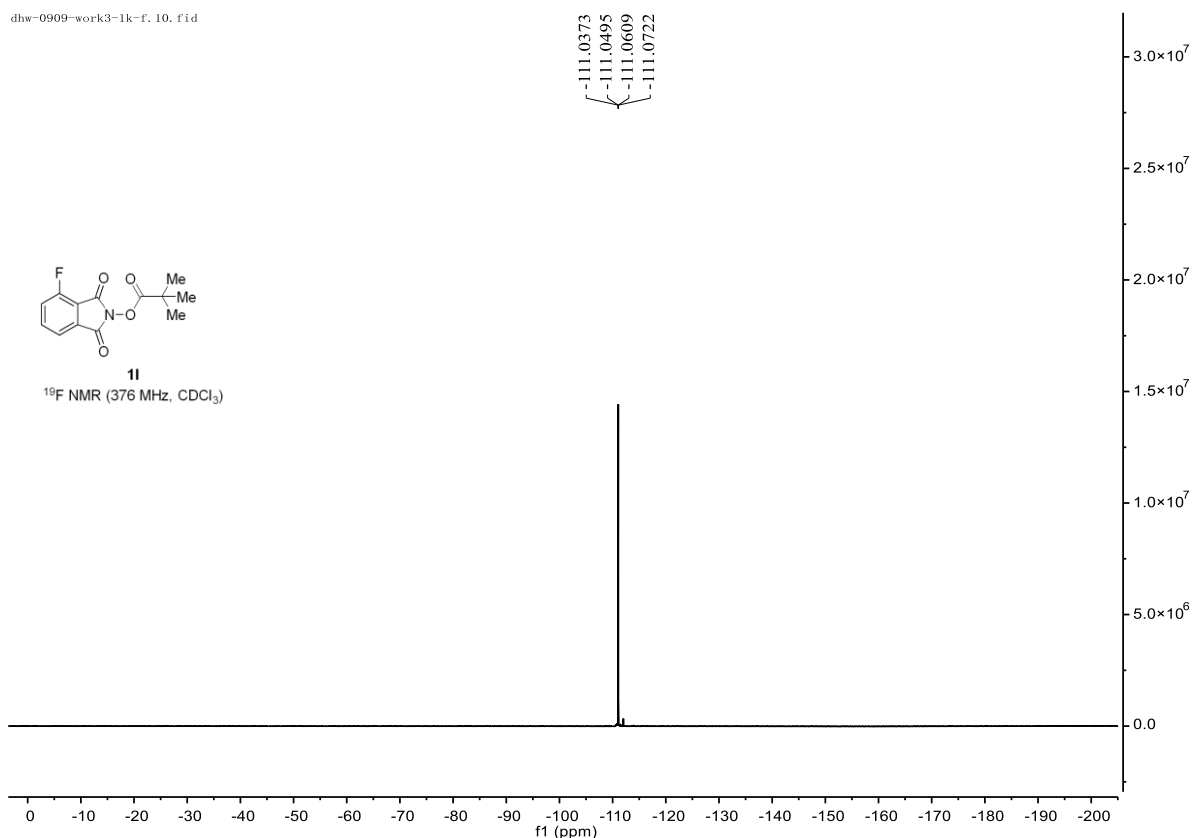


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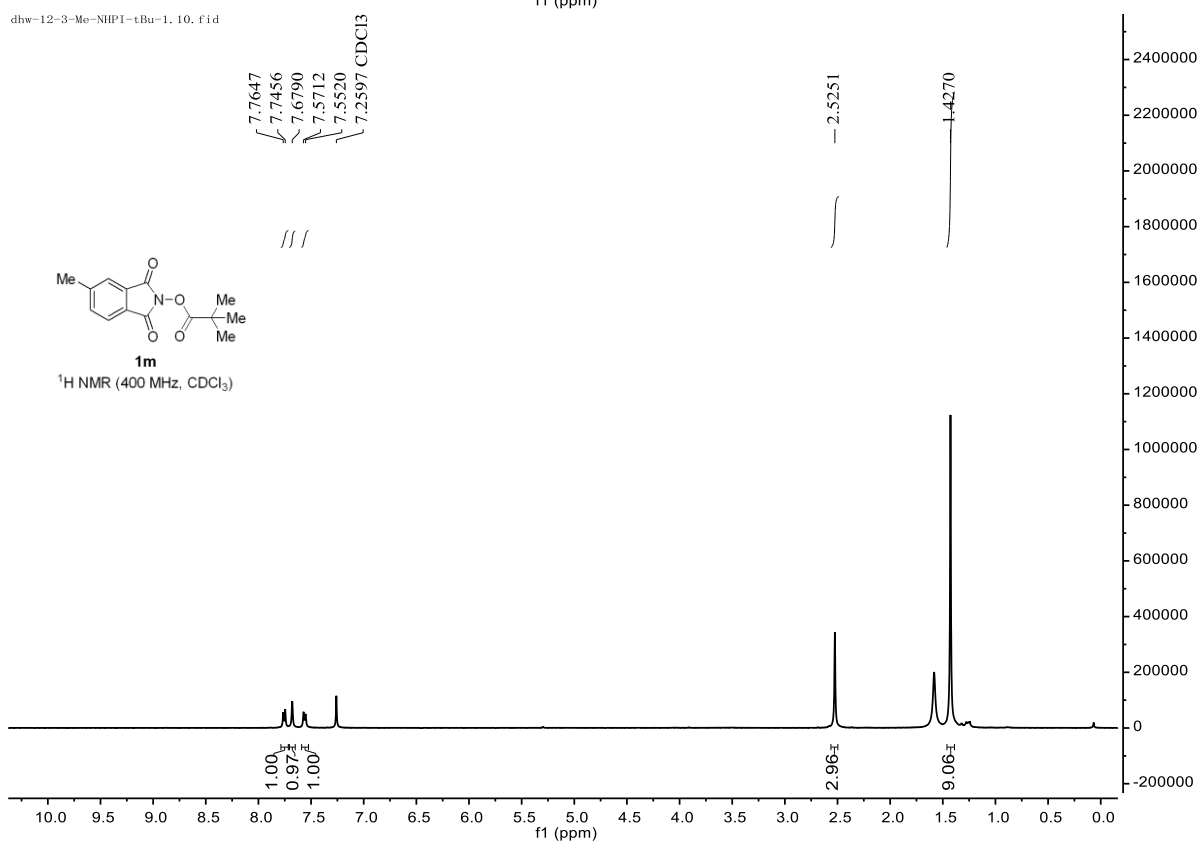




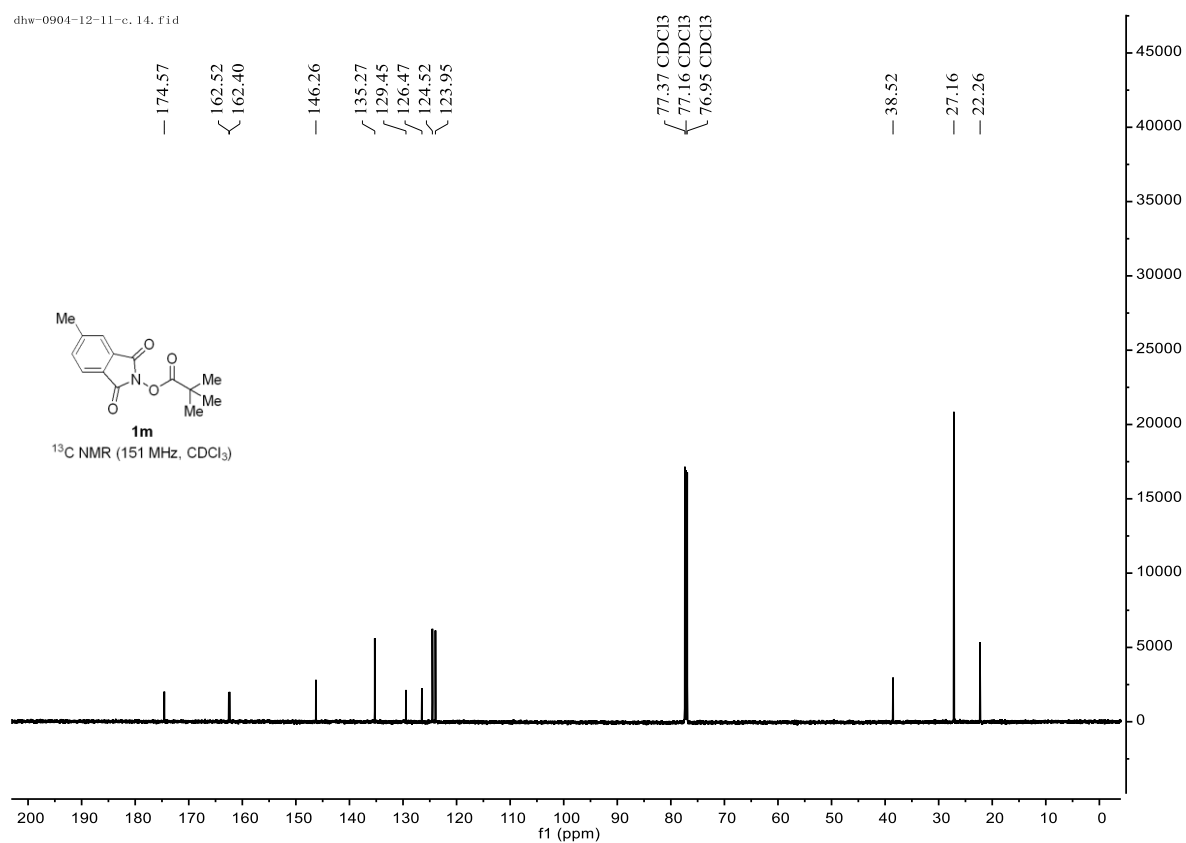
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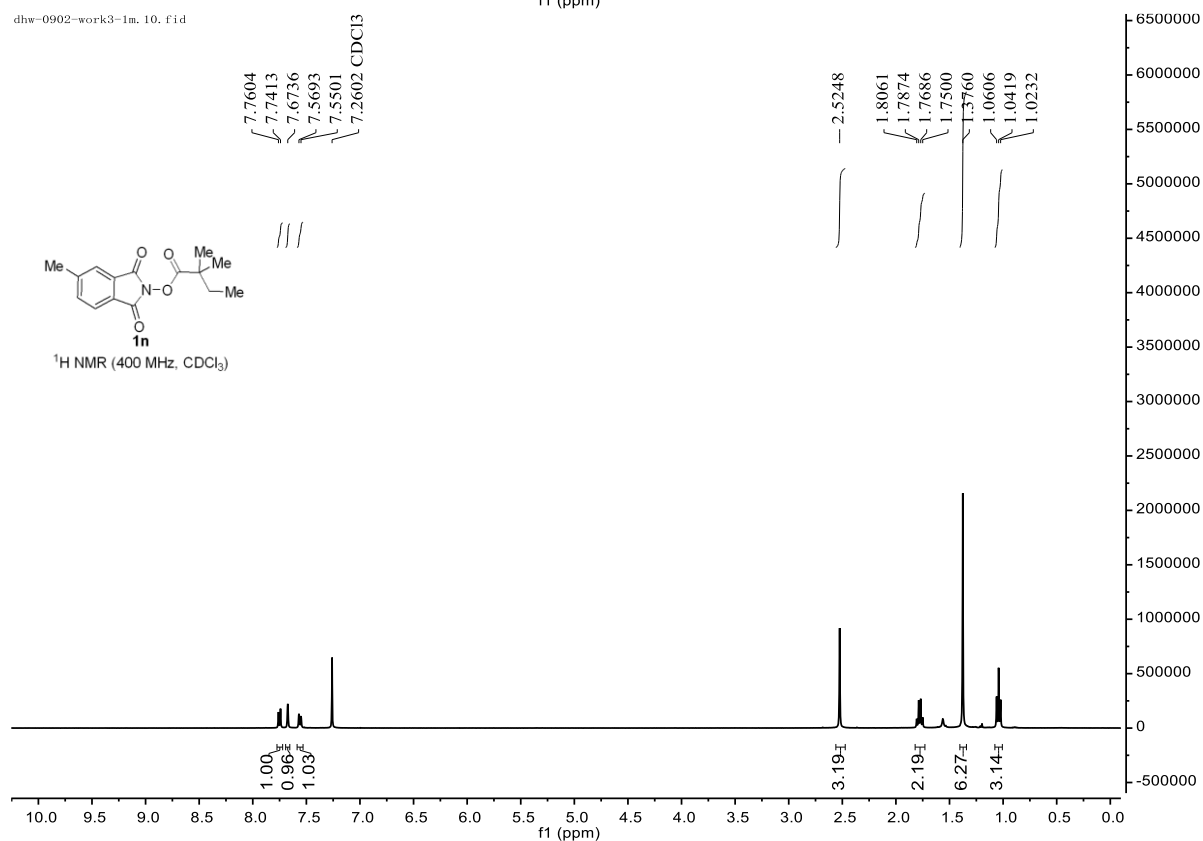
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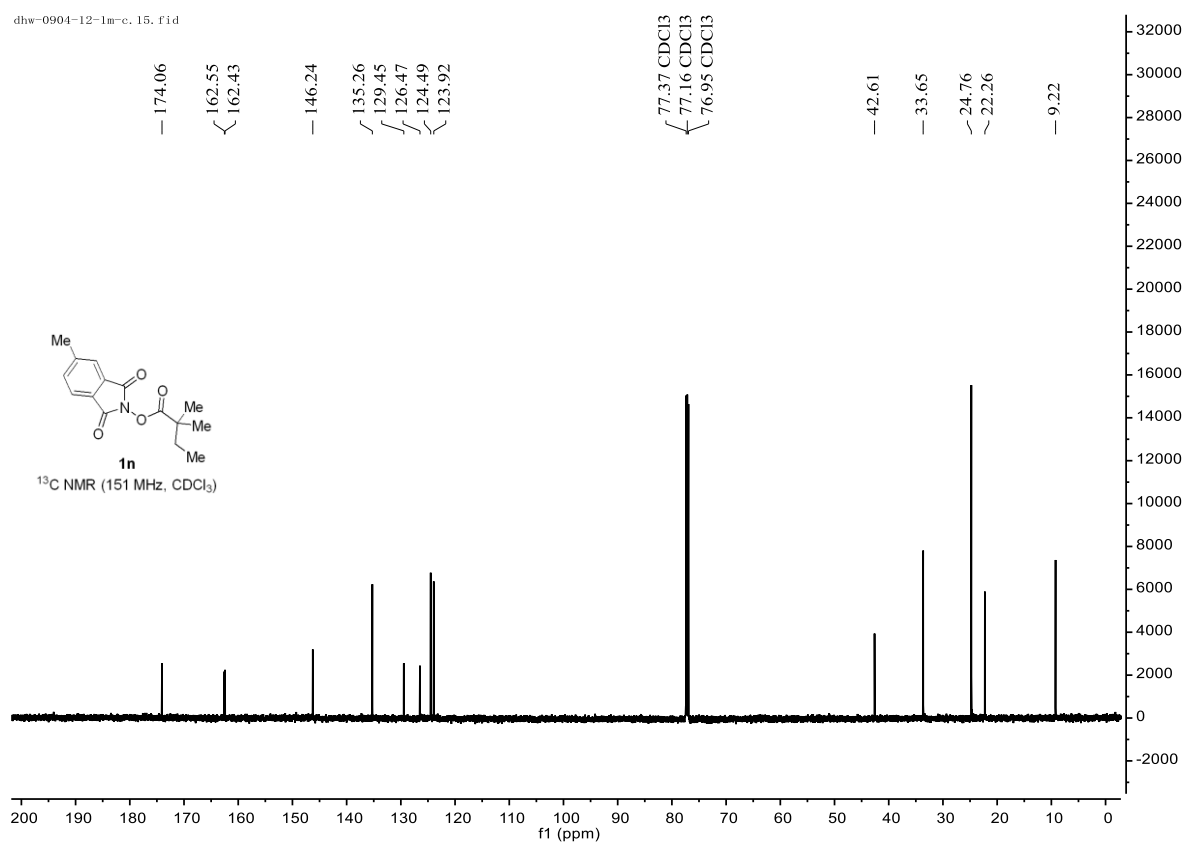
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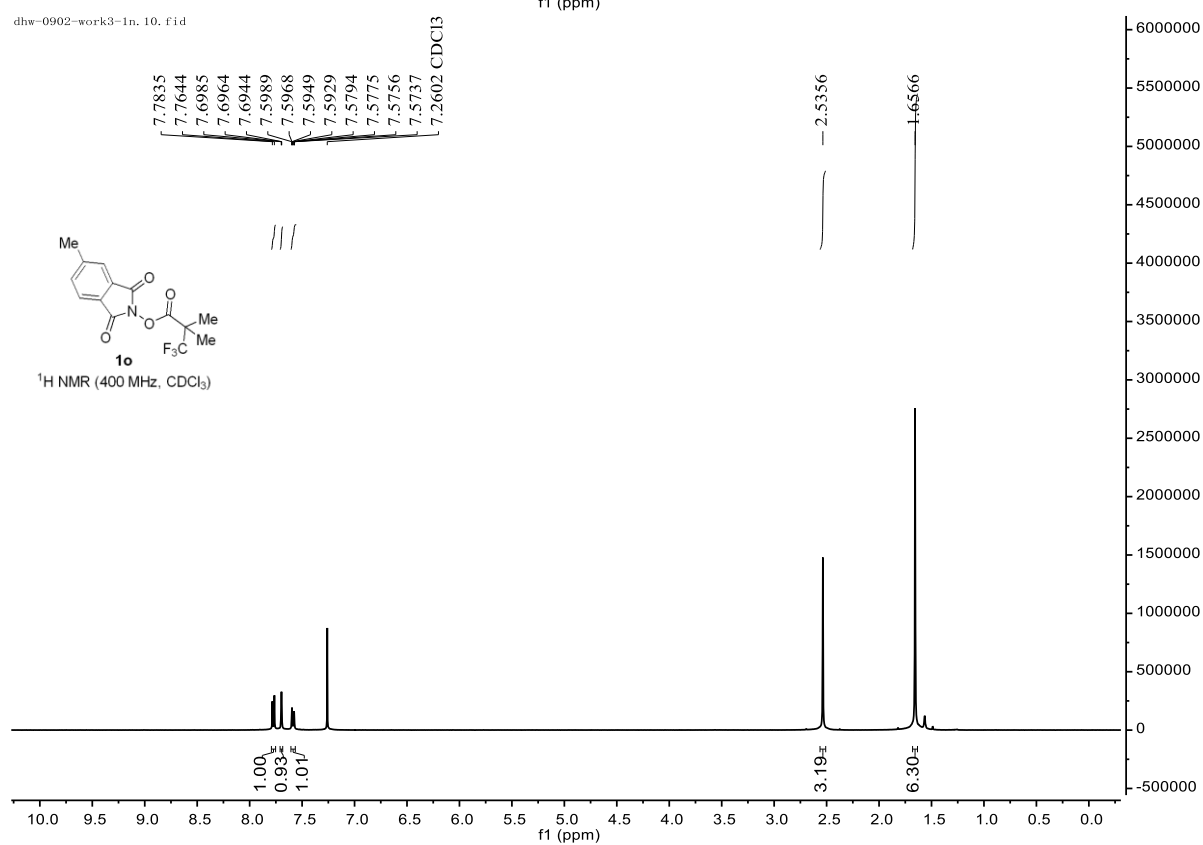
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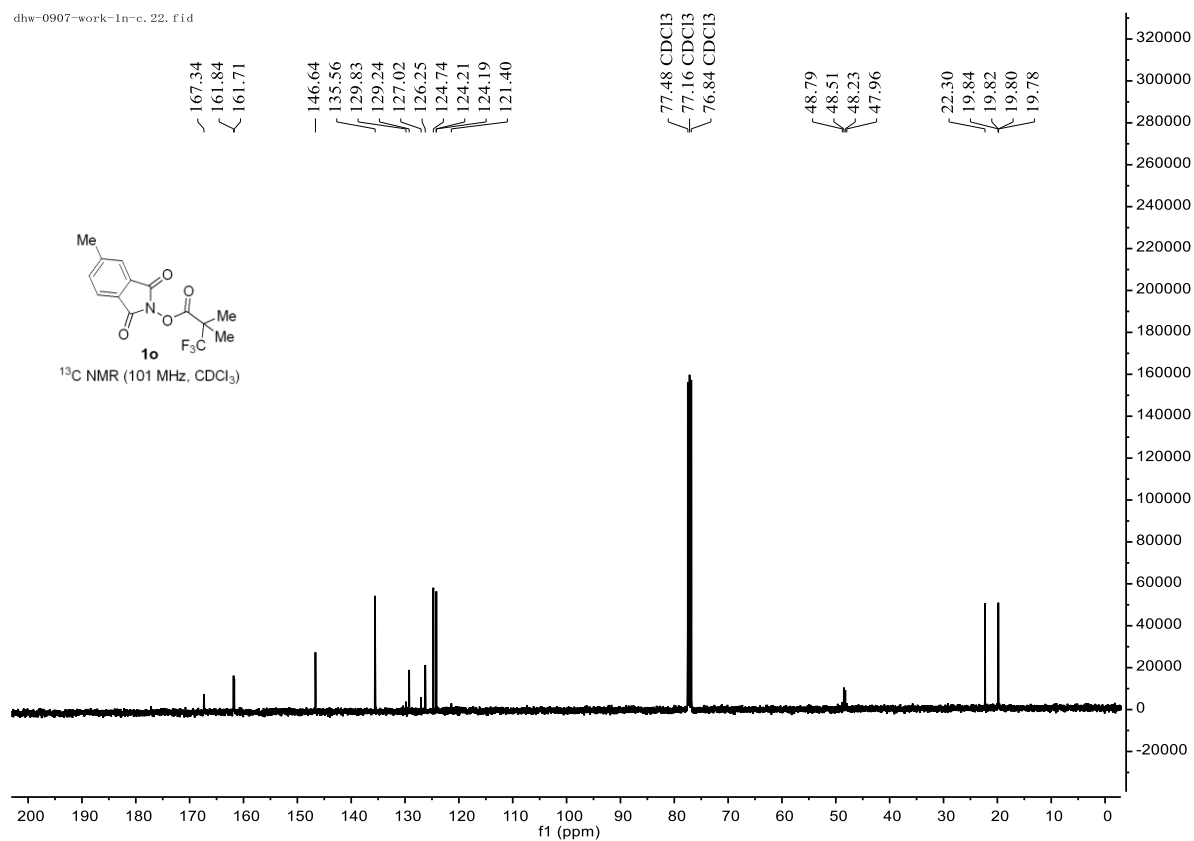
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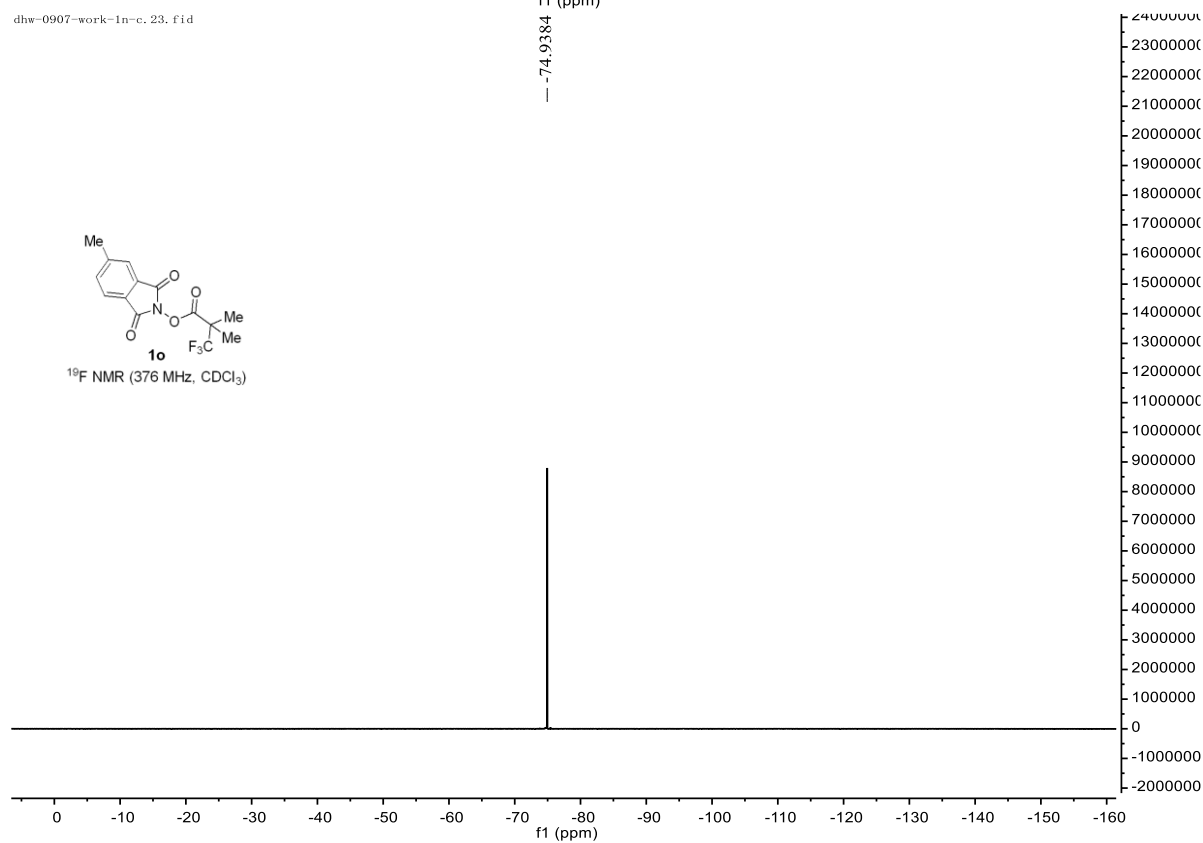
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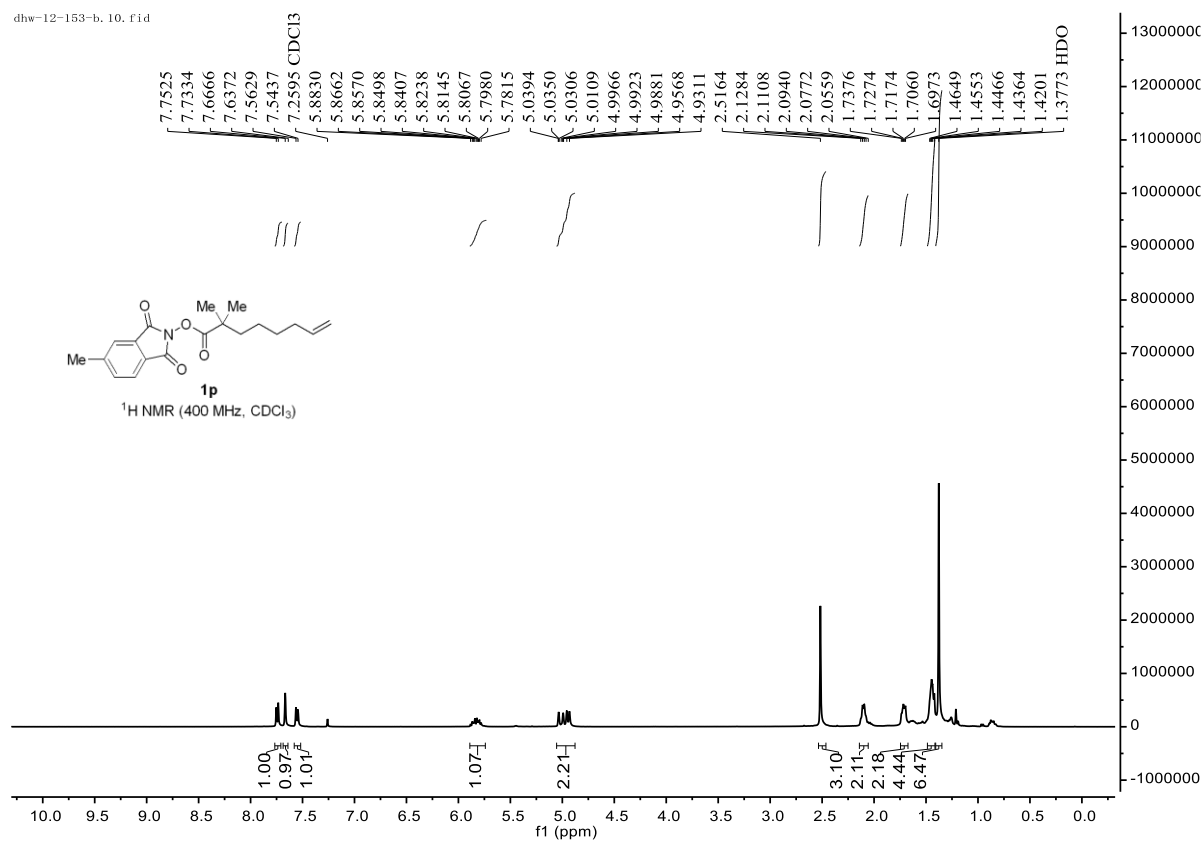
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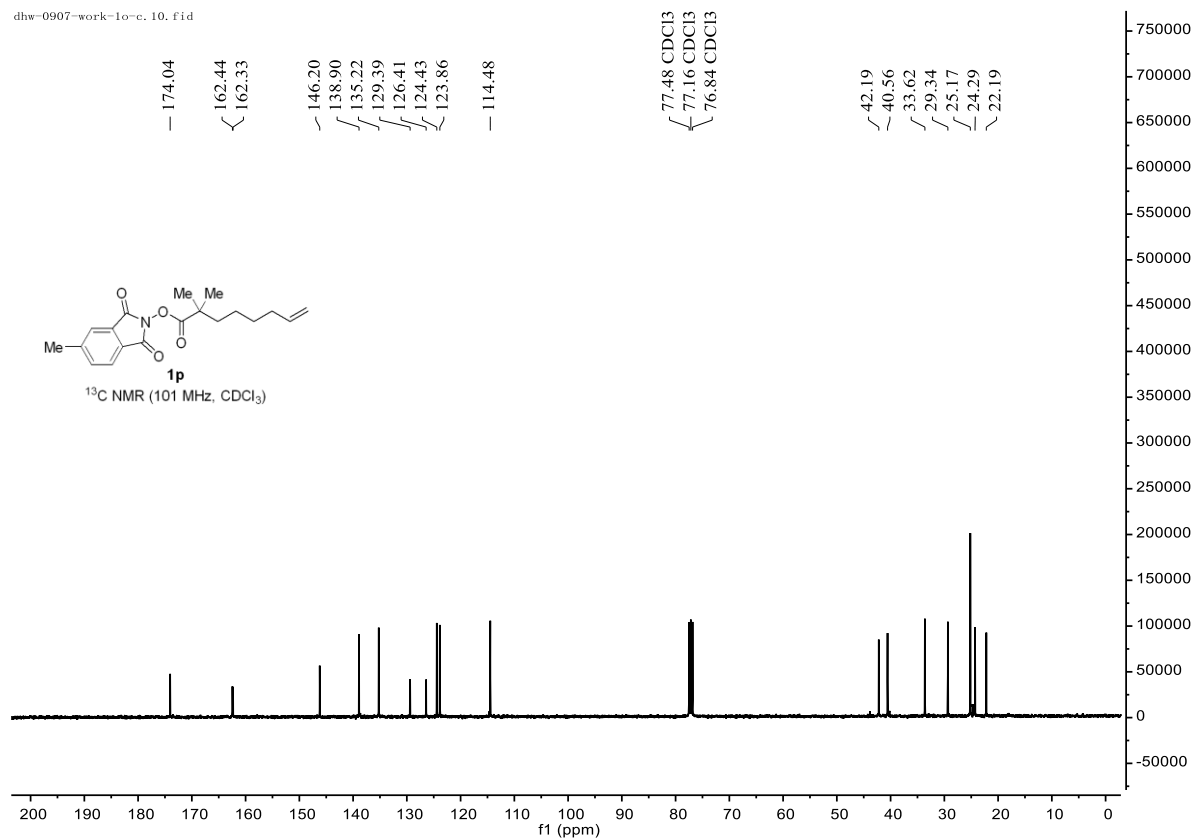
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dhw-0902-work3-1p. 10. f1d

1q

¹H NMR (400 MHz, CDCl₃)

Chemical structure of **1q**: CC1=C(C(=O)N2C(=O)c3ccc(C)cc3C2=O)C(C)C(C)COc4ccc(C)cc4

Peak list (ppm):

- 7.7675, 7.7483, 7.6803, 7.5771, 7.5579, 7.2600, 7.0109, 6.9921, 6.6667, 6.6494
- 4.0211, 4.0128, 4.0079, 3.9961
- 2.5309, 2.3133, 2.2974, 2.1885, 1.9614, 1.9498, 1.9427, 1.9267, 1.4441

Integration values:

- 1.00, 0.95, 1.00 (aromatic region)
- 1.05, 2.04 (aromatic region)
- 1.97 (methoxy singlet)
- 3.00, 3.30, 3.18, 4.22 (aliphatic region)
- 6.08 (aliphatic region)

dhw-0907-work-1p-c. 10. fid

1q

¹³C NMR (101 MHz, CDCl₃)

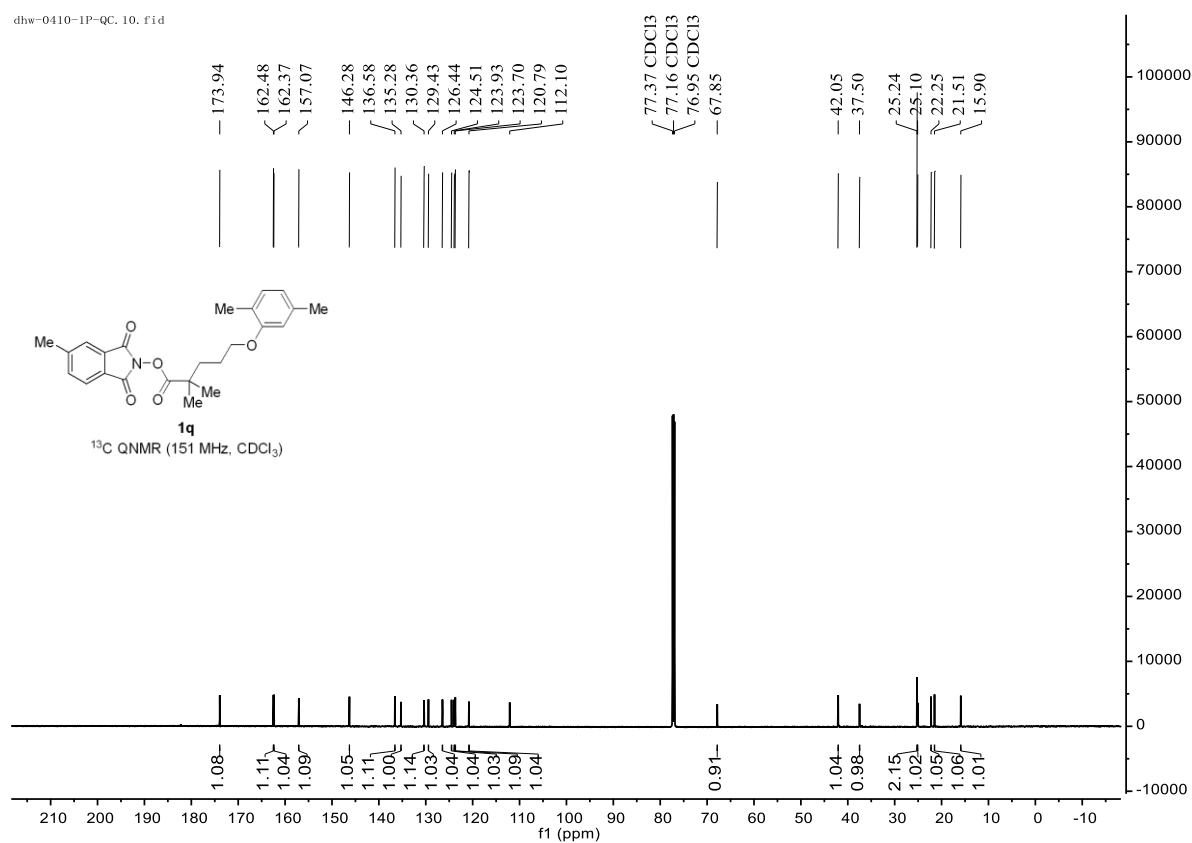
Chemical structure of **1q** is shown. The structure is a 5-methyl-2-((4-methoxyphenyl)butyl)isobenzofuran-1-one derivative. The structure is labeled **1q**.

¹³C NMR (101 MHz, CDCl₃) spectrum is shown. The x-axis is labeled f1 (ppm) and ranges from 0 to 200. The y-axis is labeled f2 (ppm) and ranges from 0 to 400,000. The spectrum shows several peaks, with the following chemical shifts (ppm) labeled above the peaks:

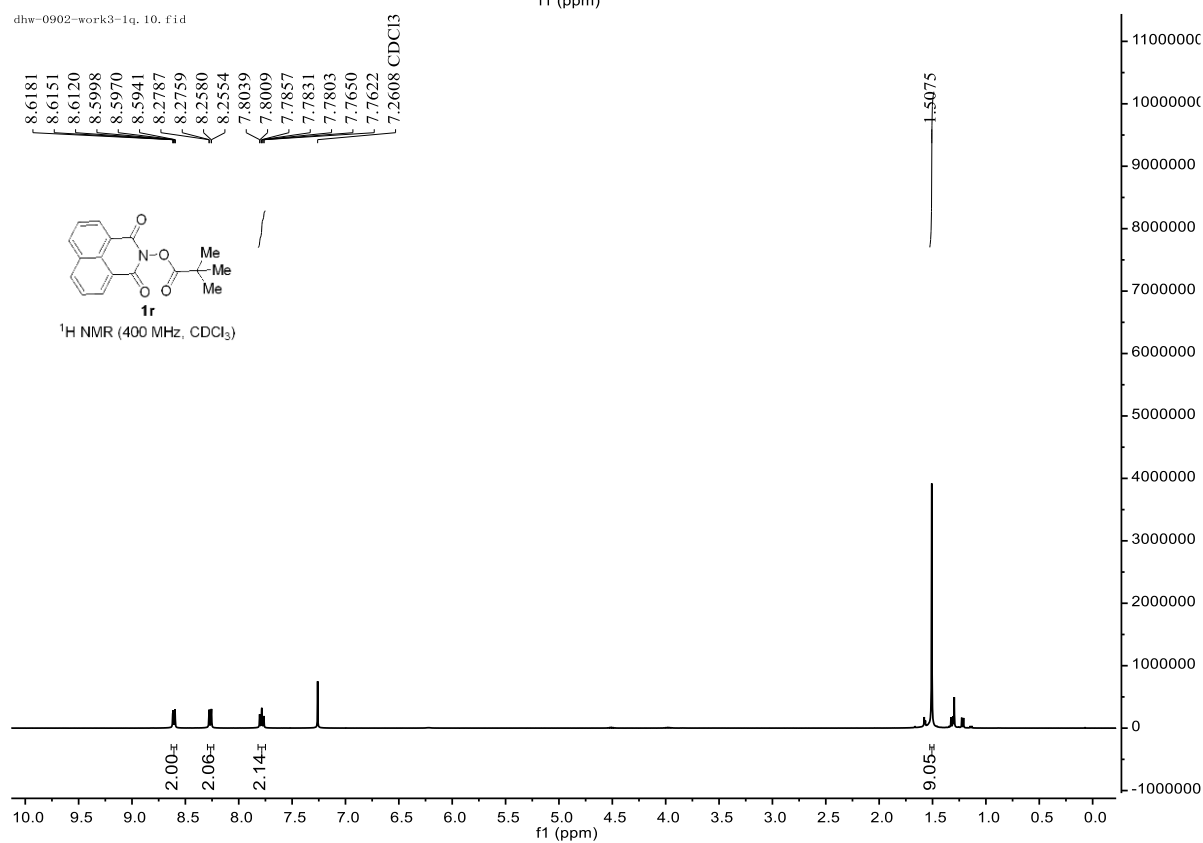
- 173.94
- 162.49
- 162.38
- 157.08
- 146.28
- 136.59
- 135.29
- 130.37
- 129.44
- 126.45
- 124.51
- 123.94
- 123.71
- 120.80
- 112.11
- 77.48 CDCl₃
- 77.16 CDCl₃
- 76.84 CDCl₃
- 67.86
- 42.06
- 37.51
- 25.25
- 25.11
- 22.26
- 21.51
- 15.90

The spectrum shows a complex pattern of peaks, with the most intense peak at 77.16 ppm, which is the solvent peak for CDCl₃. The peaks are labeled with their chemical shifts in ppm.

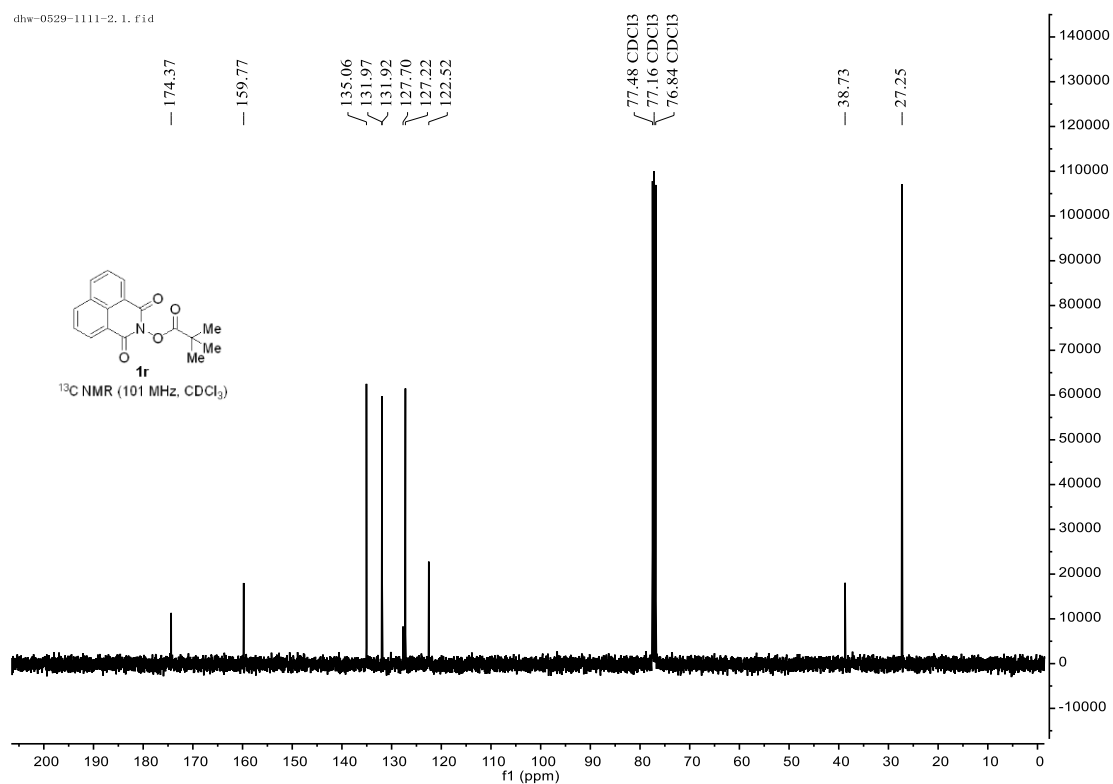
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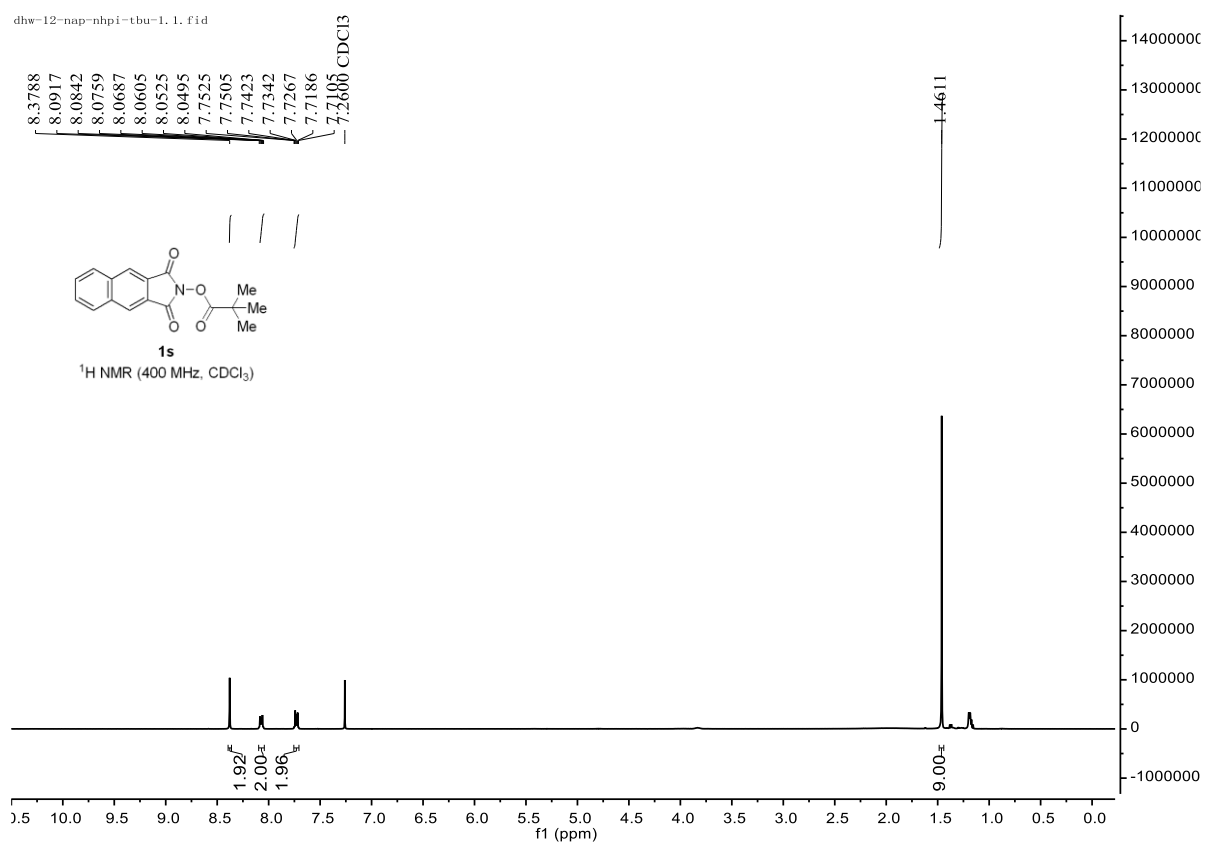
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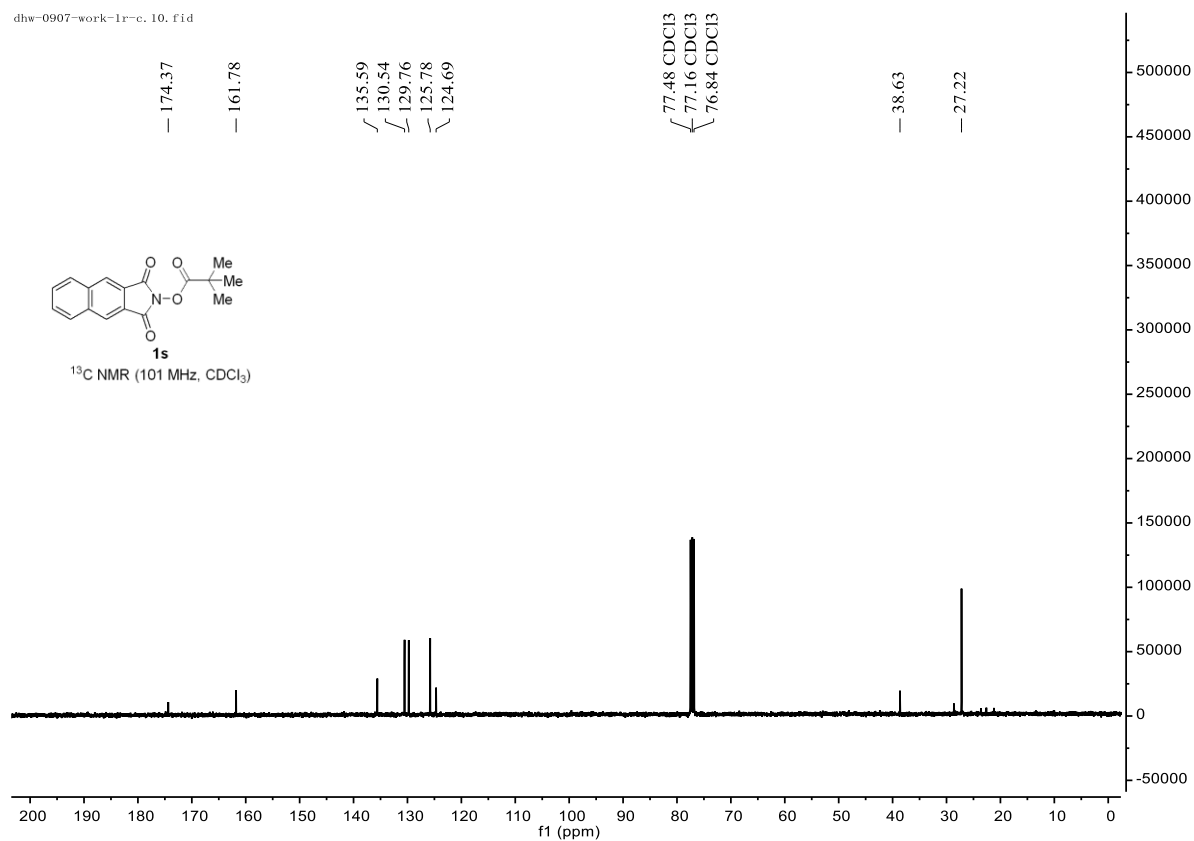
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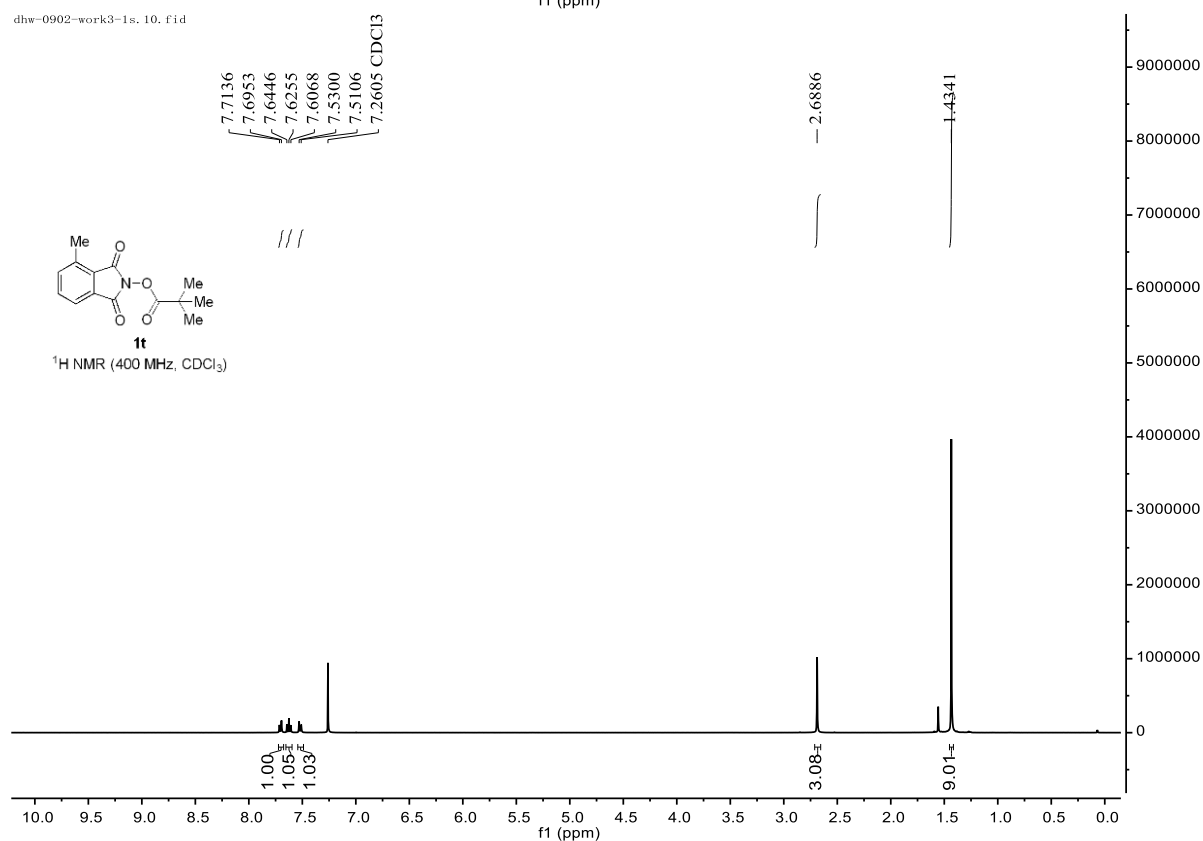
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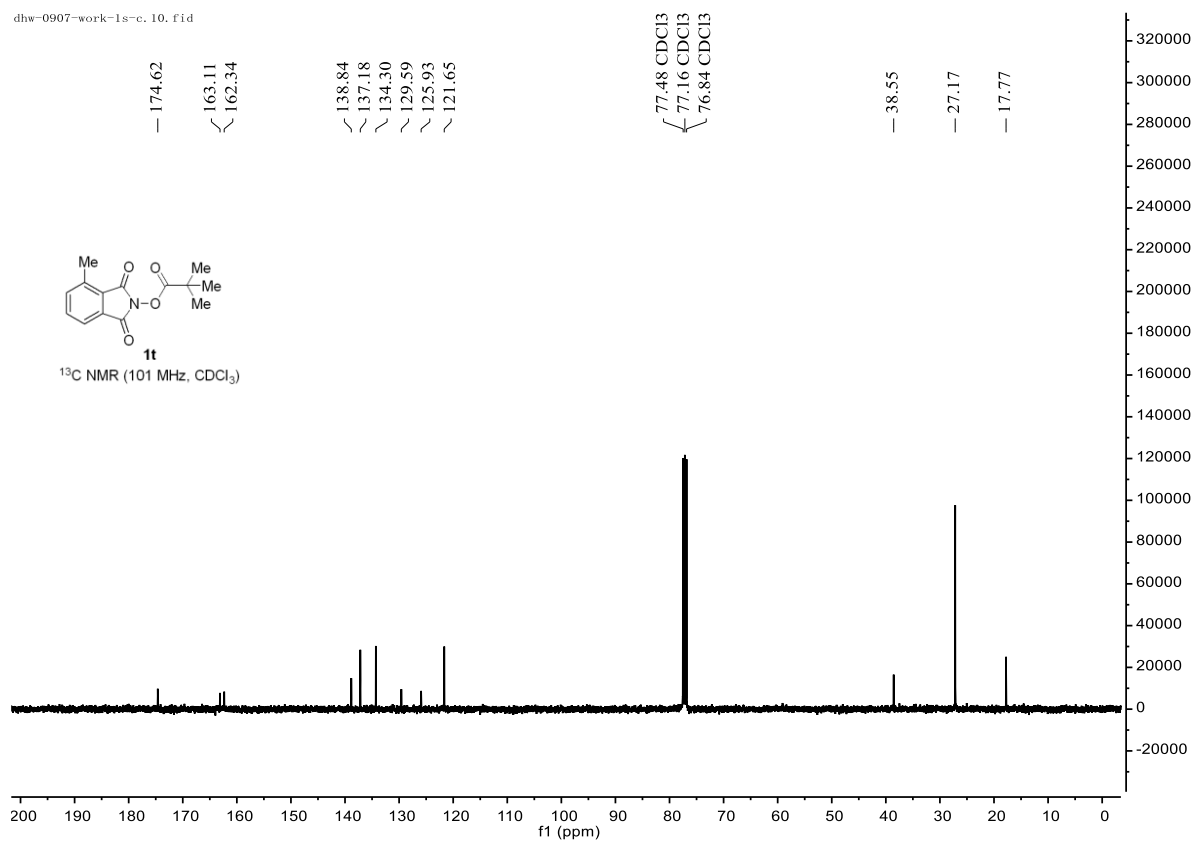
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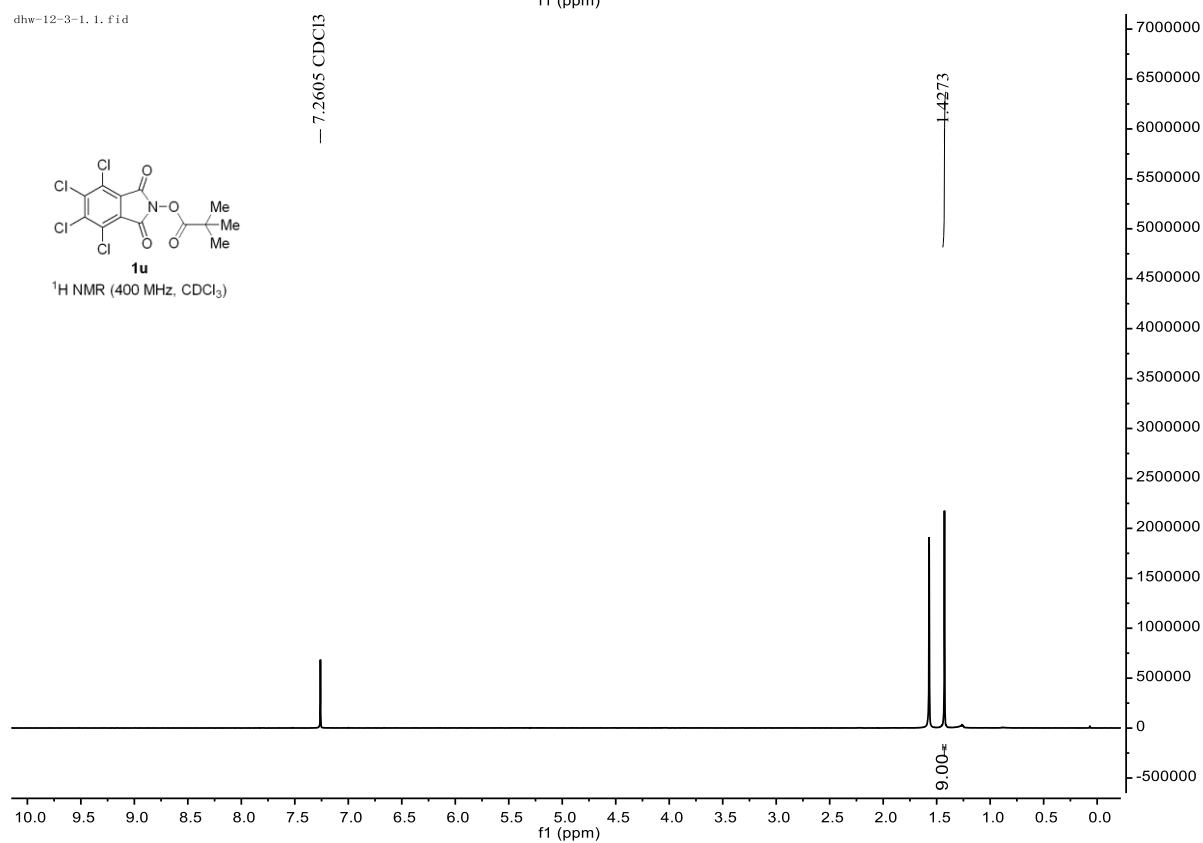
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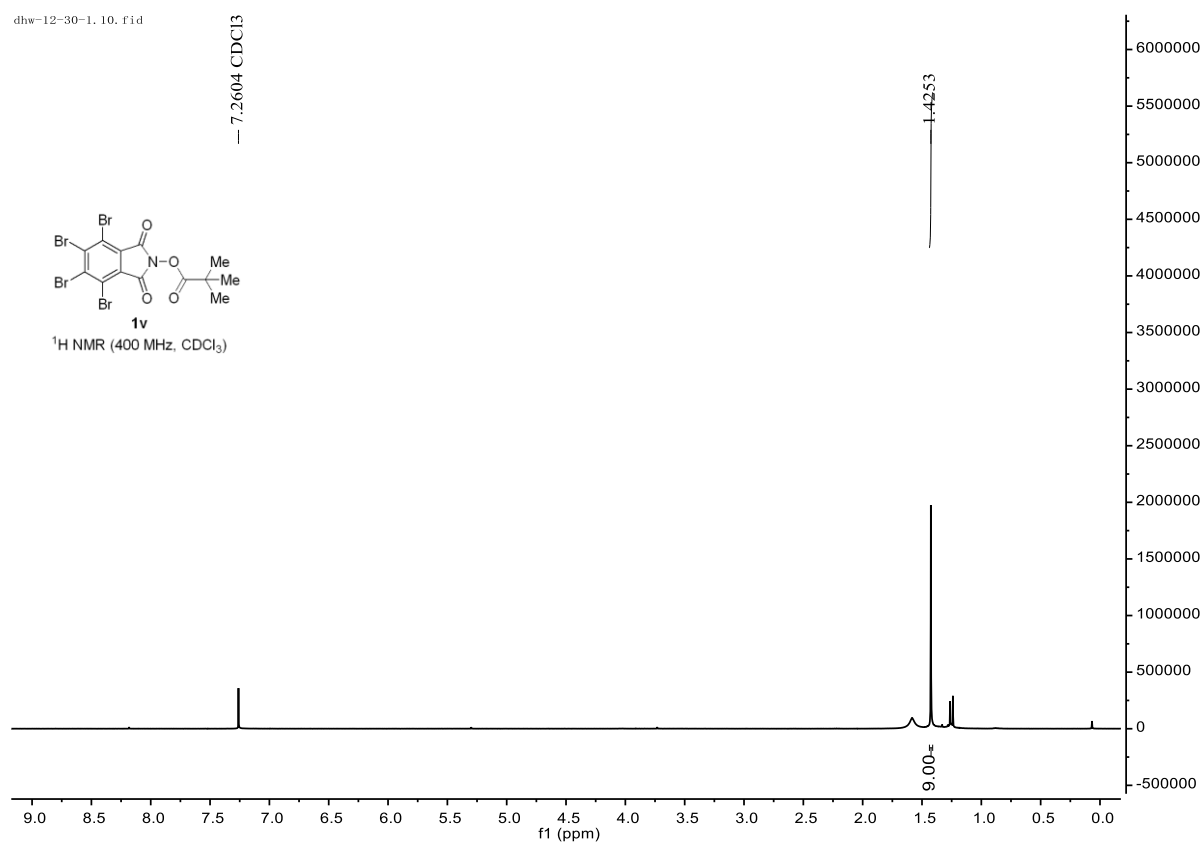
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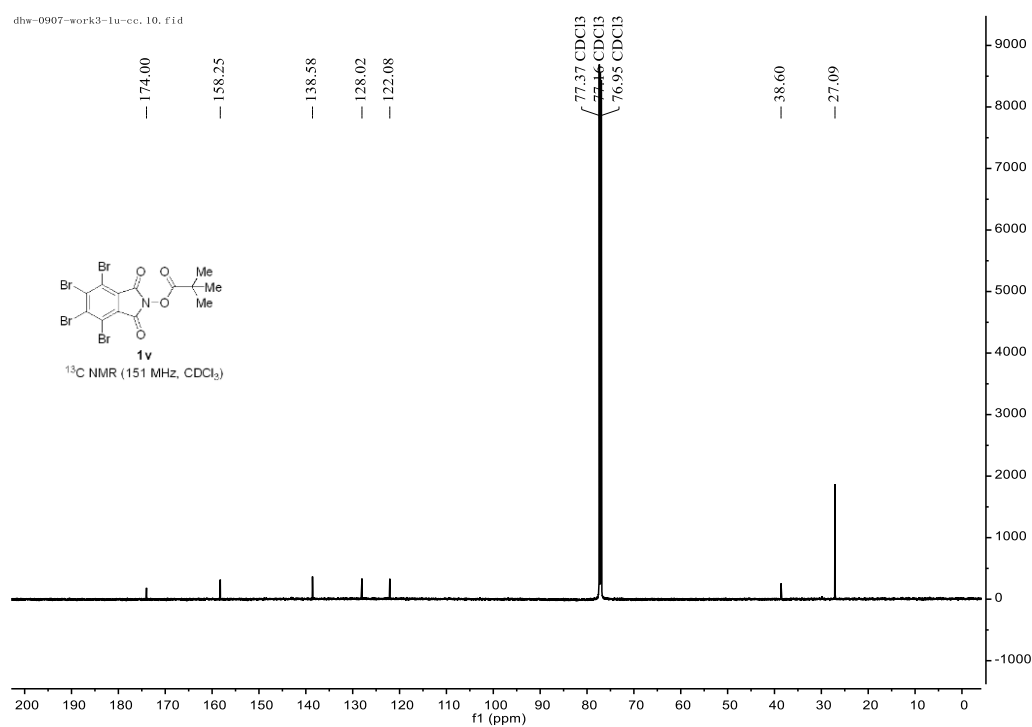
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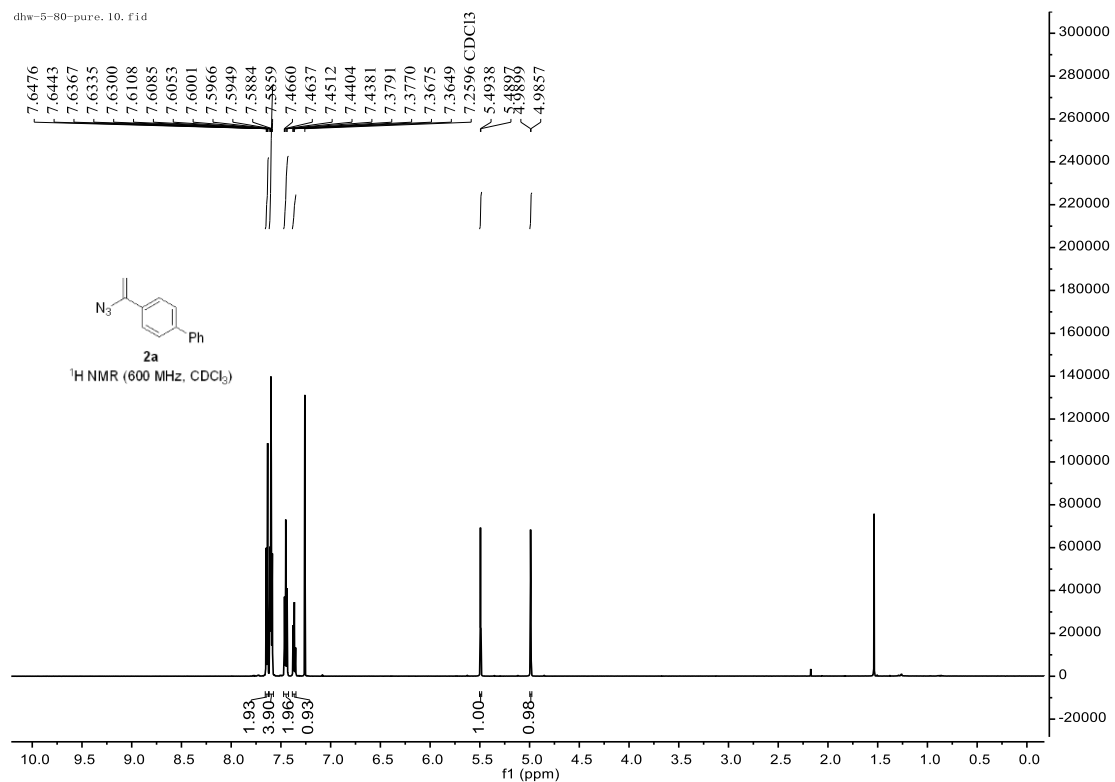
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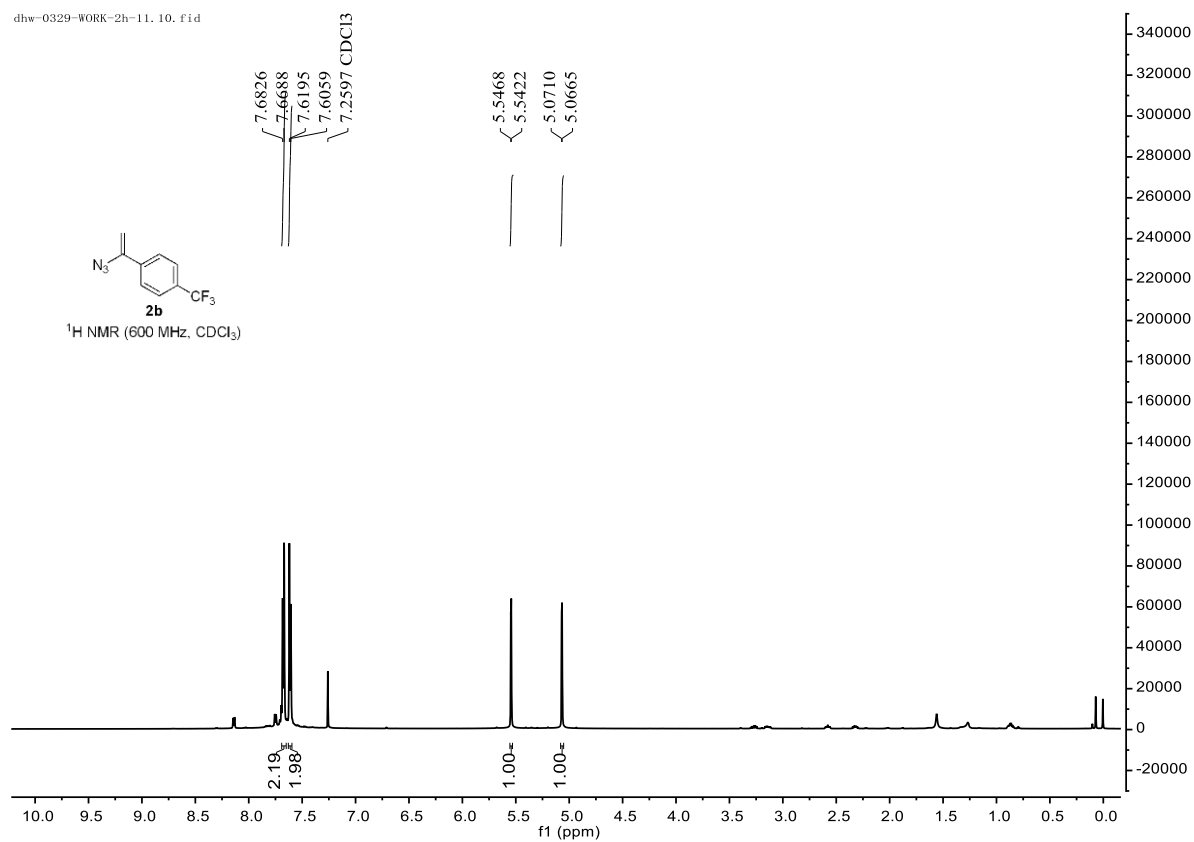
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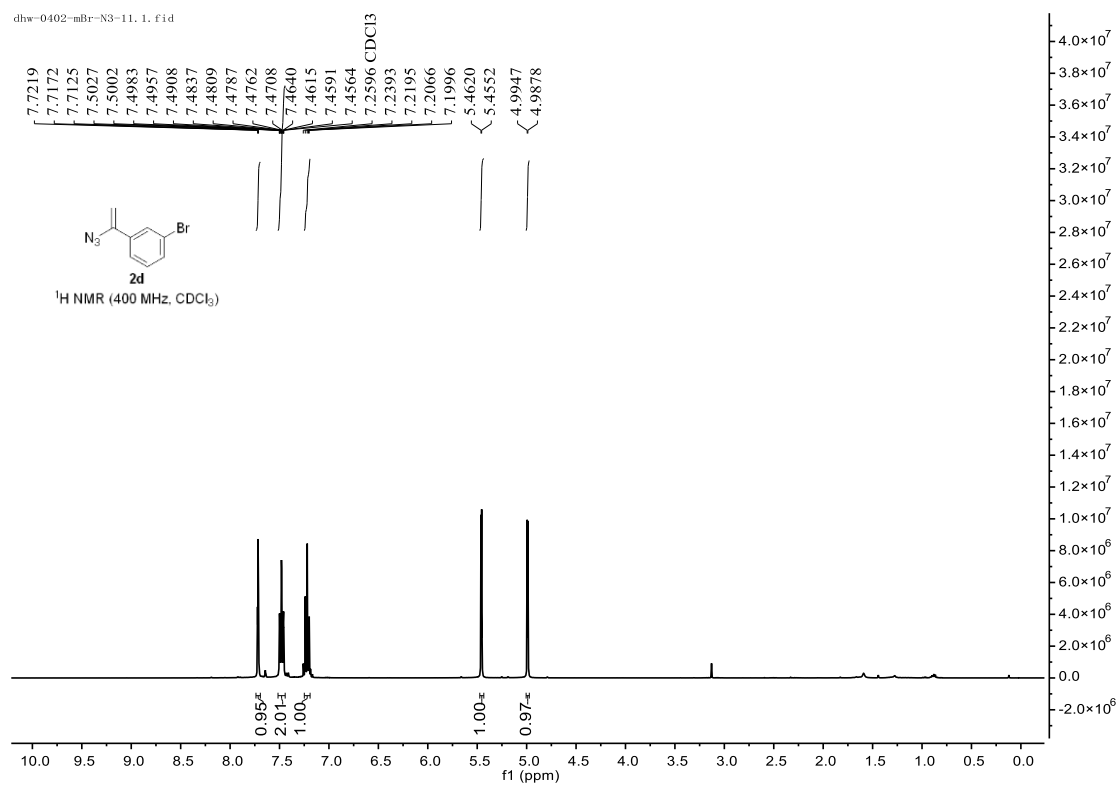
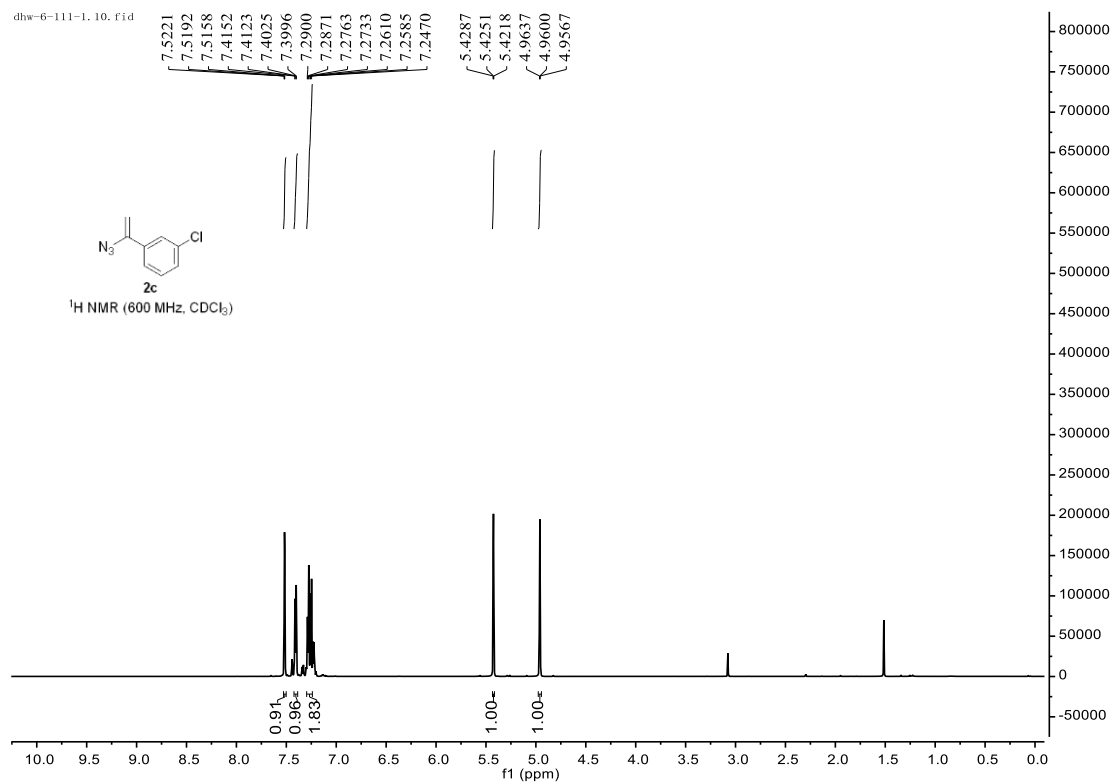


dhw-5-80-pure, 10, f1d

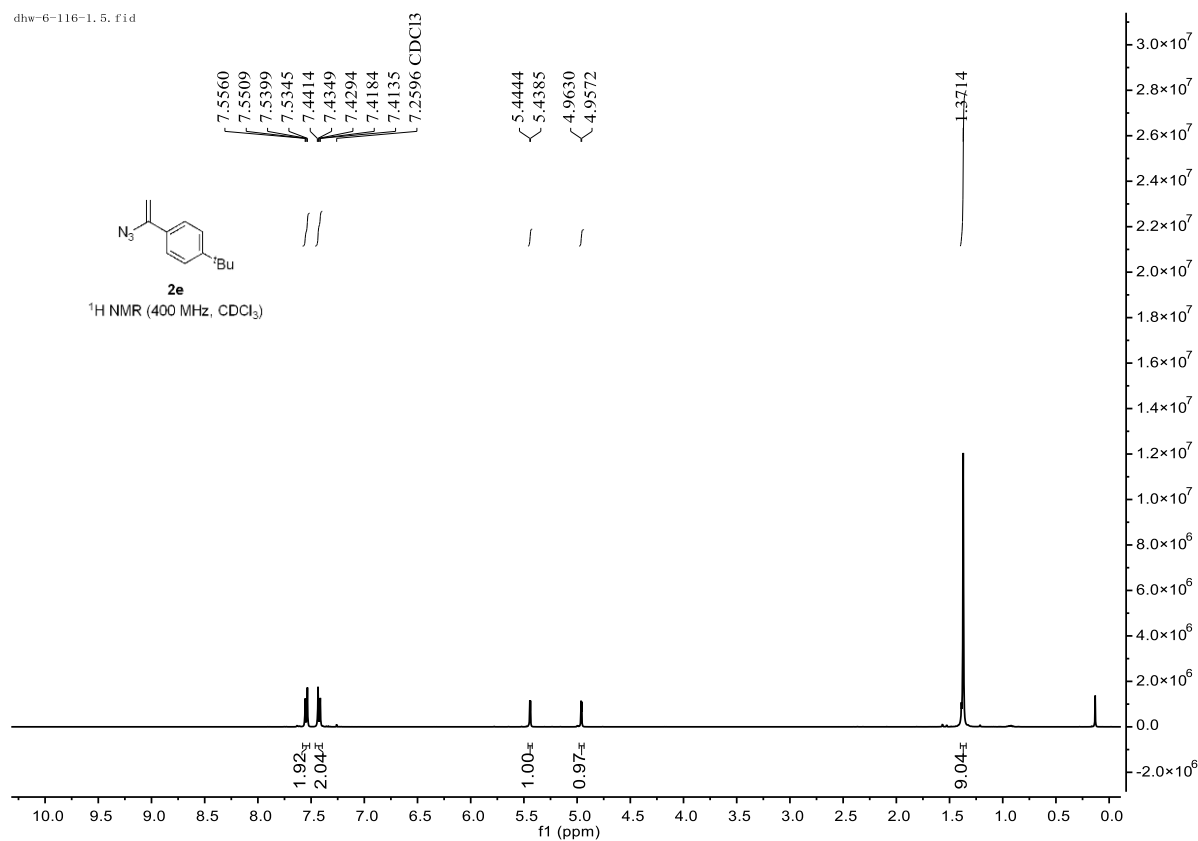


dhw-0329-WORK-2h-11, 10, f1d

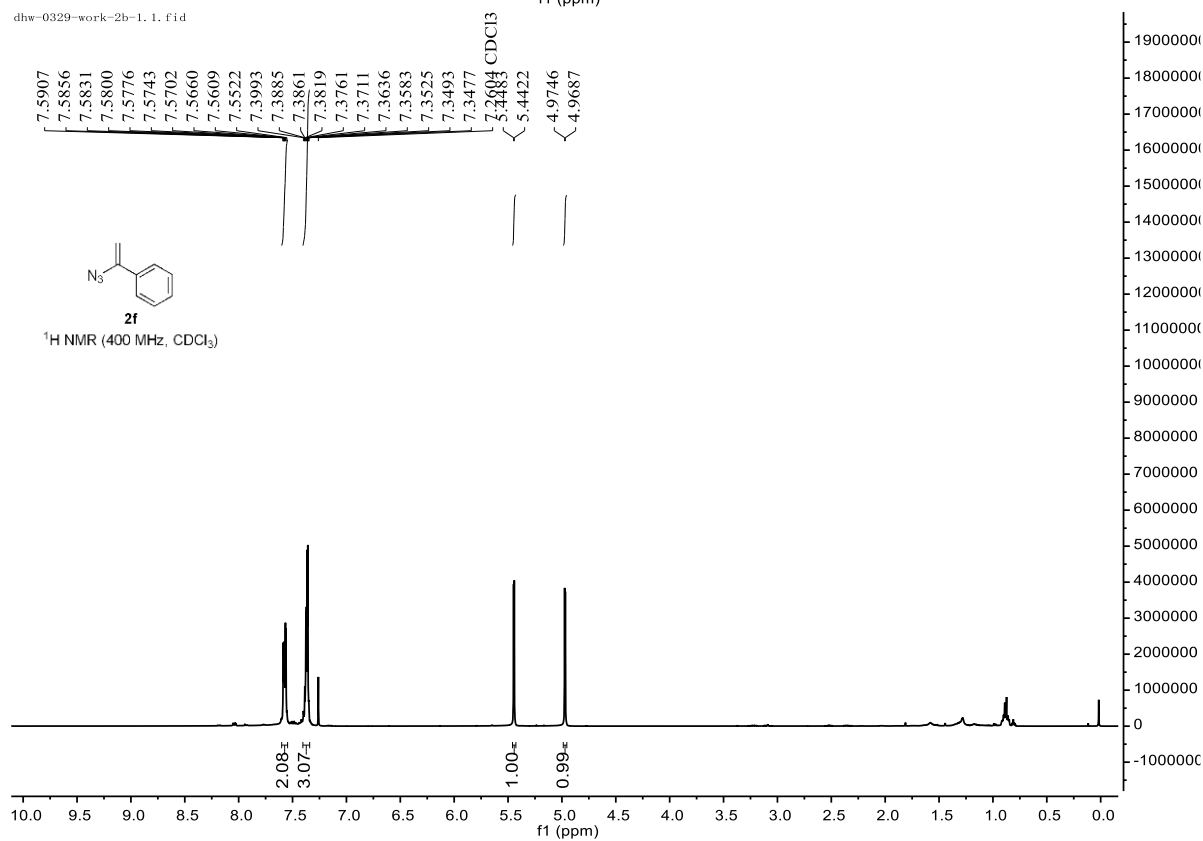




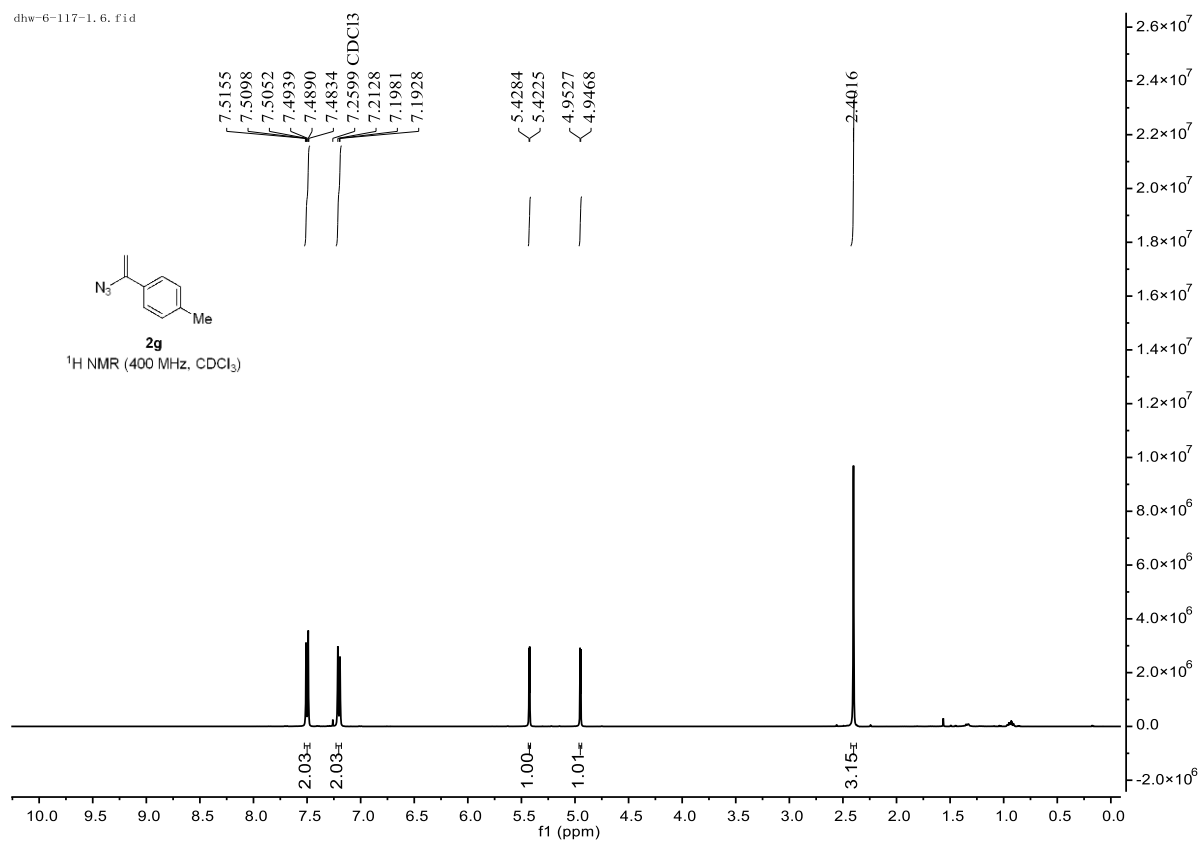
dhw-6-116-1.5.fid



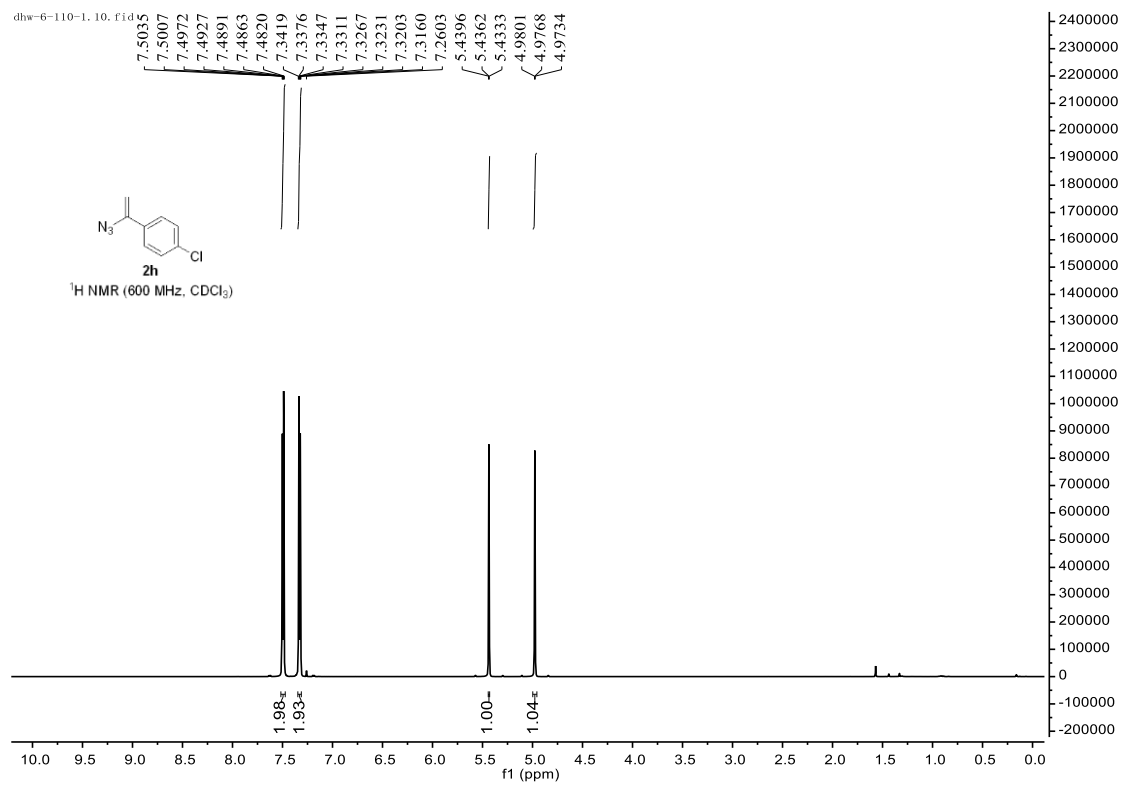
dhw-0329-work-2b-1.1.fid

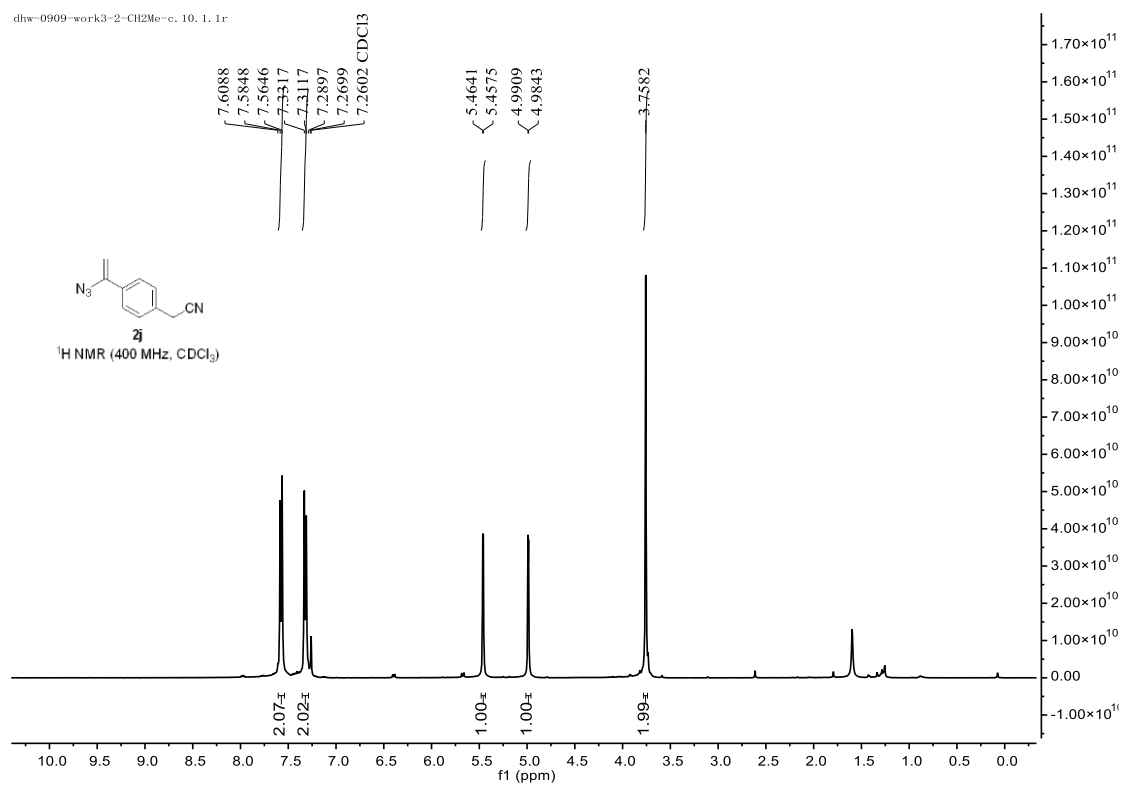
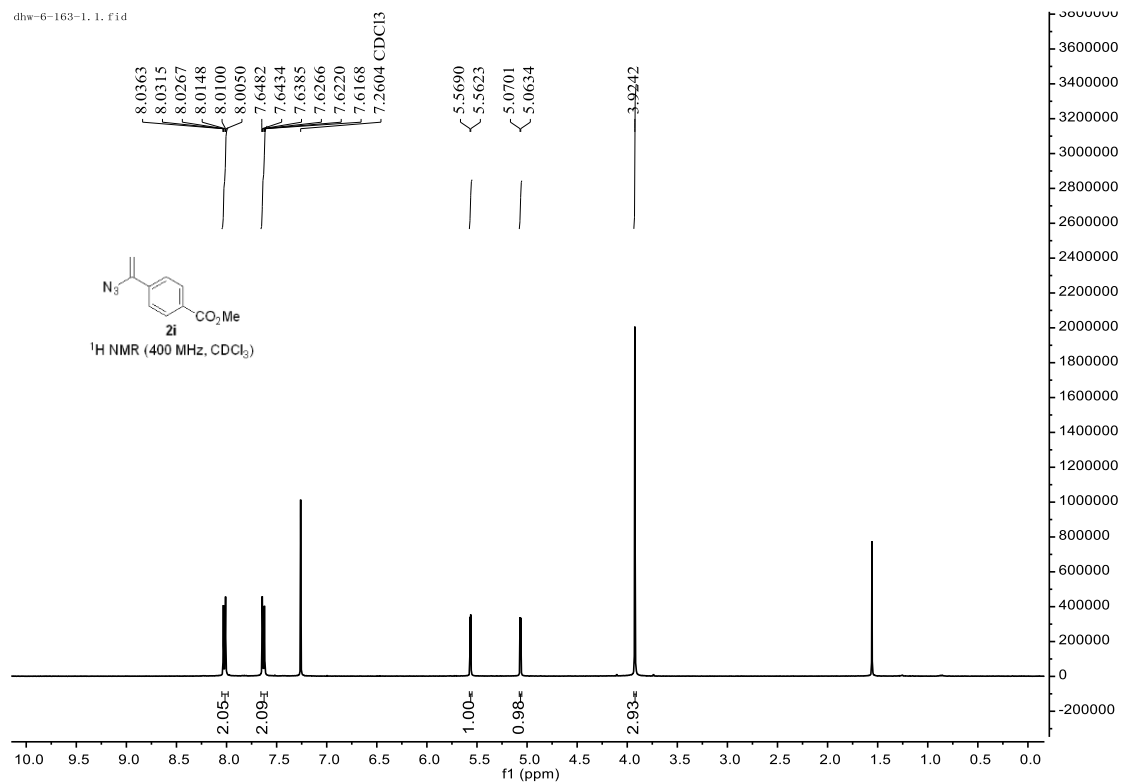


dhw-6-117-1.6.fid

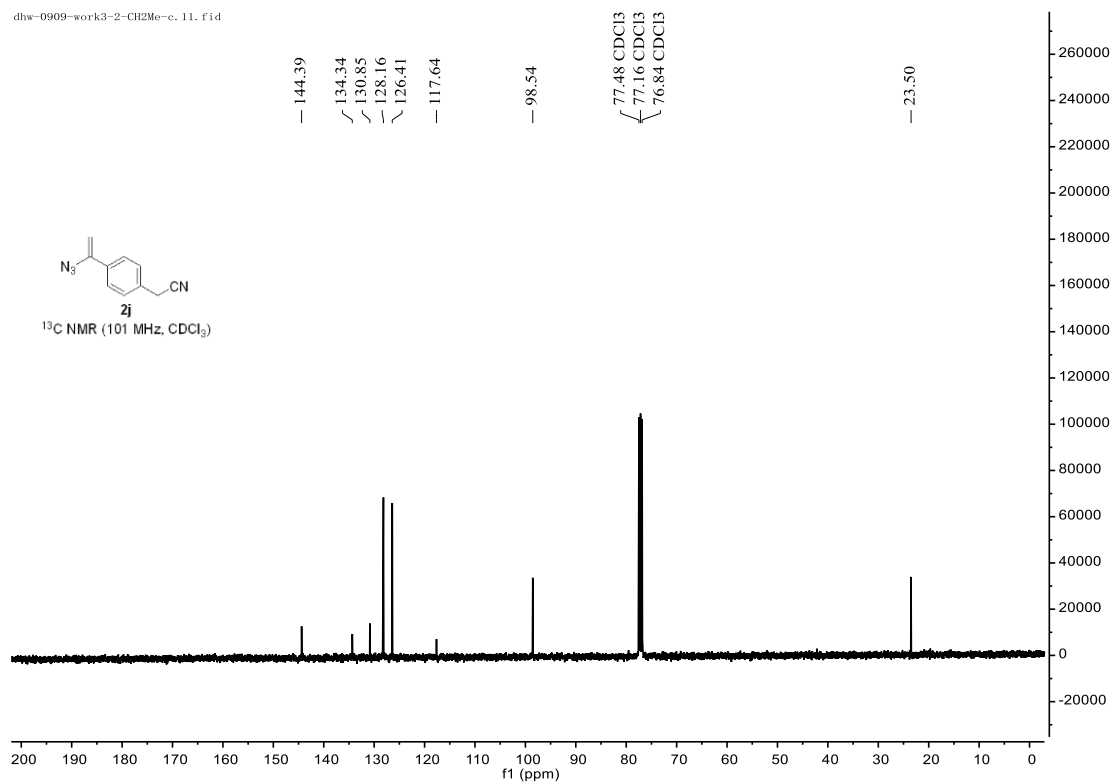


dhw-6-110-1.10.fid

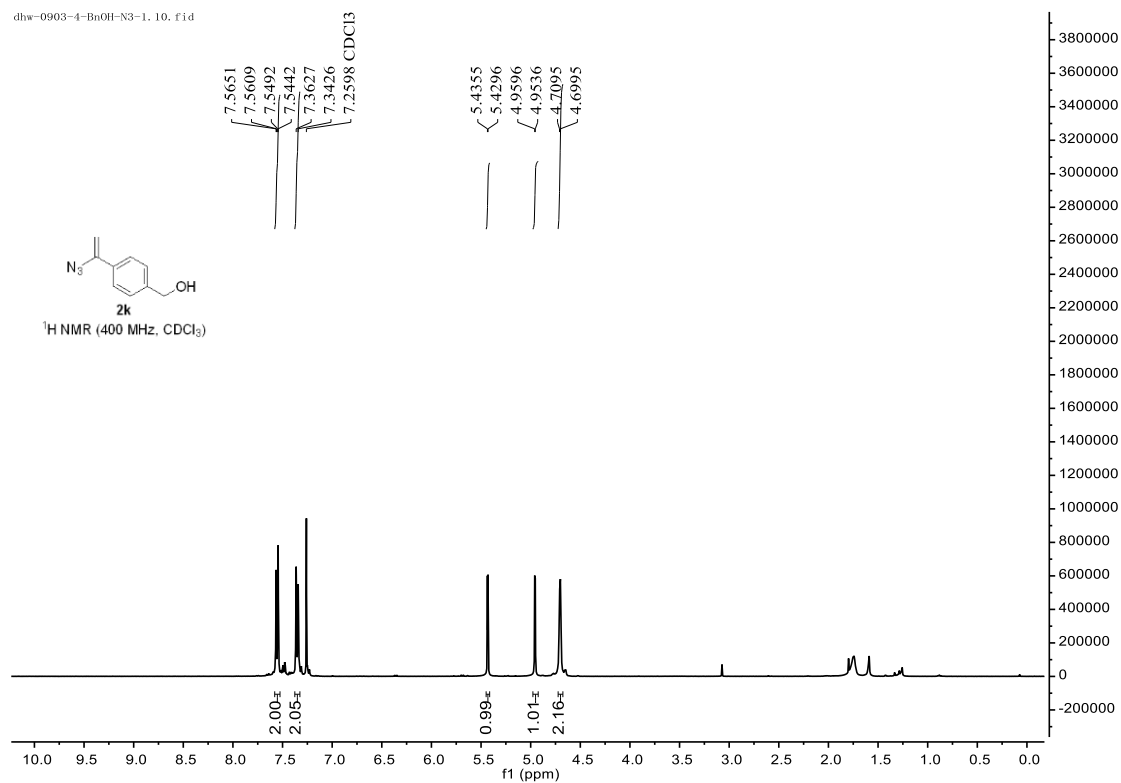


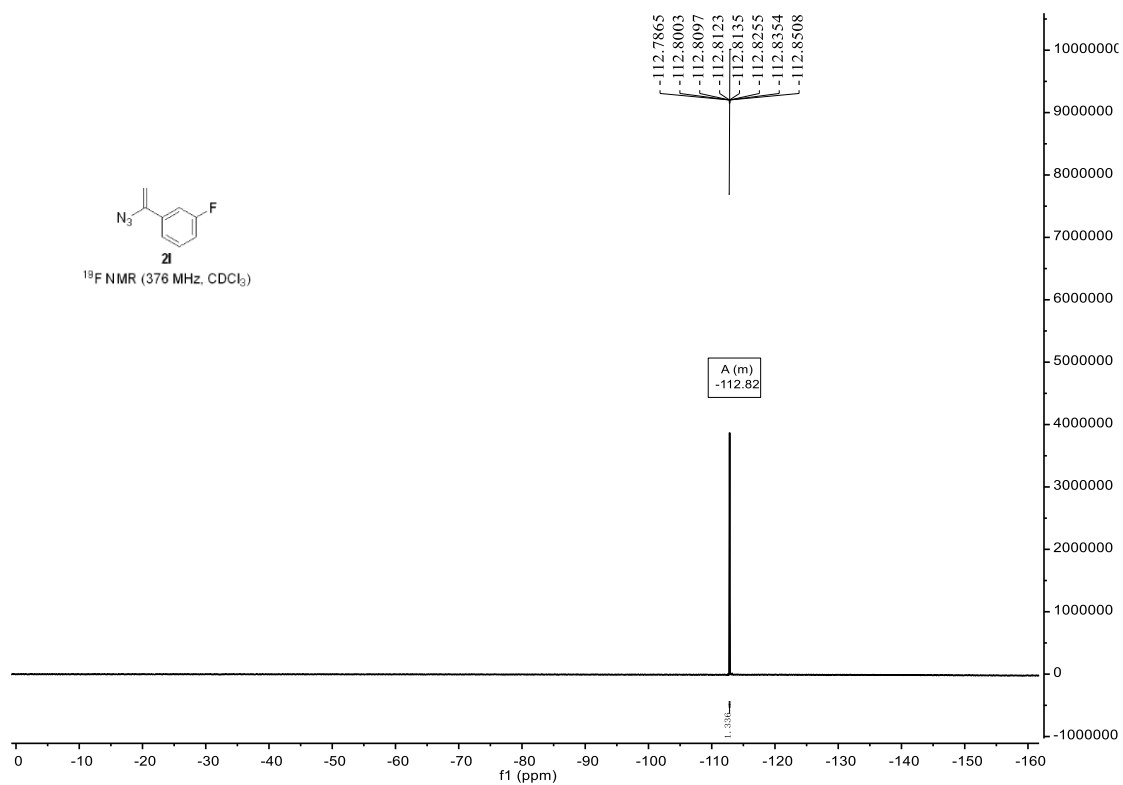
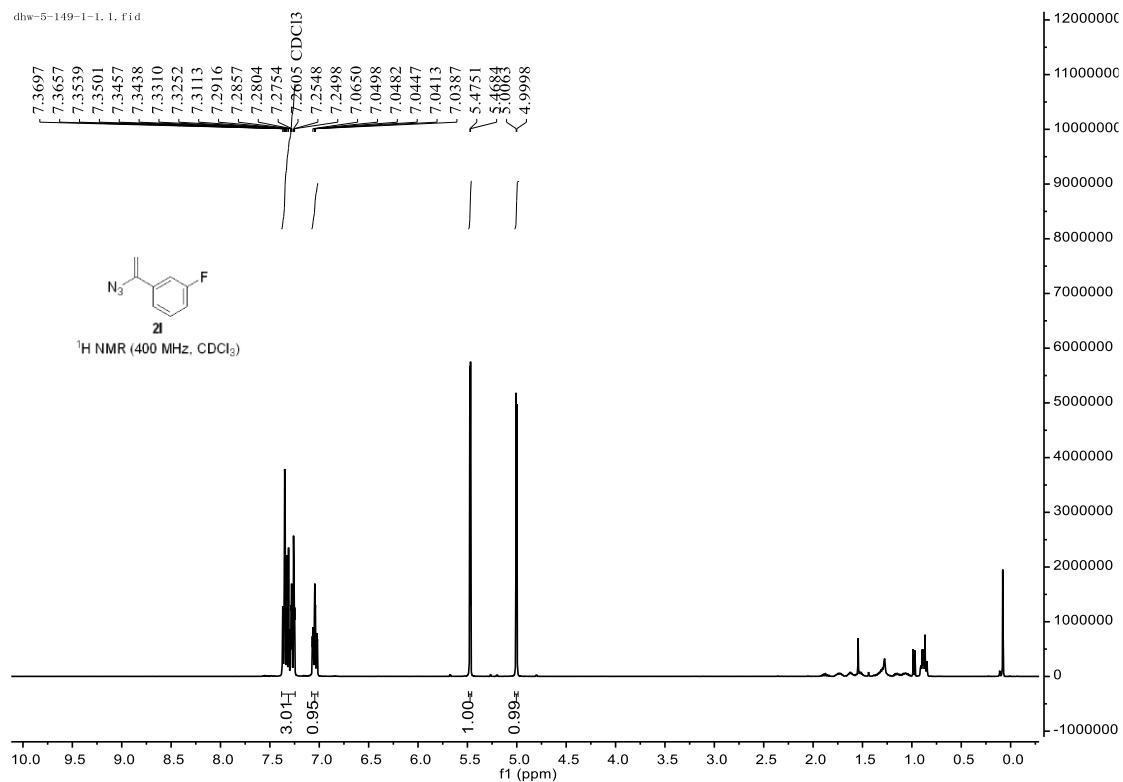


dhw-0909-work3-2-CH2Me-c, 11, f1d

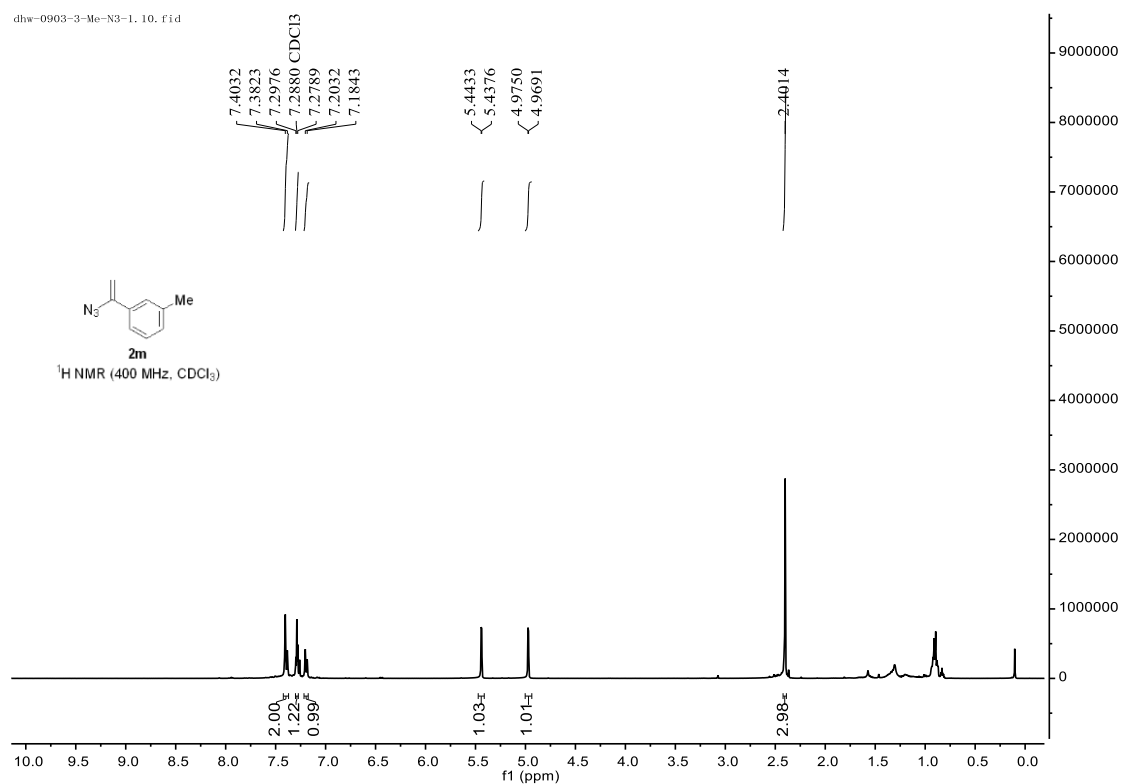


dhw-0903-4-BnOH-N3-1, 10, f1d

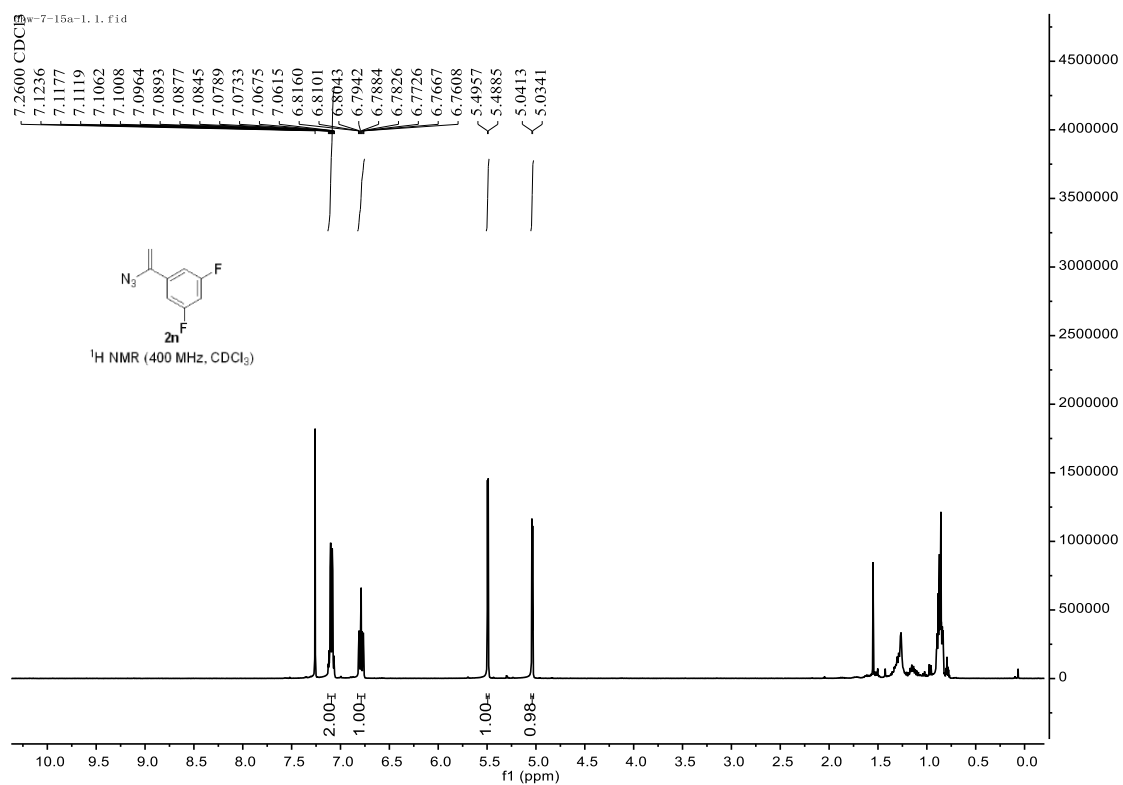




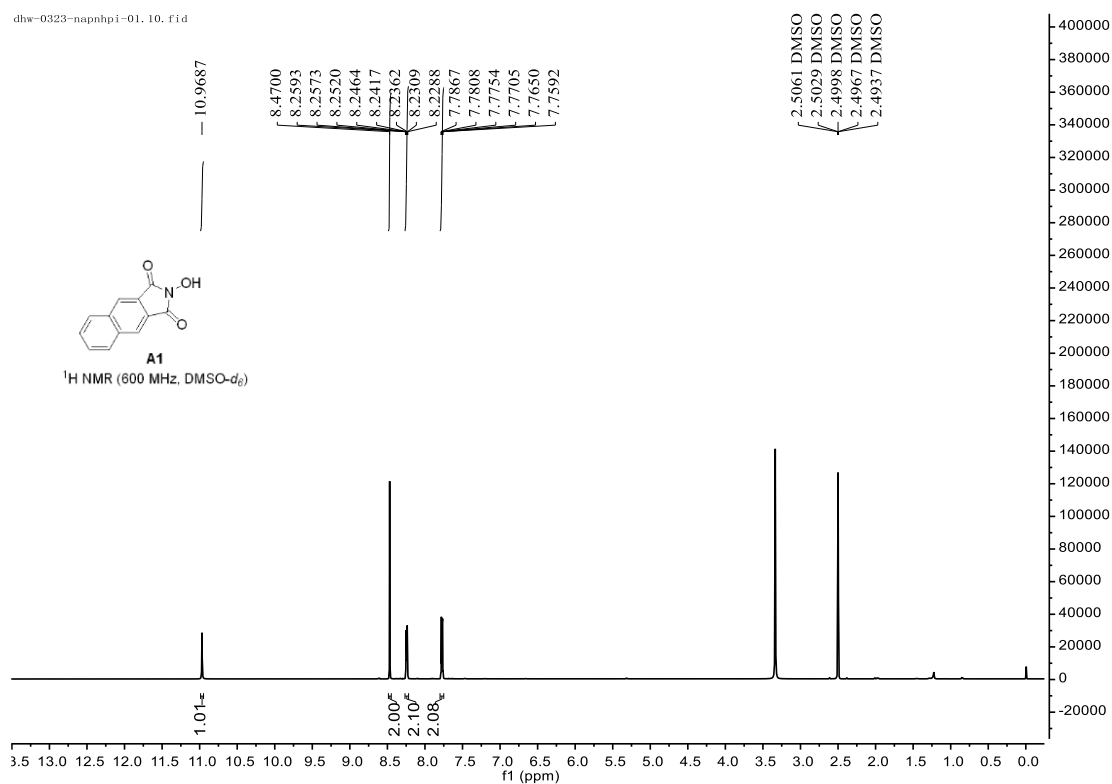
dhw-0903-3-Me-N3-1, 10, f1d



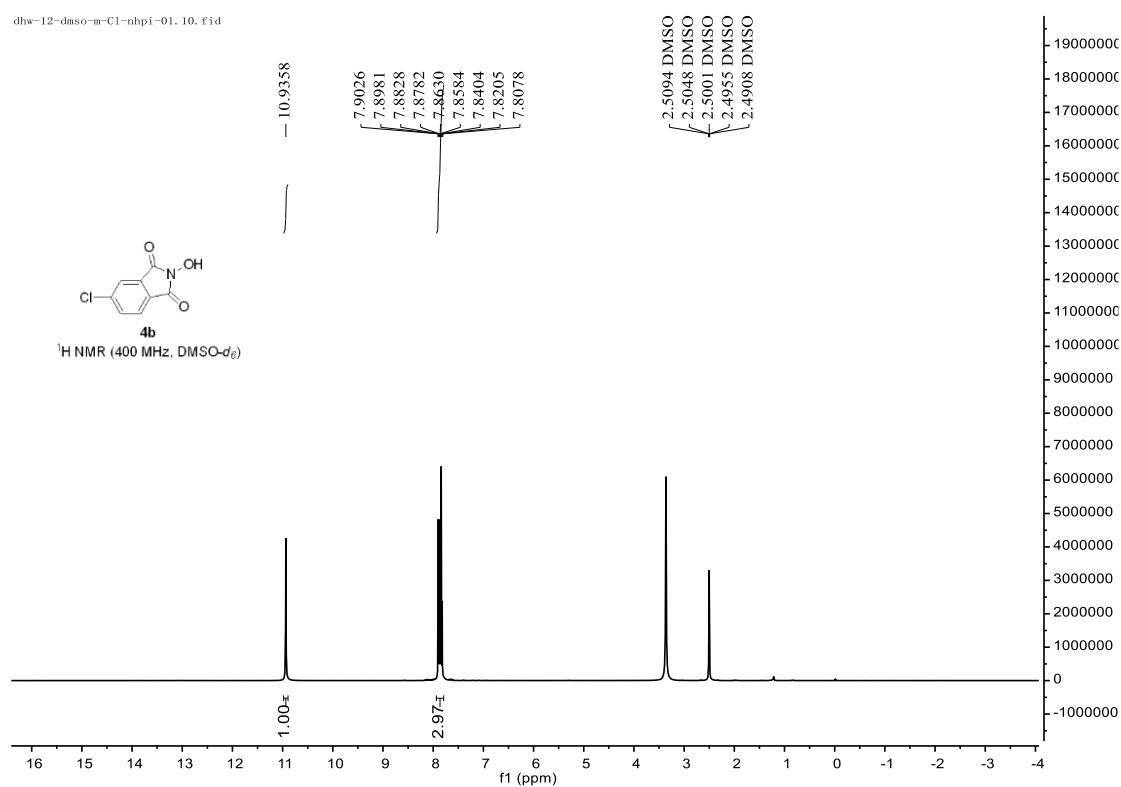
dhw-7-15a-1, 1, f1d



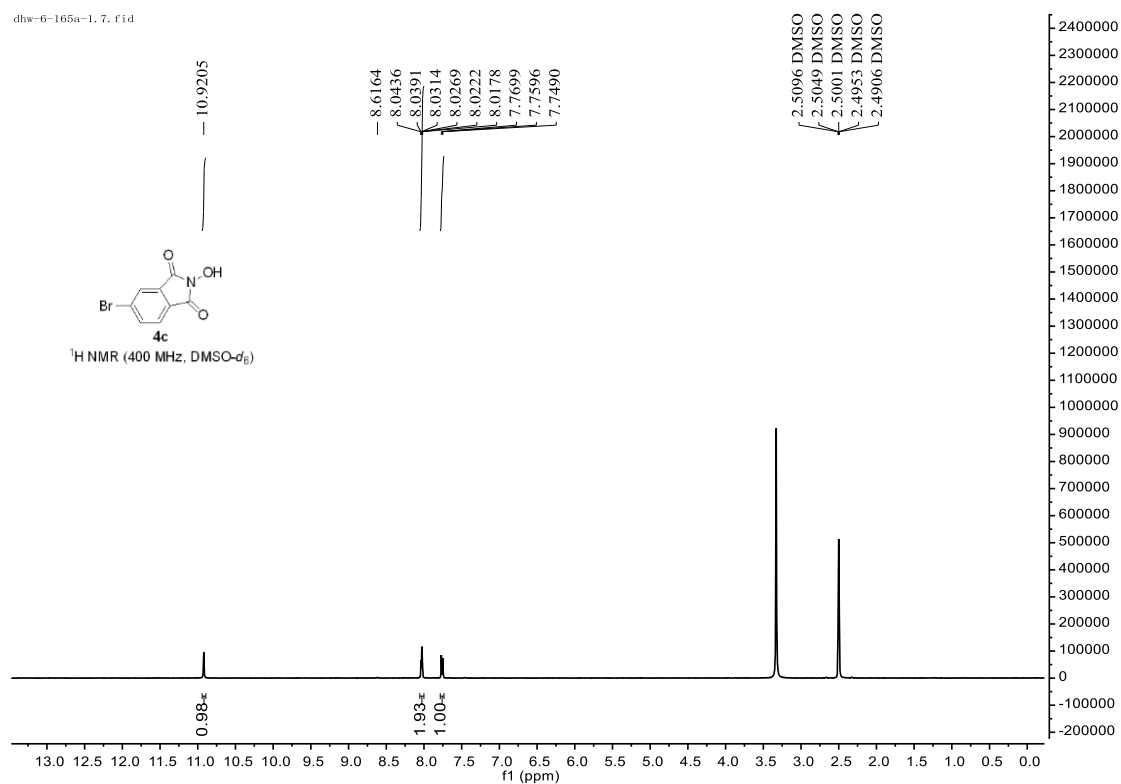
dhw-0323-napnhipi-01.10.fid



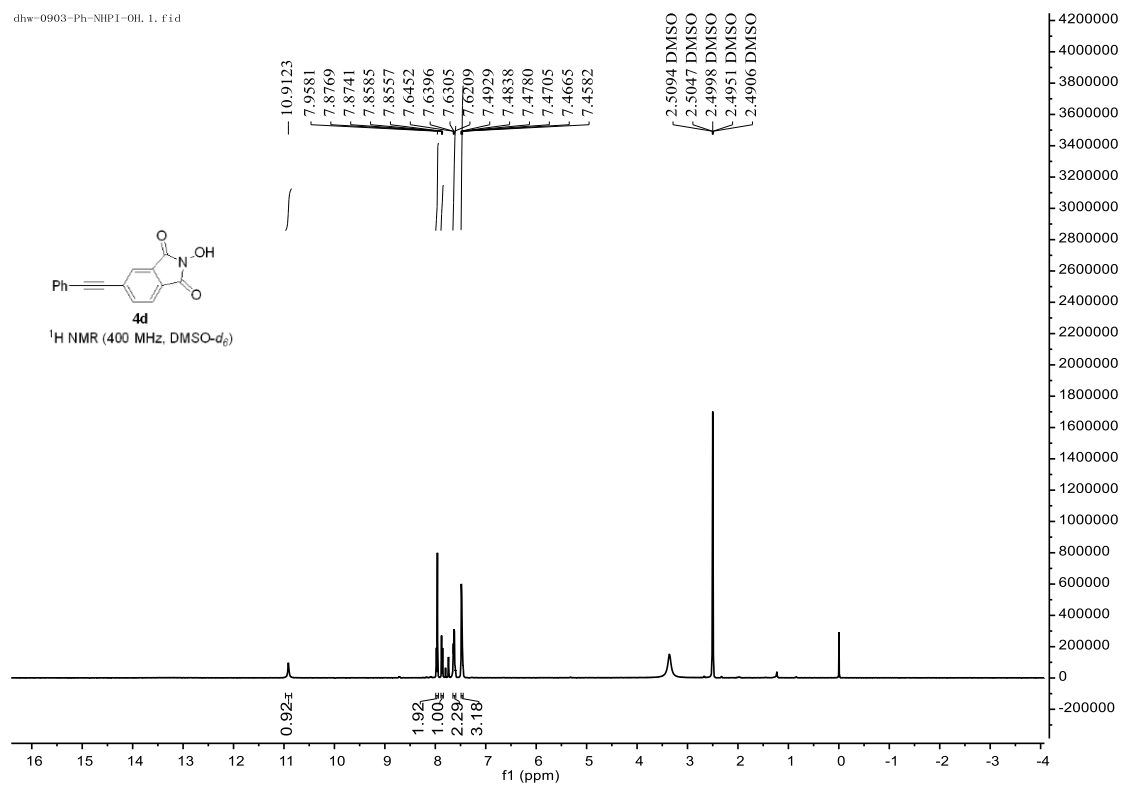
dhw-12-dms0-m-Cl-nhipi-01.10.fid



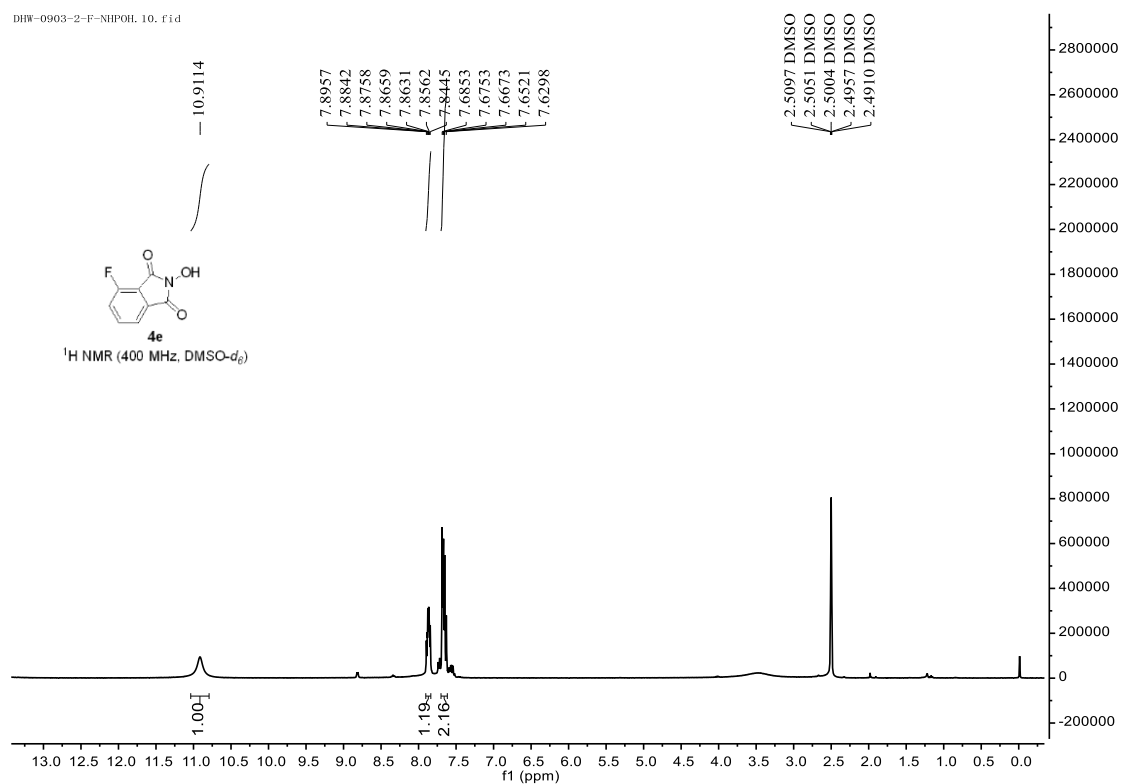
dhw-6-165a-1.7.fid



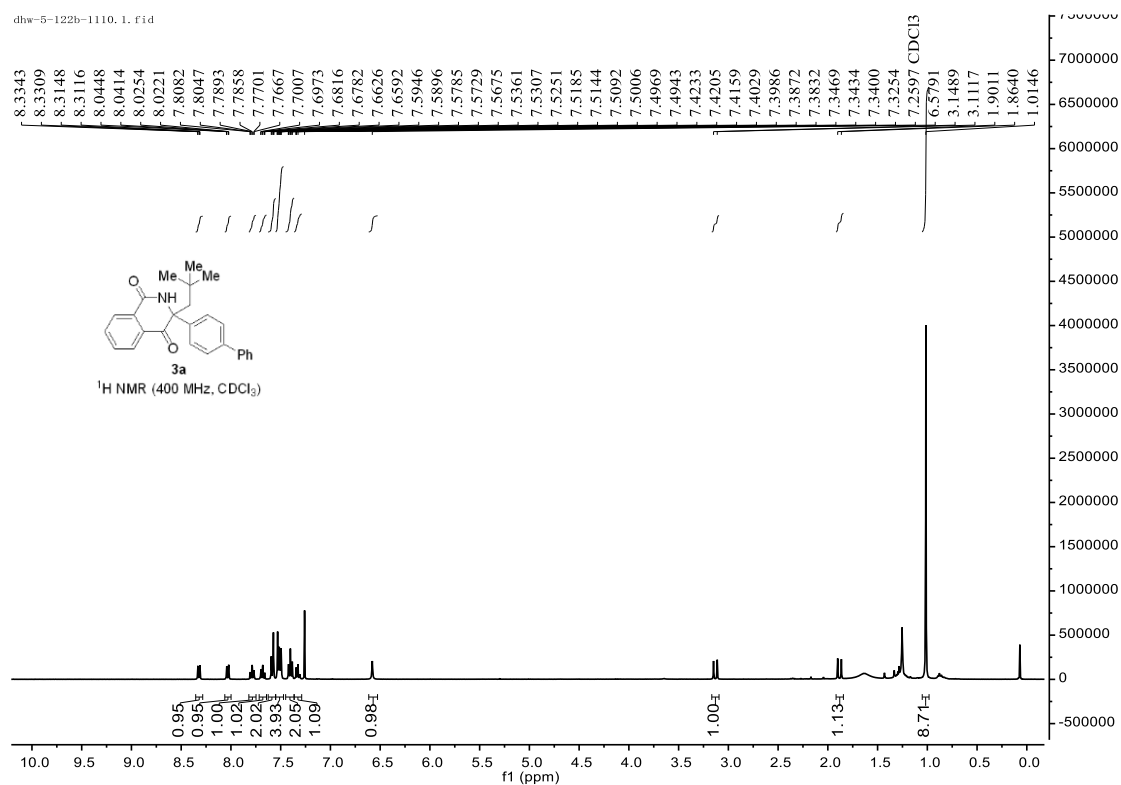
dhw-0903-Ph-NHPI-OH.1.fid



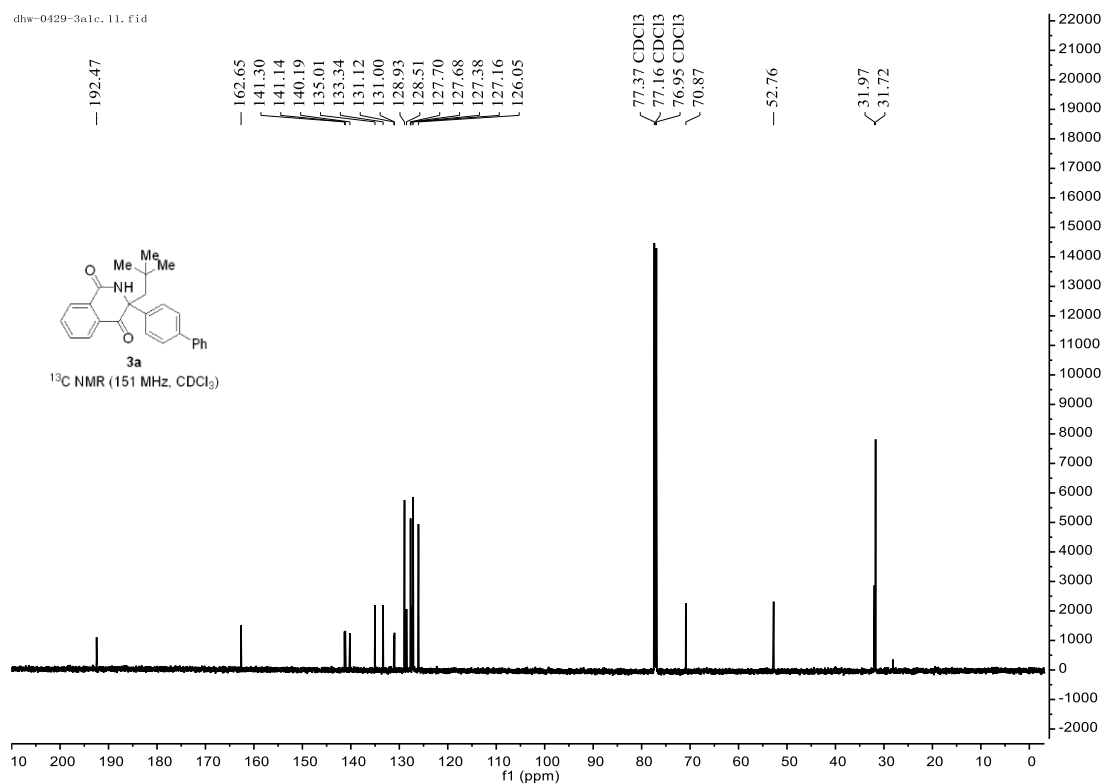
dhw-0903-2-F-NHPOH, 10, f1d



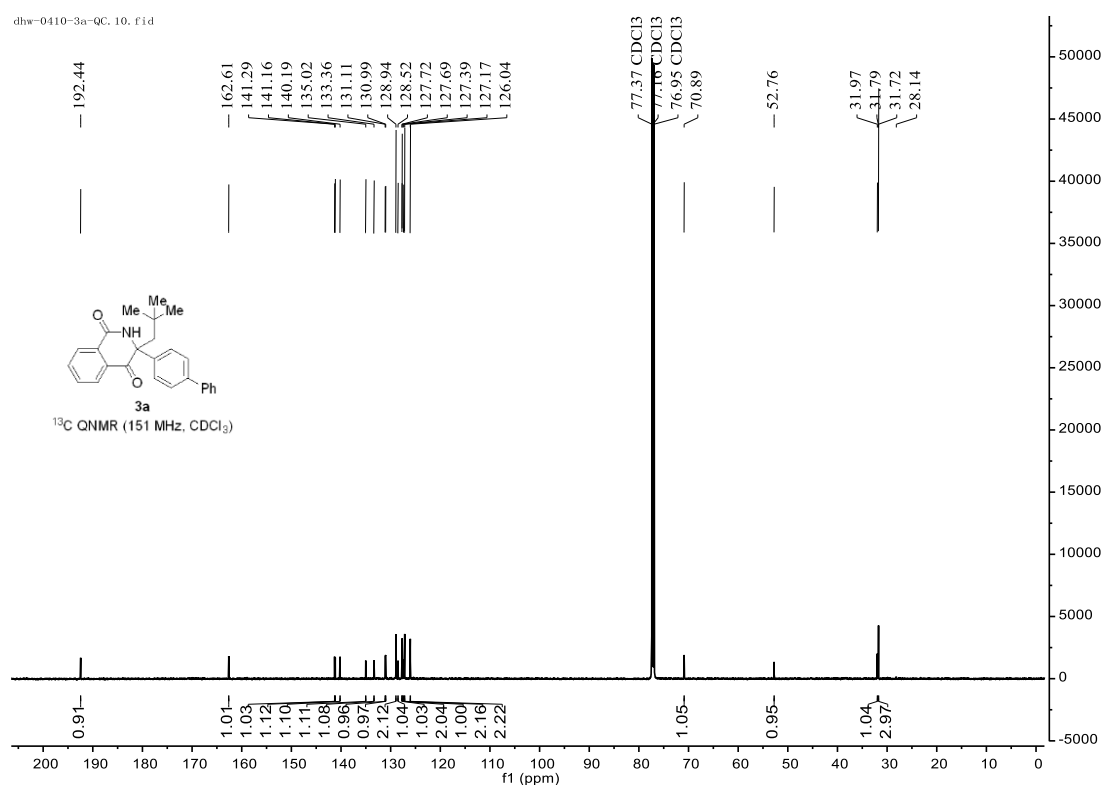
dhw-5-122b-1110, 1, f1d

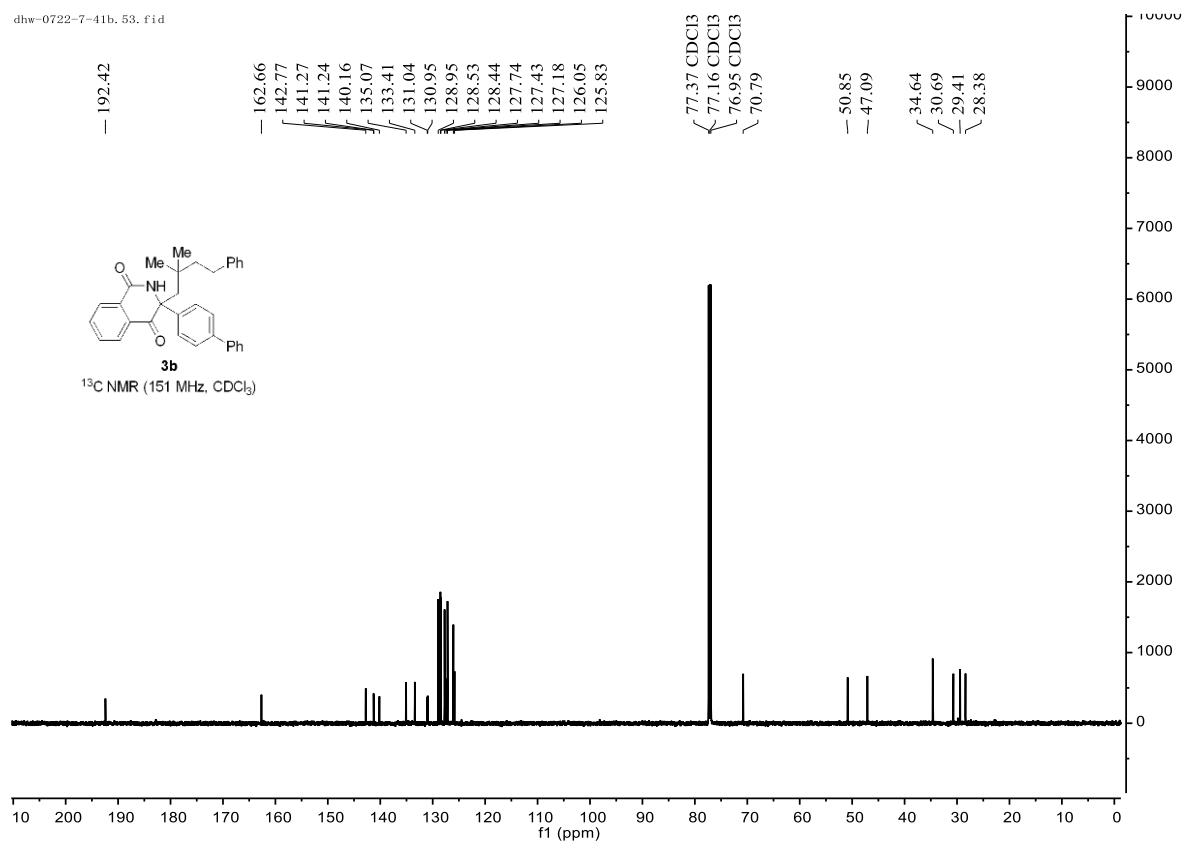
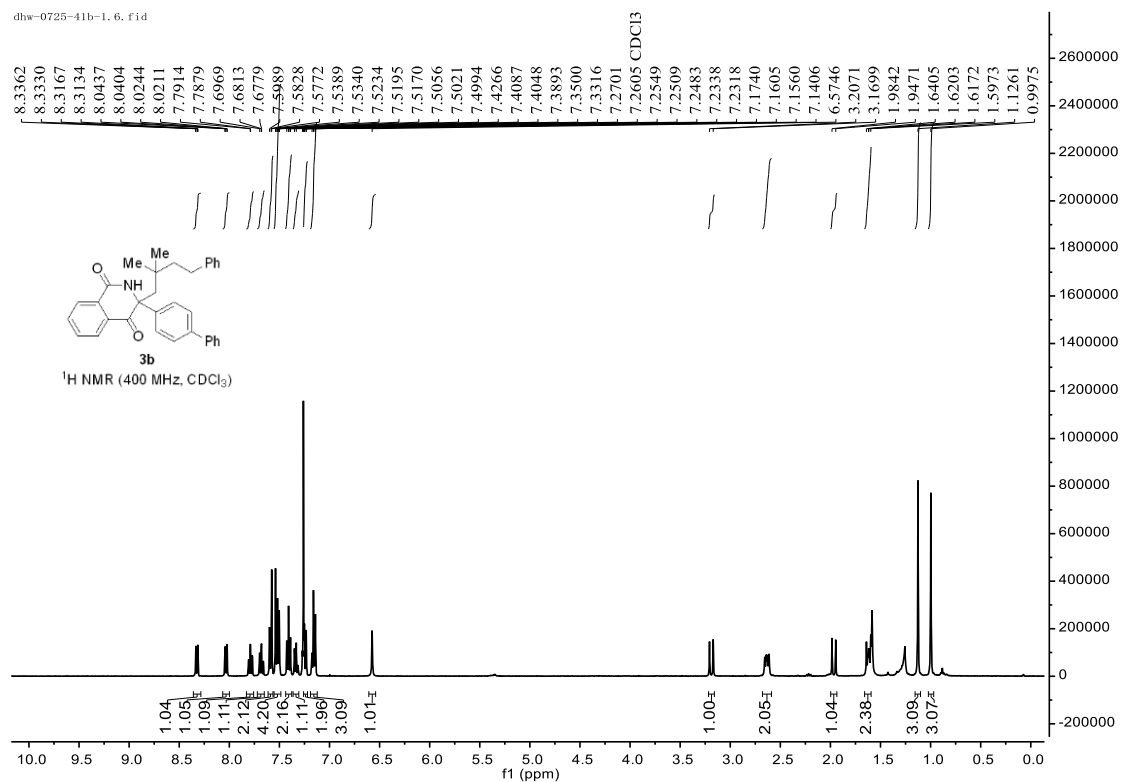


dhw-0429-3a1c, 11, fid

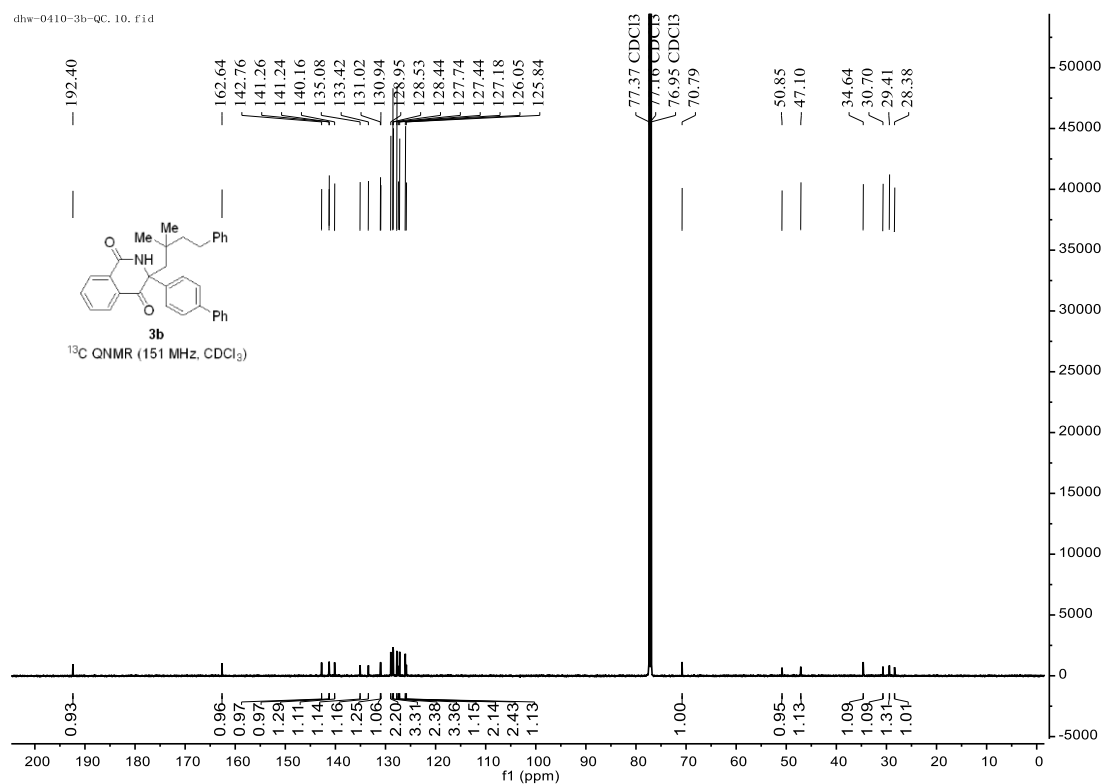


dhw-0410-3a-QC, 10, fid

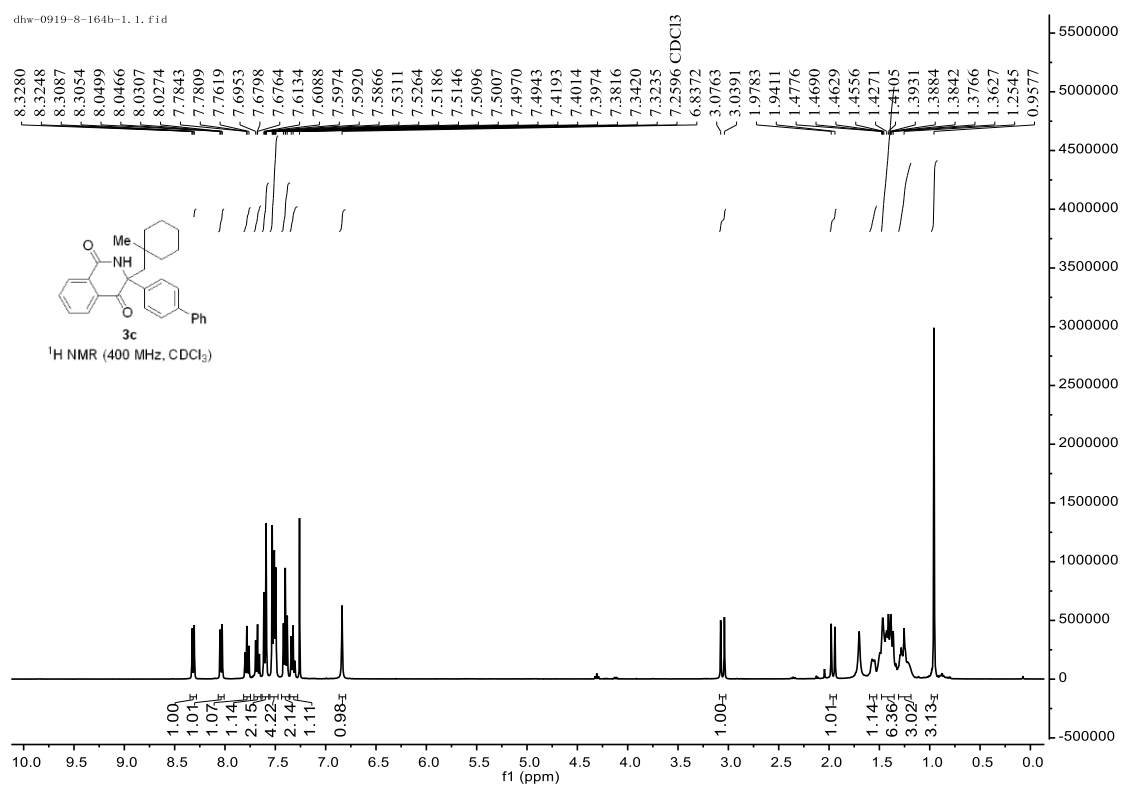




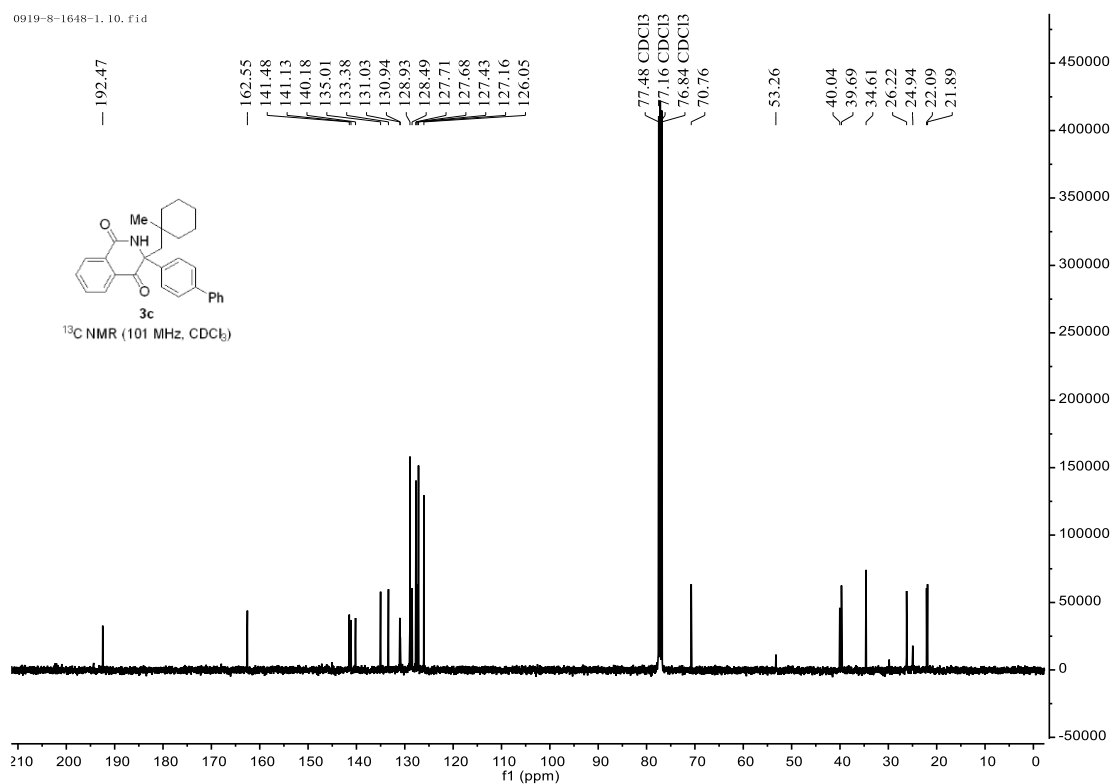
dhw-0410-3b-QC, 10, f1d



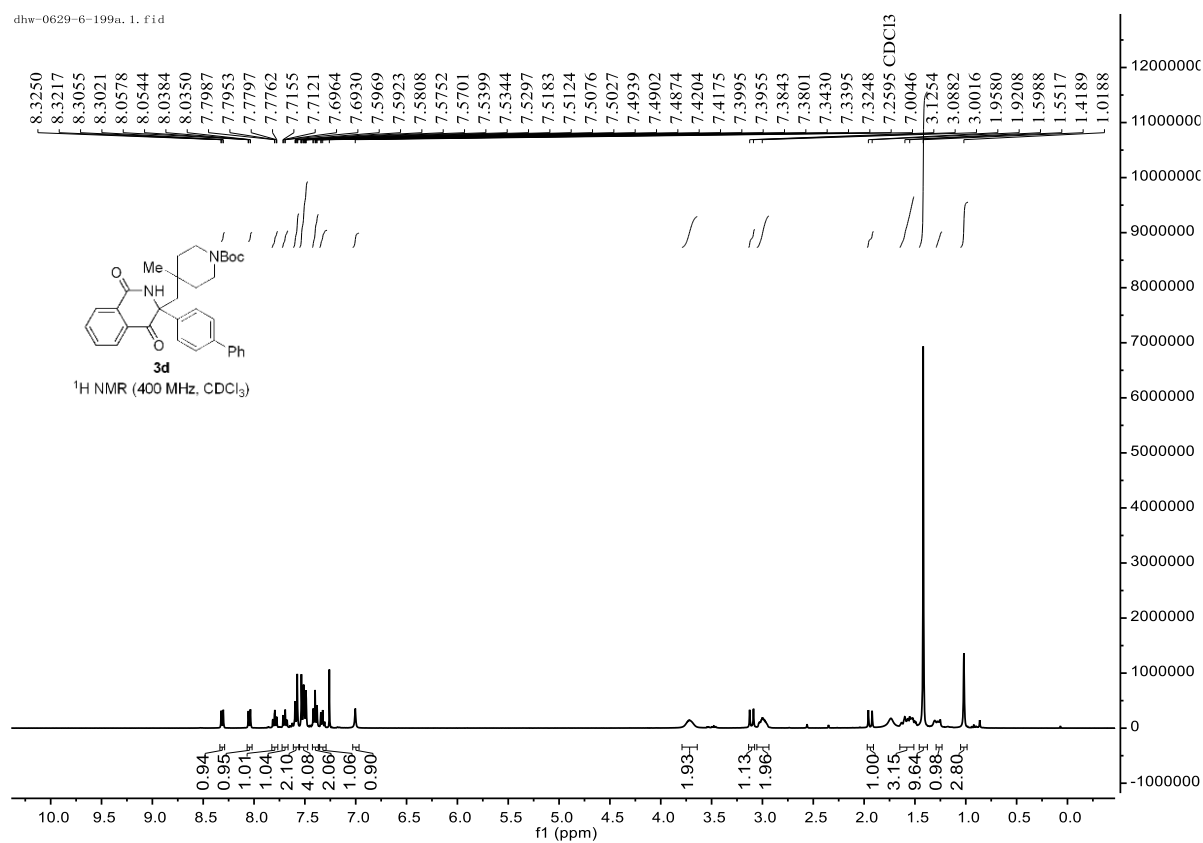
dhw-0919-8-164b-1, 1, f1d



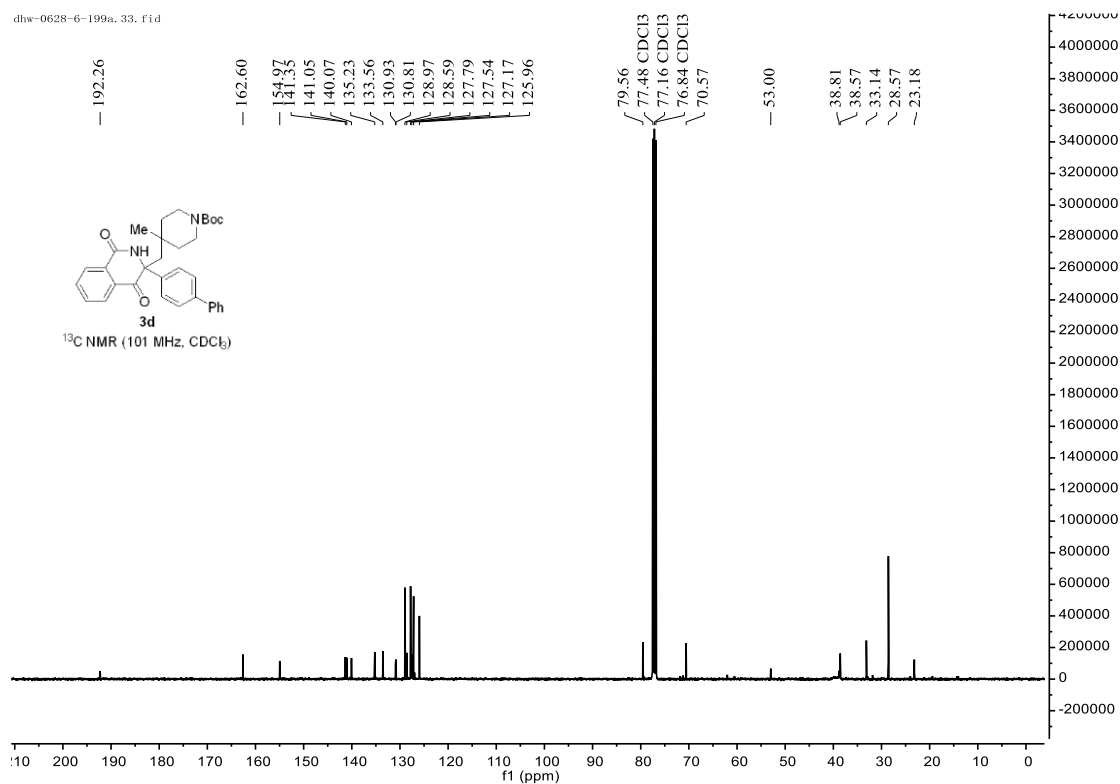
0919-8-1648-1.10. f1d



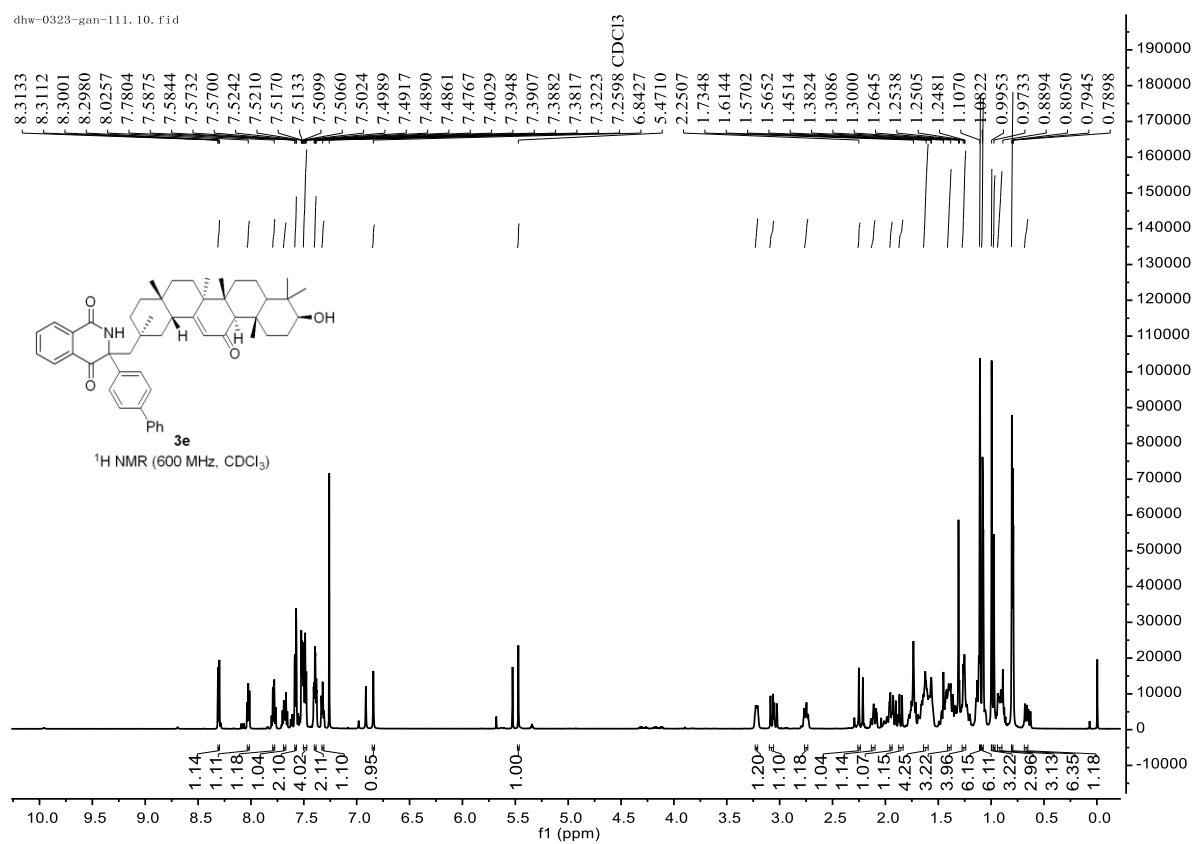
dhw-0629-6-199a.1. f1d

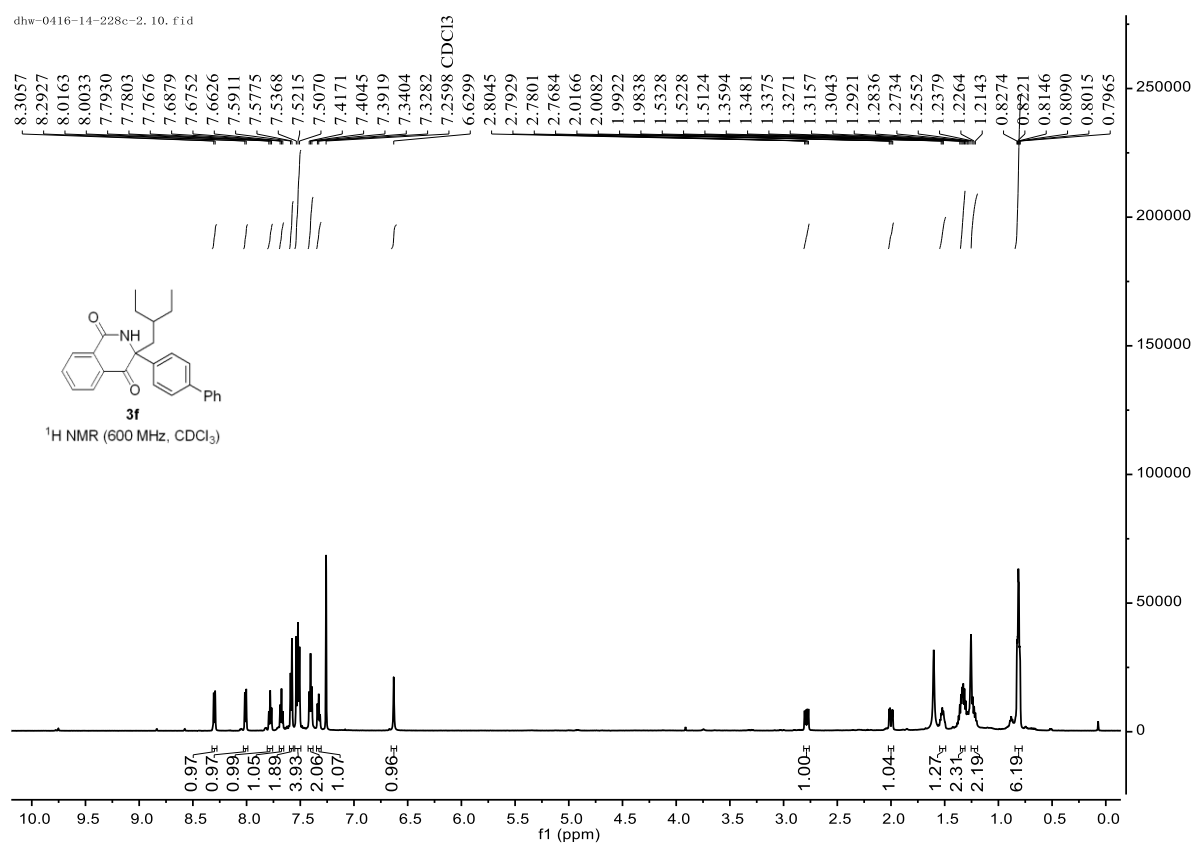
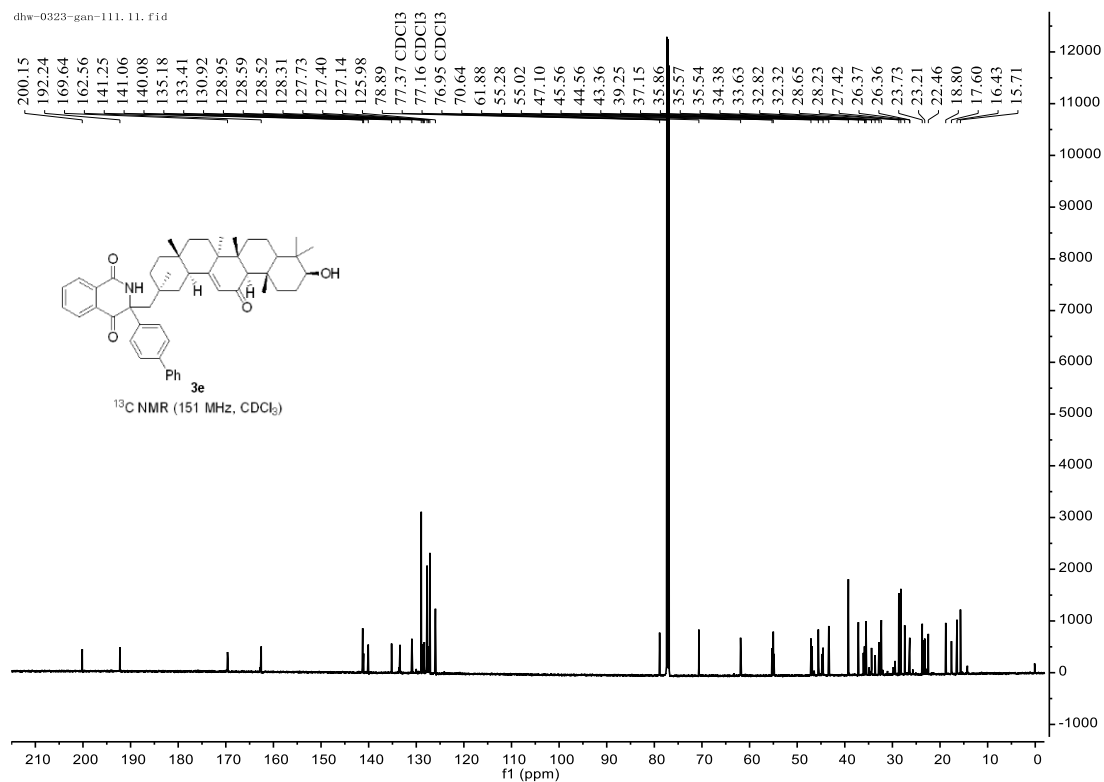


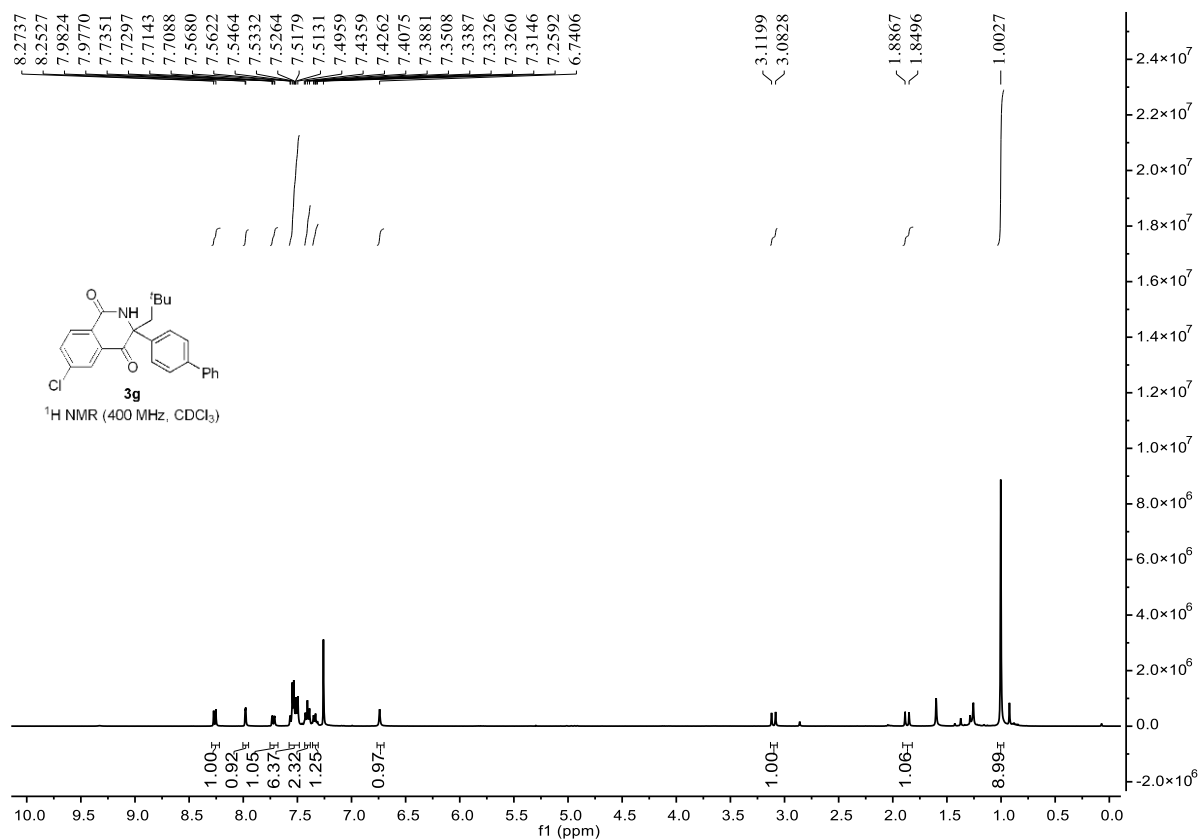
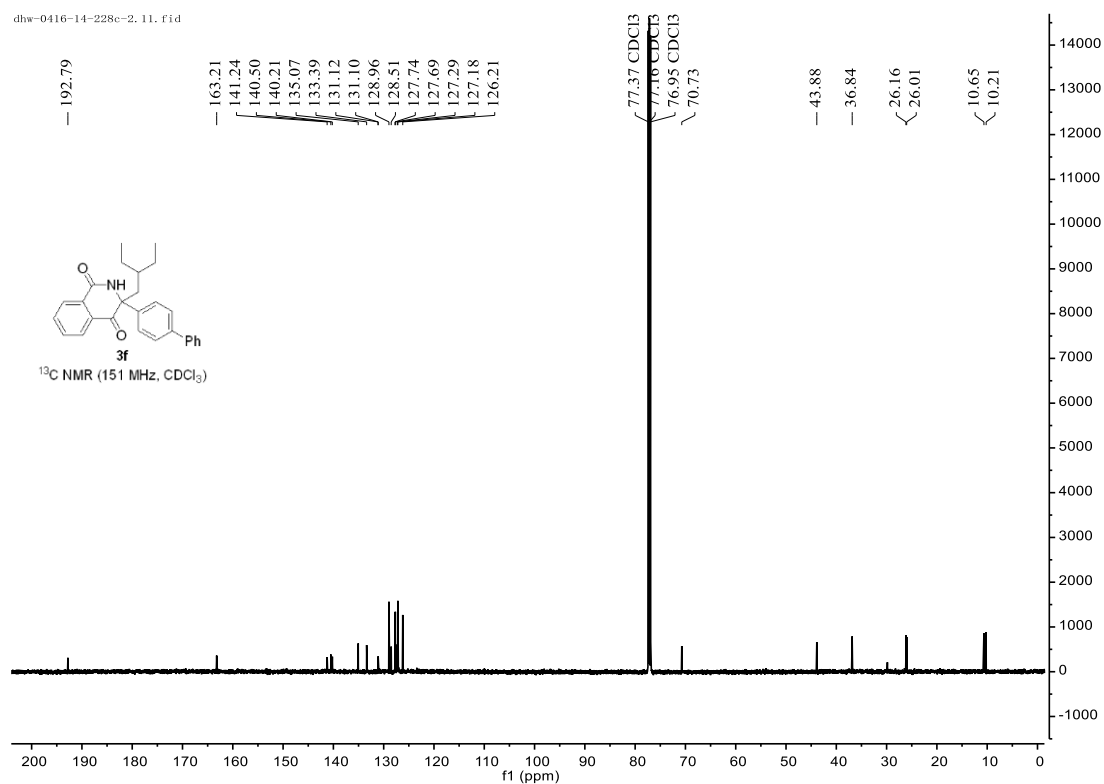
dhw-0628-6-199a, 33, fid

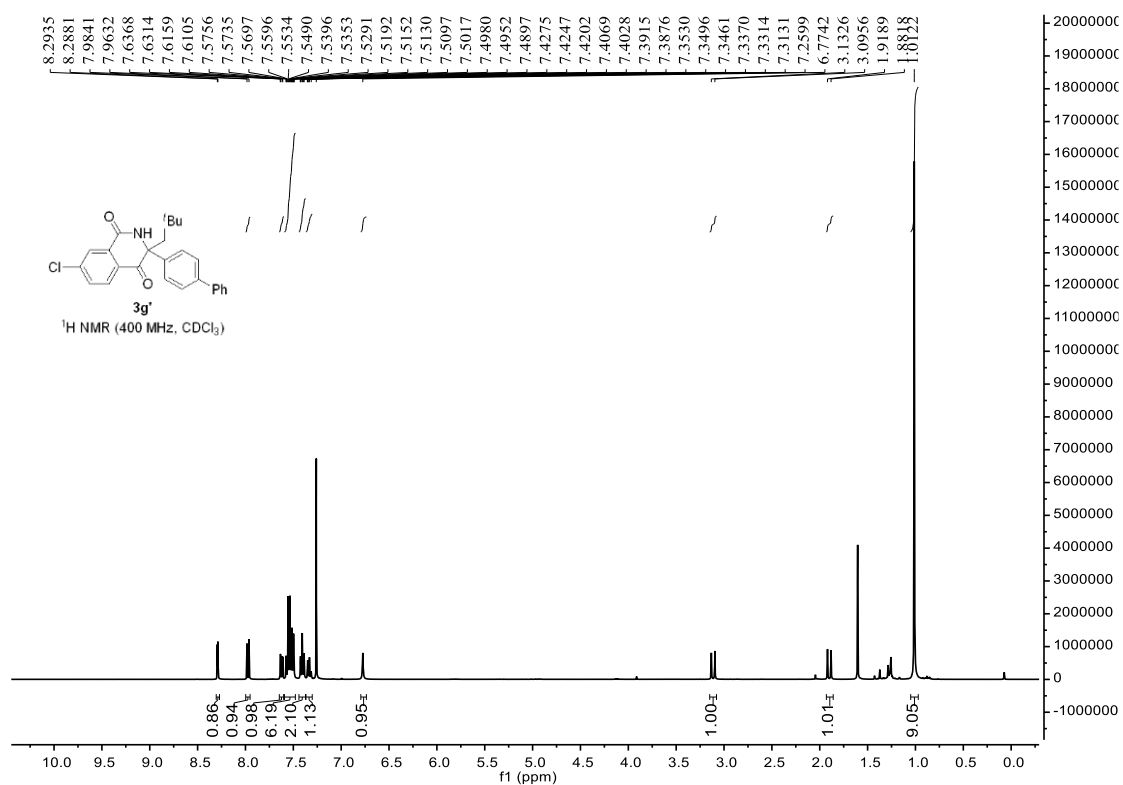
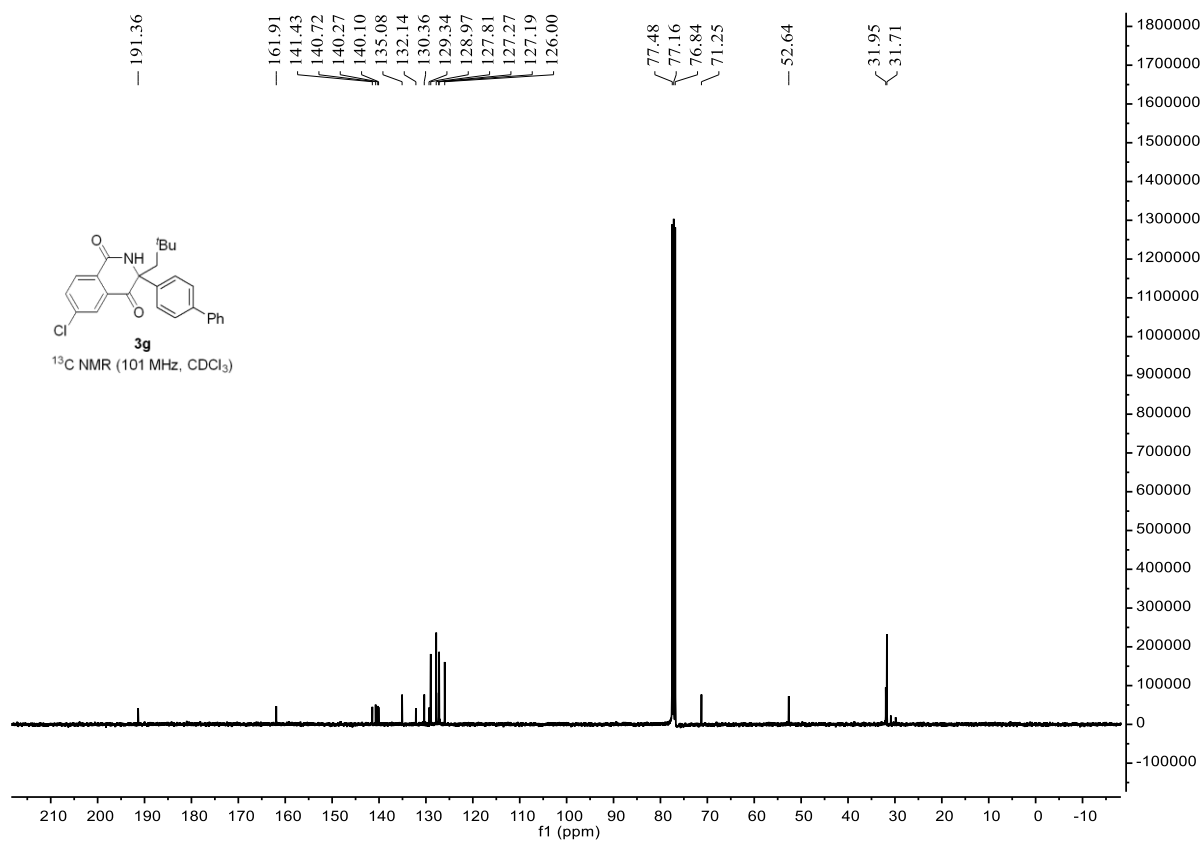


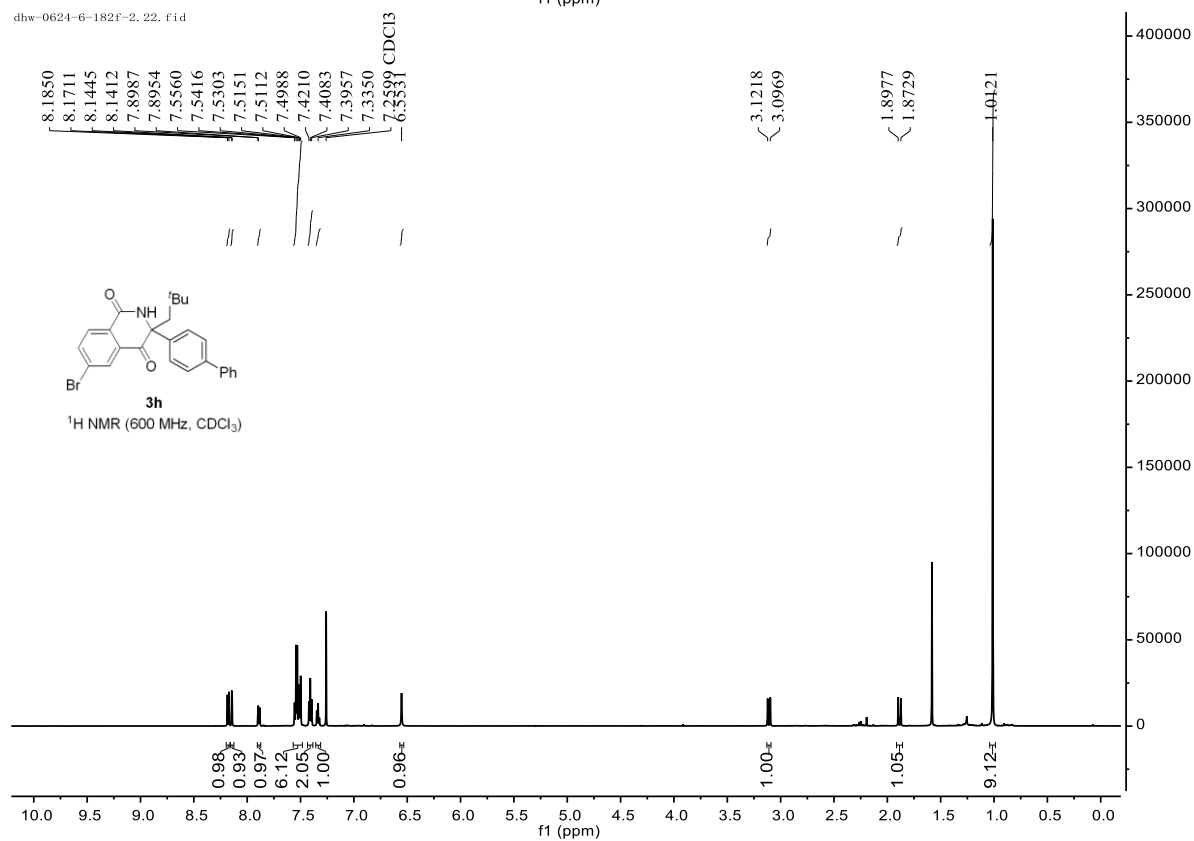
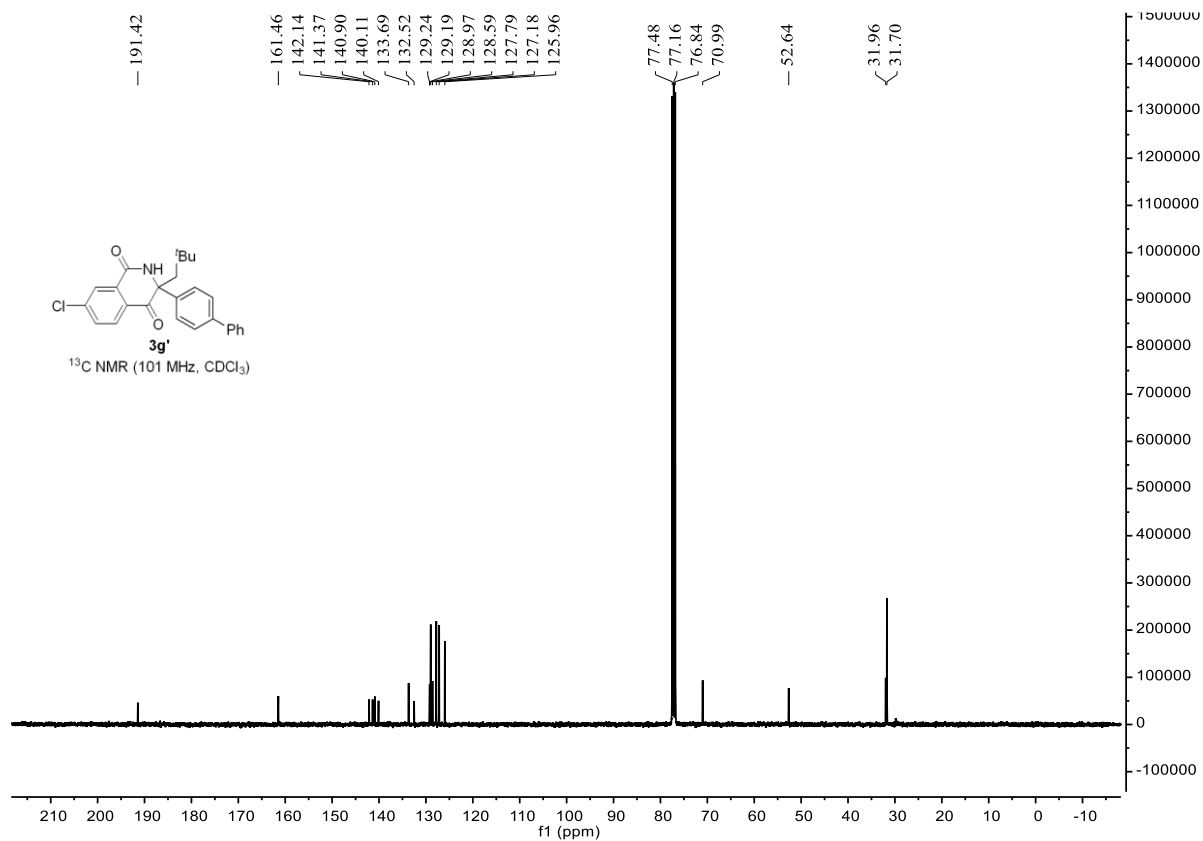
dhw-0323-gan-111, 10, fid



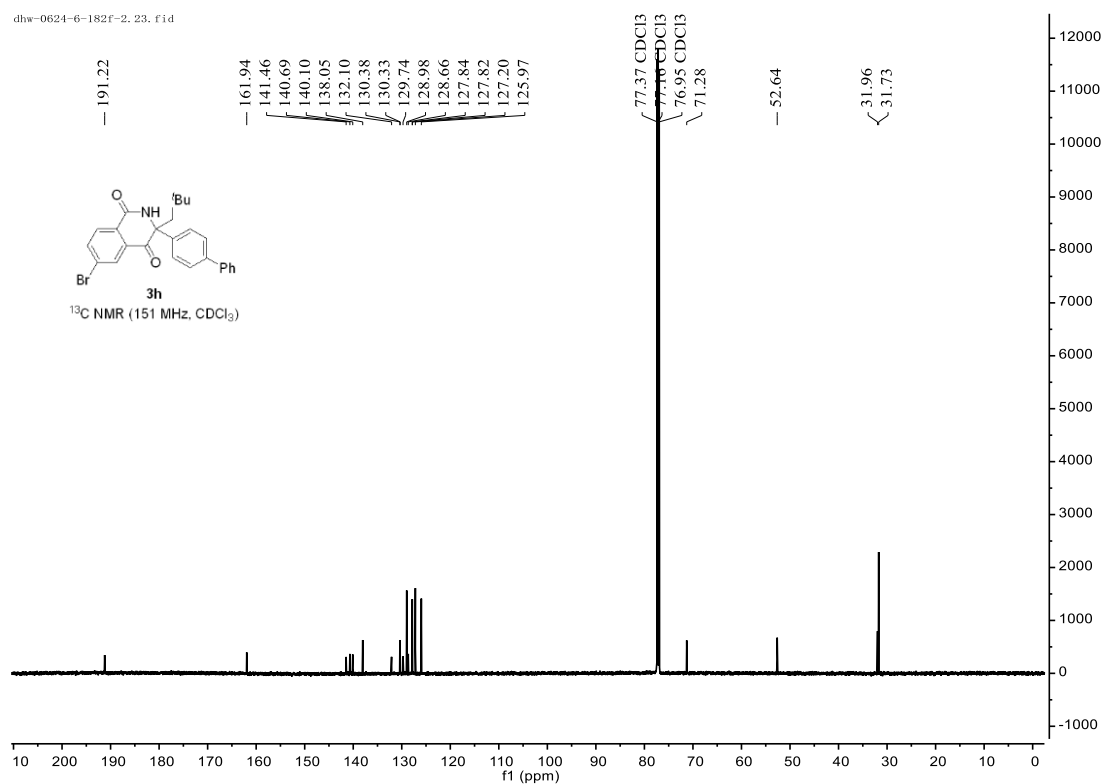




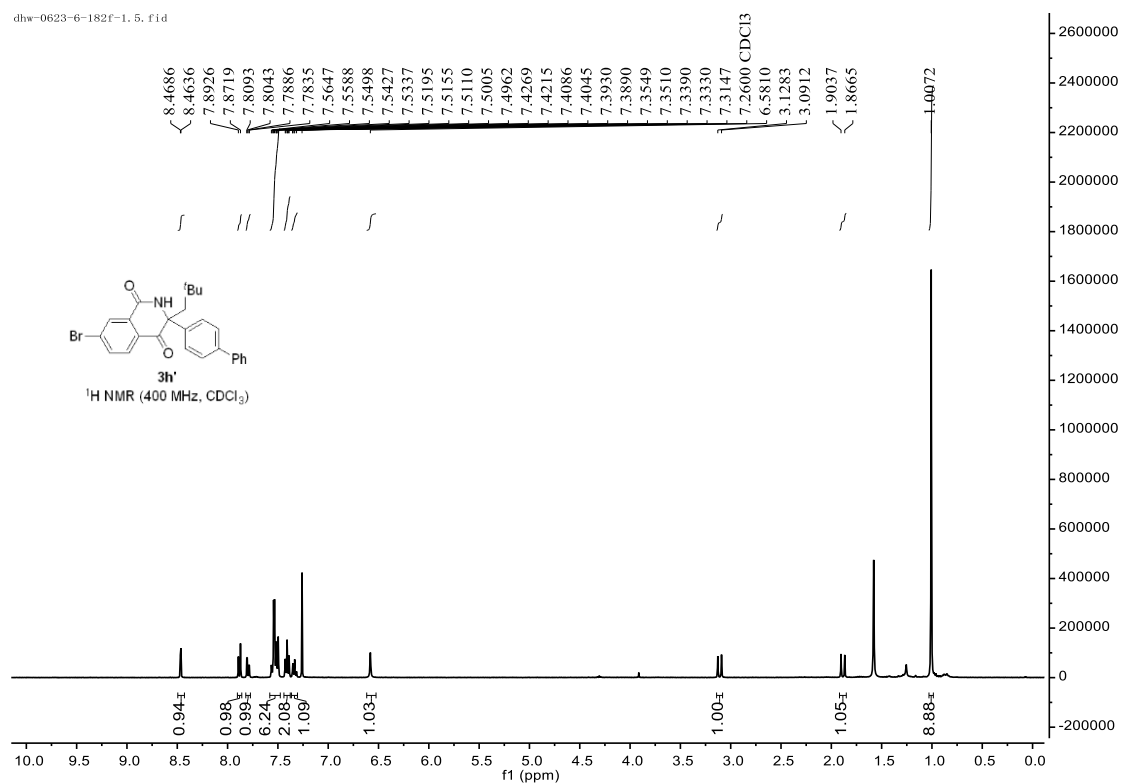




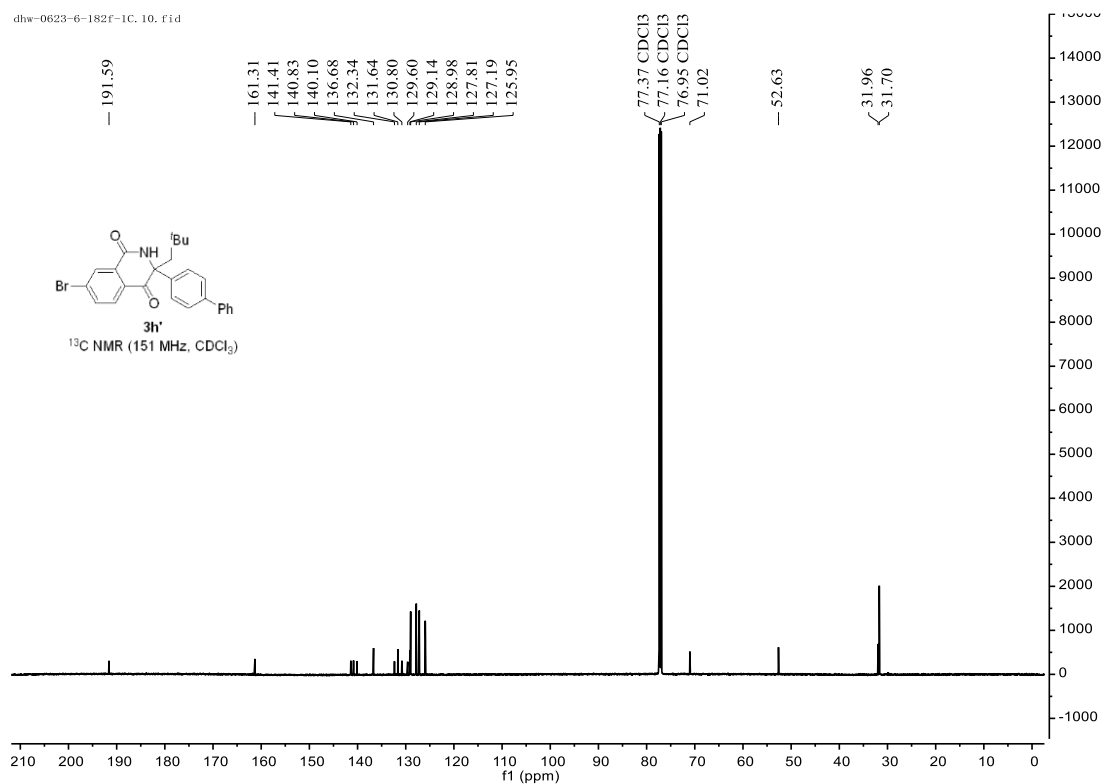
dhw-0624-6-182f-2, 23, f1d



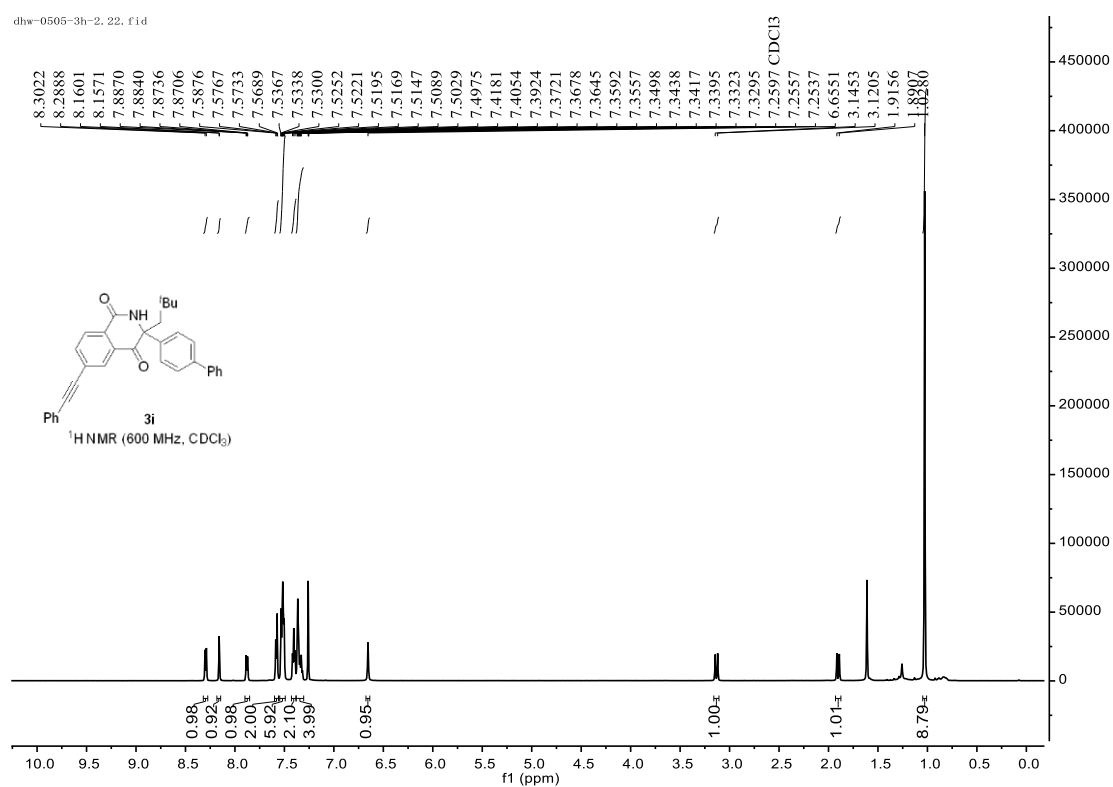
dhw-0623-6-182f-1, 5, f1d



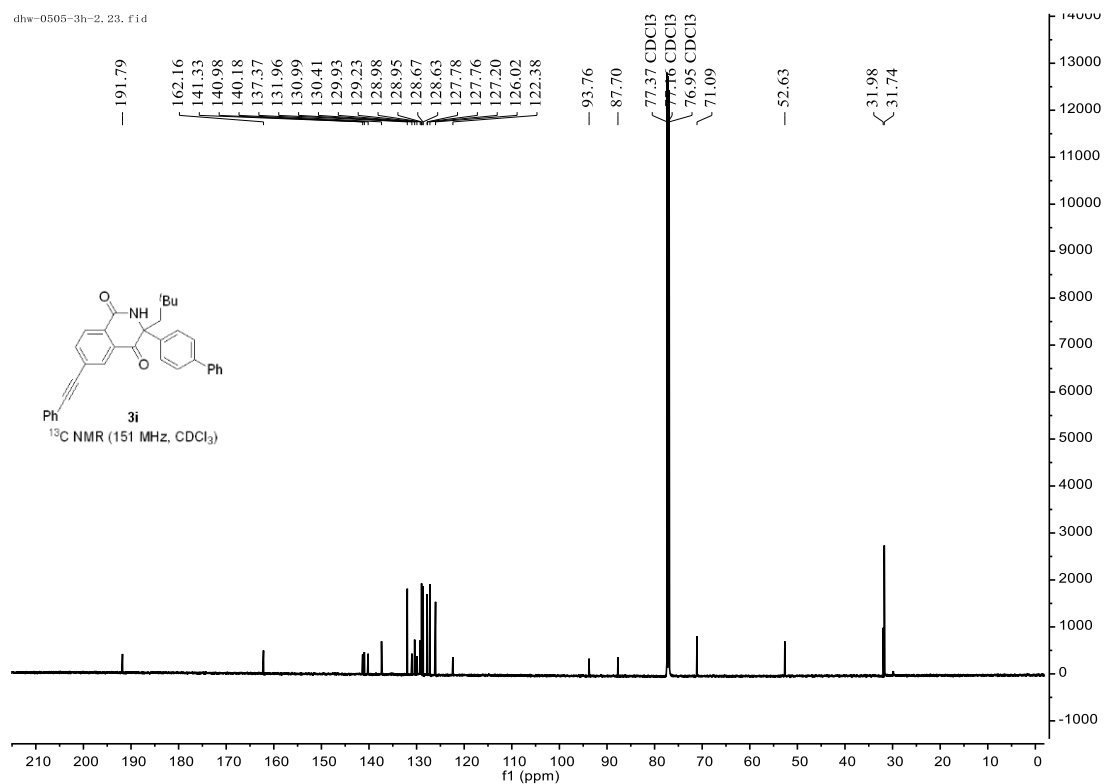
dhw-0623-6-182f-1C, 10, f1d



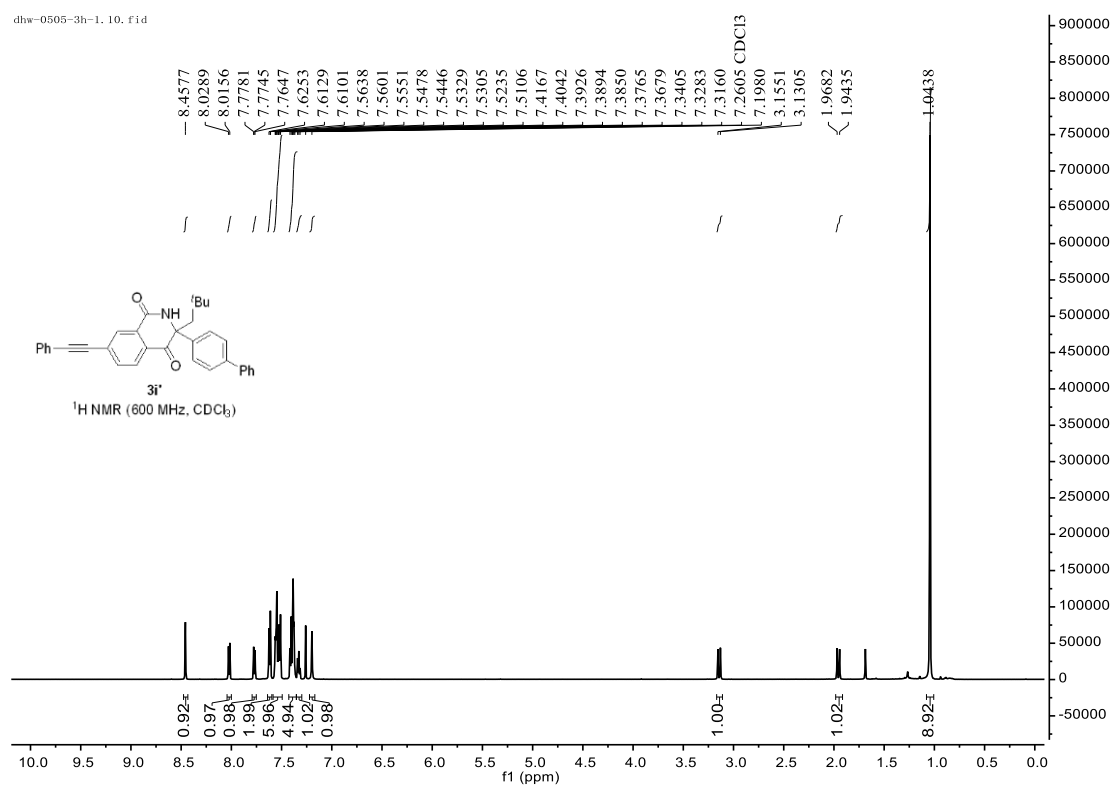
dhw-0505-3h-2. 22, f1d



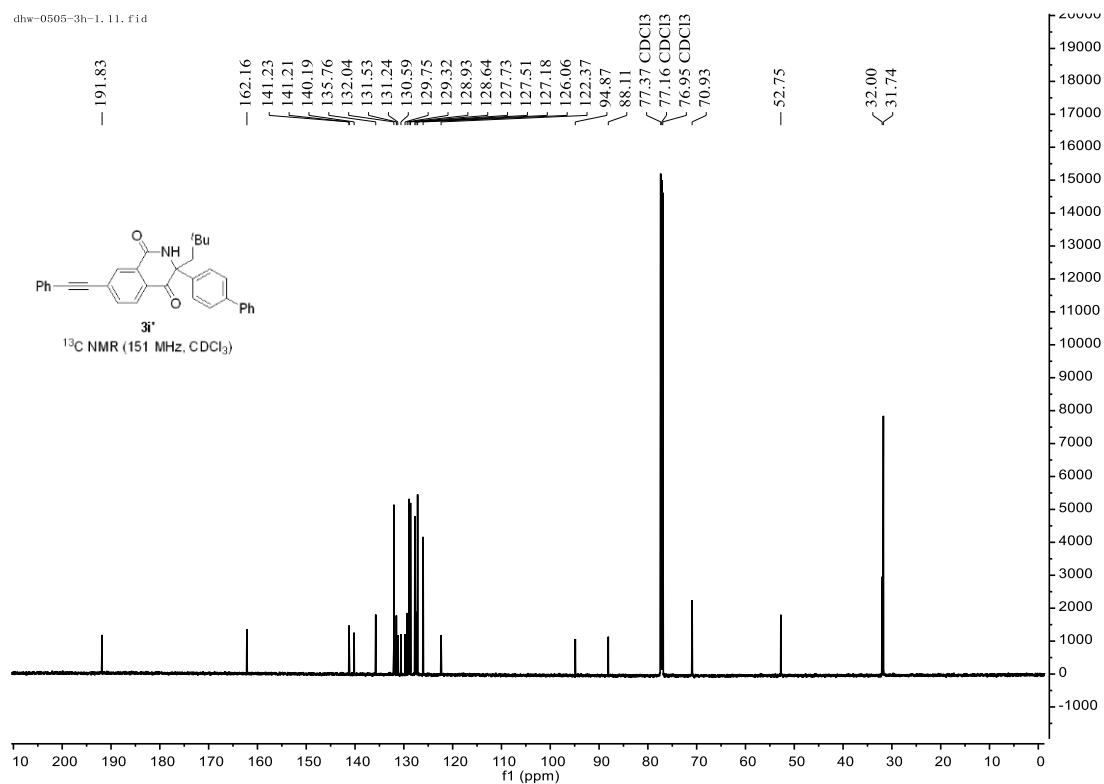
dhw-0505-3h-2, 23, f1d



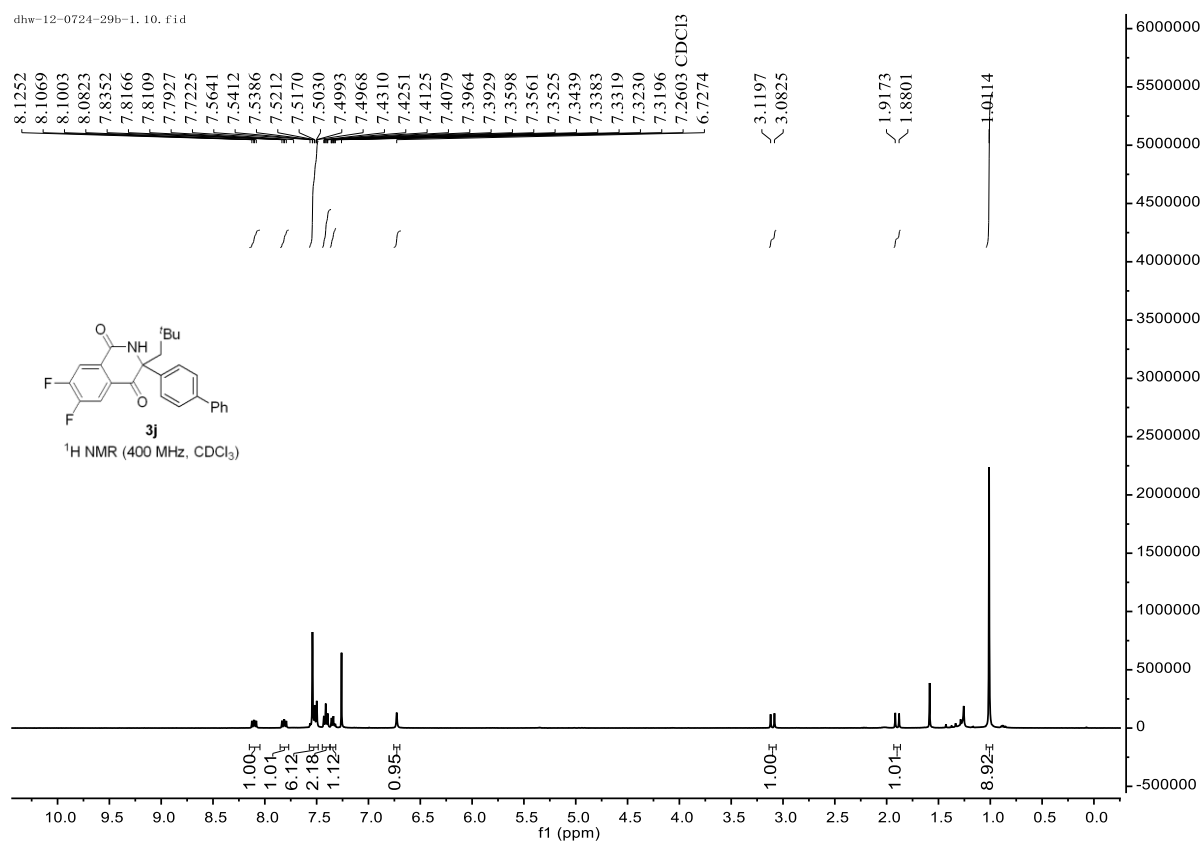
dhw-0505-3h-1, 10, f1d



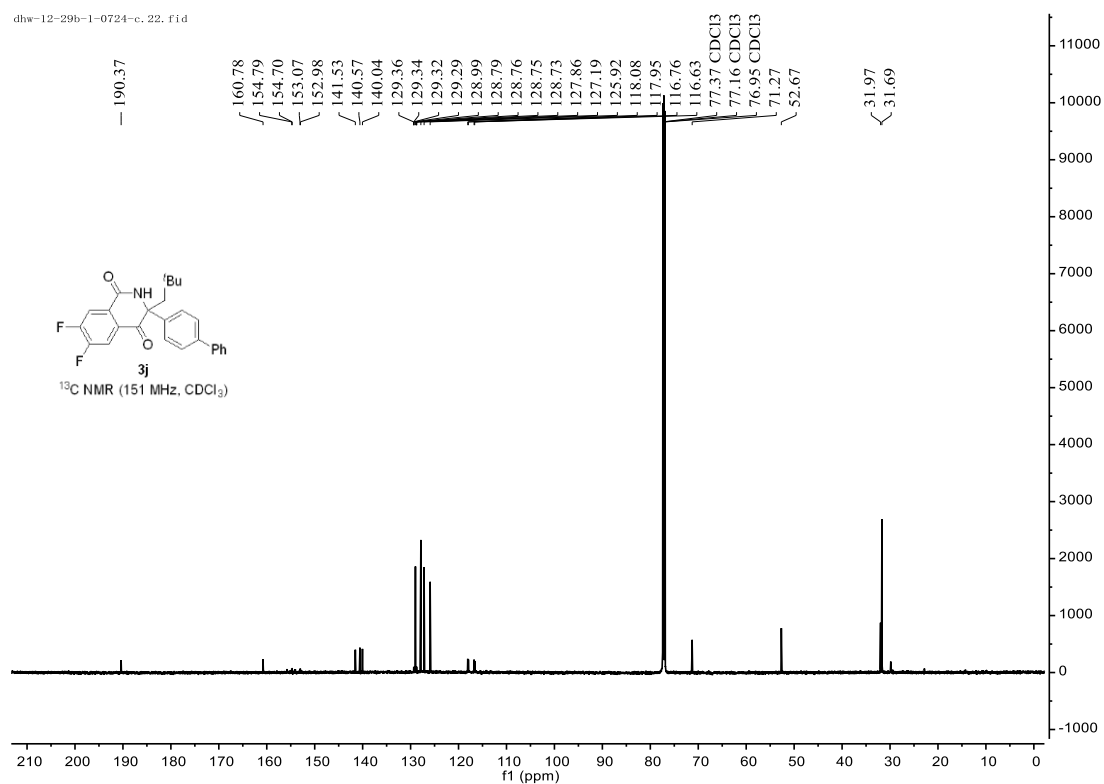
dhw-0505-3h-1, 11, f1d



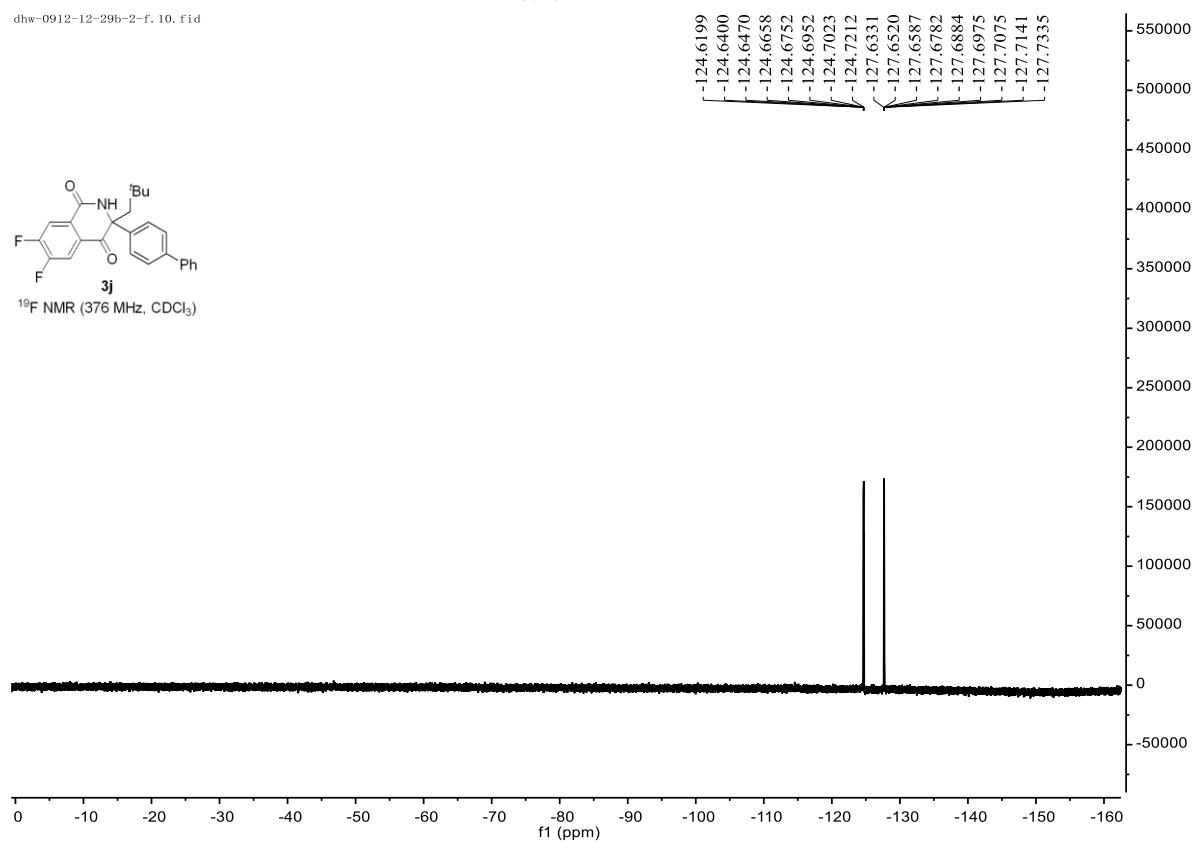
dhw-12-0724-29b-1, 10, f1d

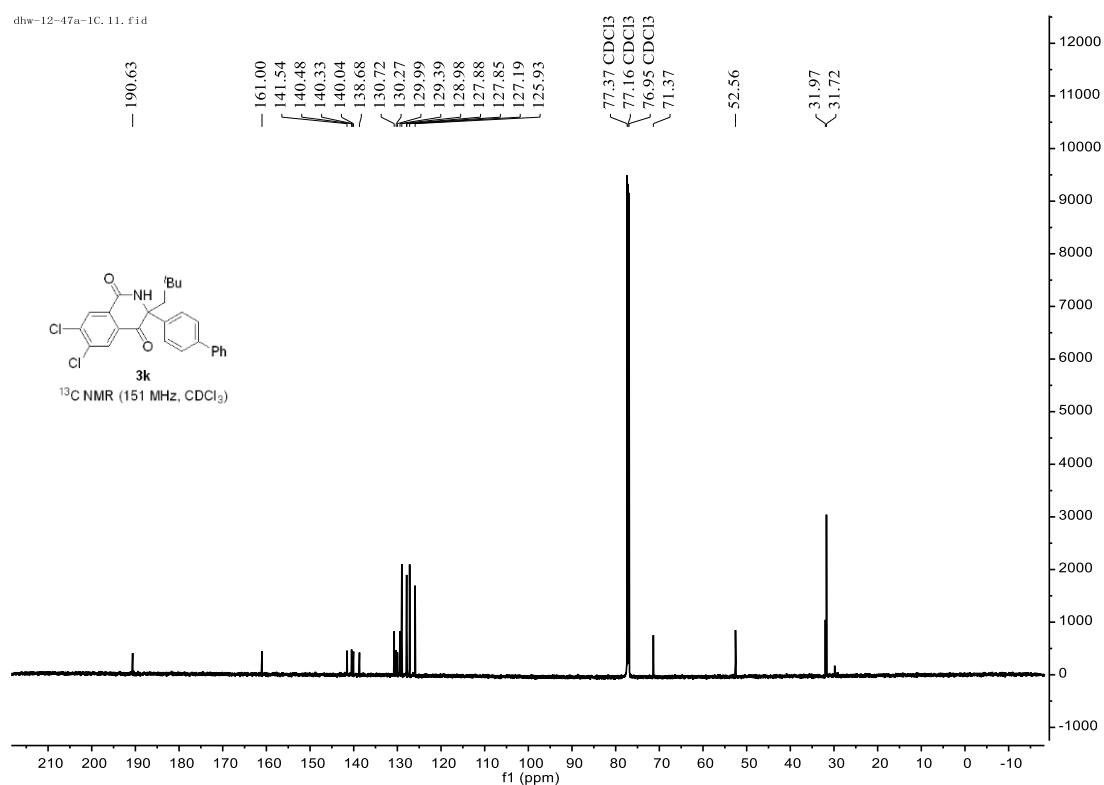
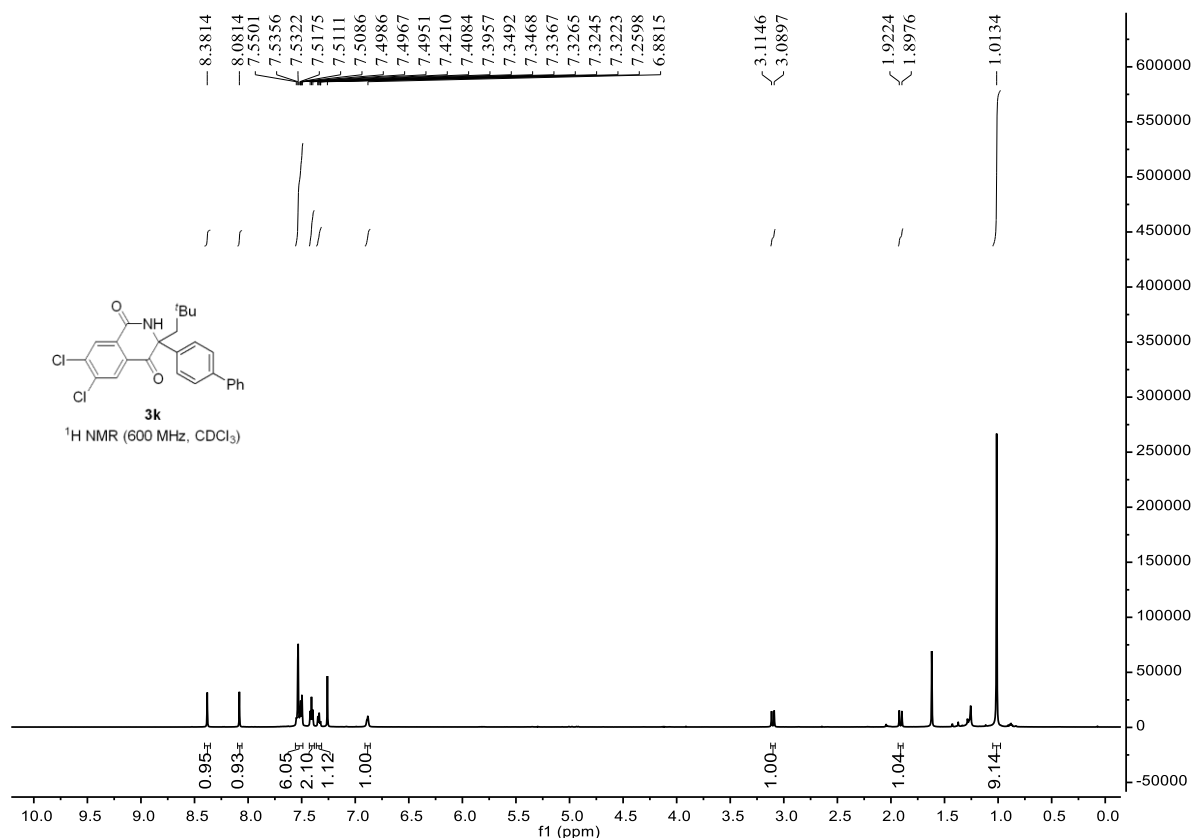


dhw-12-29b-1-0724-c, 22, f1d



dhw-0912-12-29b-2-f, 10, f1d





dhw-0504-3k-2, 52, f1d

CC(C)N1C(=O)c2ccccc2C(=O)N1C3=CC=CC=C3

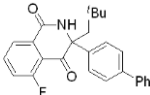
3l

¹H NMR (600 MHz, CDCl₃)

Chemical shift (ppm): 7.8731, 7.8601, 7.6515, 7.6444, 7.6375, 7.6301, 7.6225, 7.6172, 7.6147, 7.5836, 7.5805, 7.5695, 7.5663, 7.5384, 7.5352, 7.5242, 7.5205, 7.5158, 7.5033, 7.4707, 7.4546, 7.4391, 7.4188, 7.4074, 7.4046, 7.3943, 7.3913, 7.3435, 7.3327, 7.3296, 7.3192, 7.2572, 6.4398.

Integration values: 0.97, 1.16, 2.12, 4.22, 1.15, 2.12, 1.10, 0.95, 1.00, 1.05, 8.86.

dhw-0504-3k-2, 53, F1d



^{13}C NMR (151 MHz, CDCl_3)

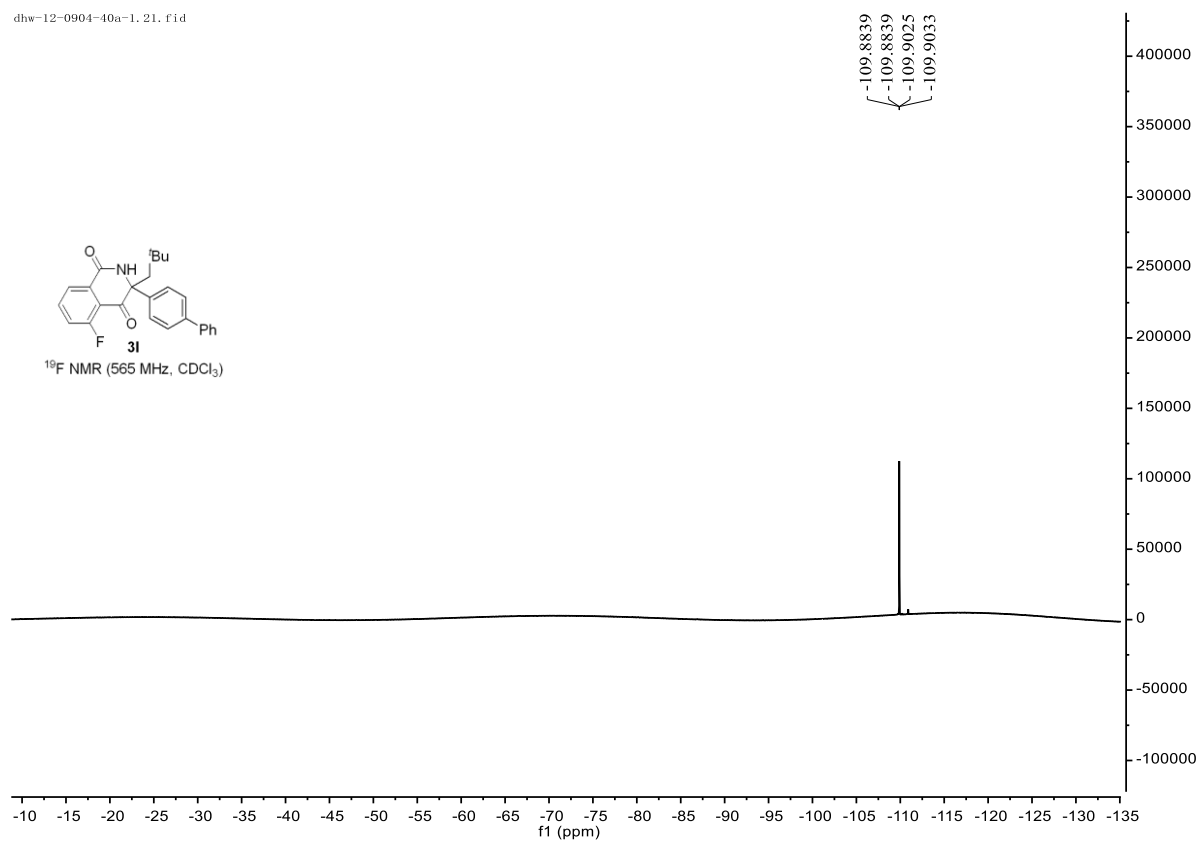
191.46
191.44
162.79
161.03
160.23
160.20
141.35
140.70
140.09
134.71
134.65
132.83
128.97
127.80
127.78
127.17
125.98
123.89
123.85
123.83
123.74
118.44
118.41
77.37 CDCl_3
77.16 CDCl_3
76.95 CDCl_3
70.79
52.58
31.94
31.75

13000
12000
11000
10000
9000
8000
7000
6000
5000
4000
3000
2000
1000
0
-1000

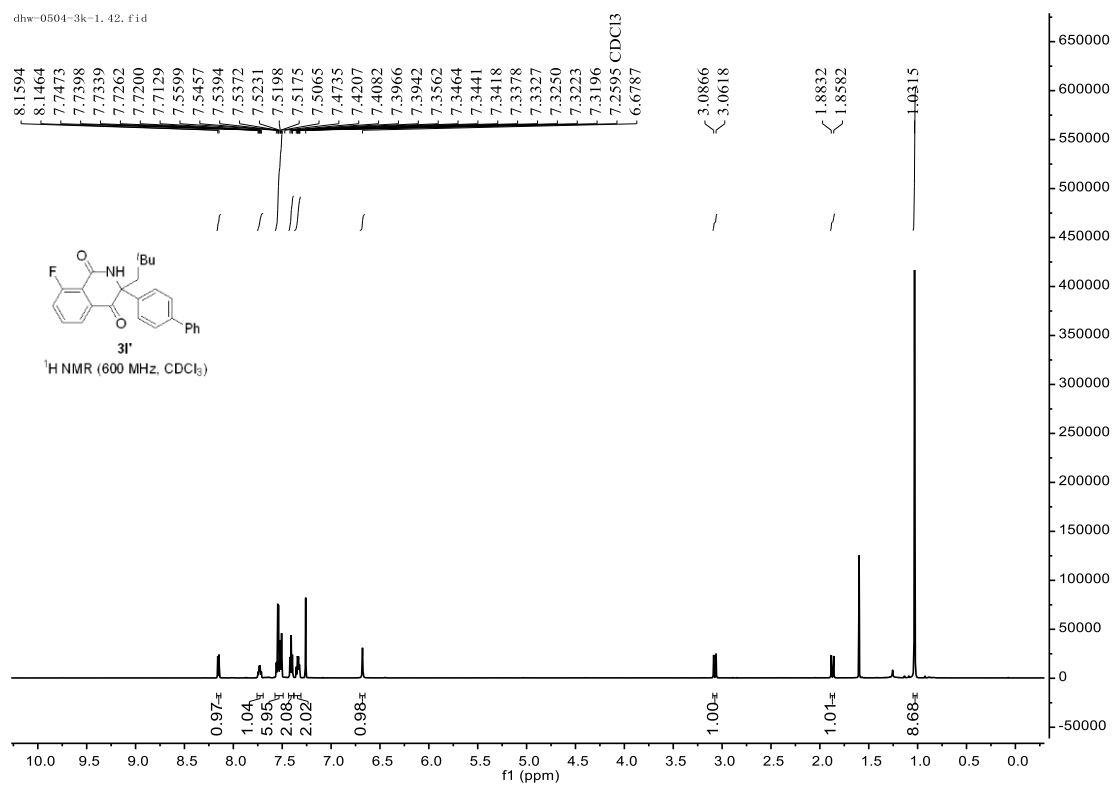
210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

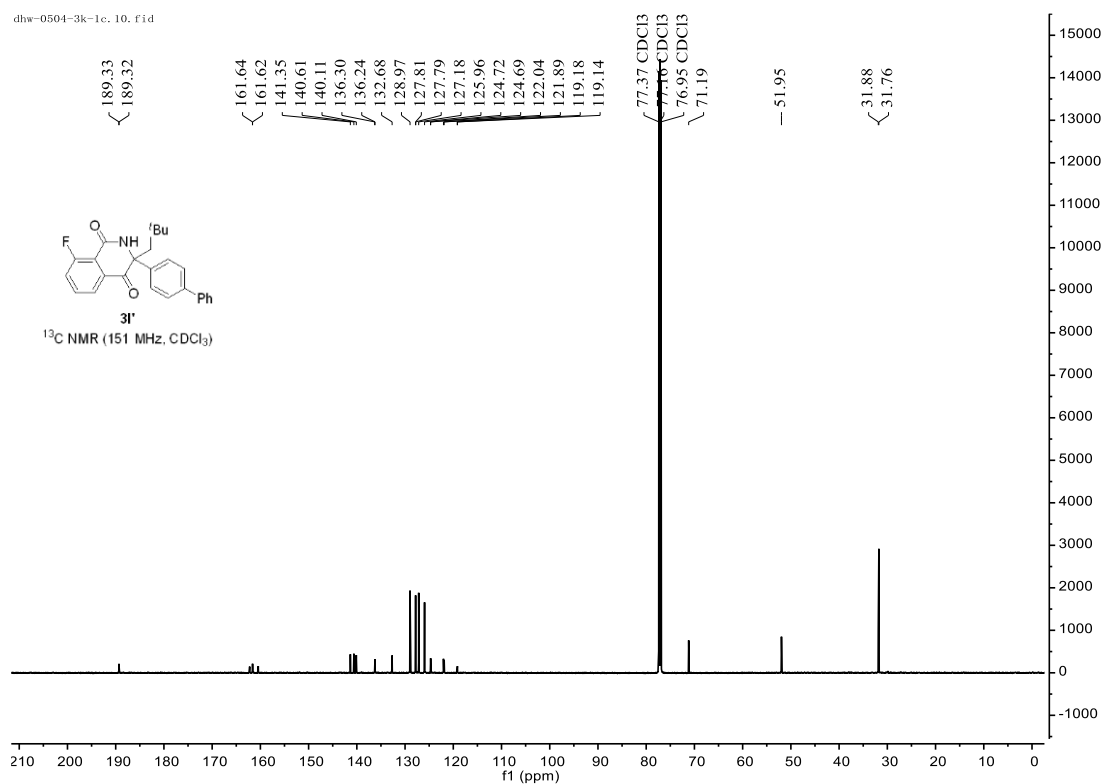
dhw-12-0904-40a-1.21.fid



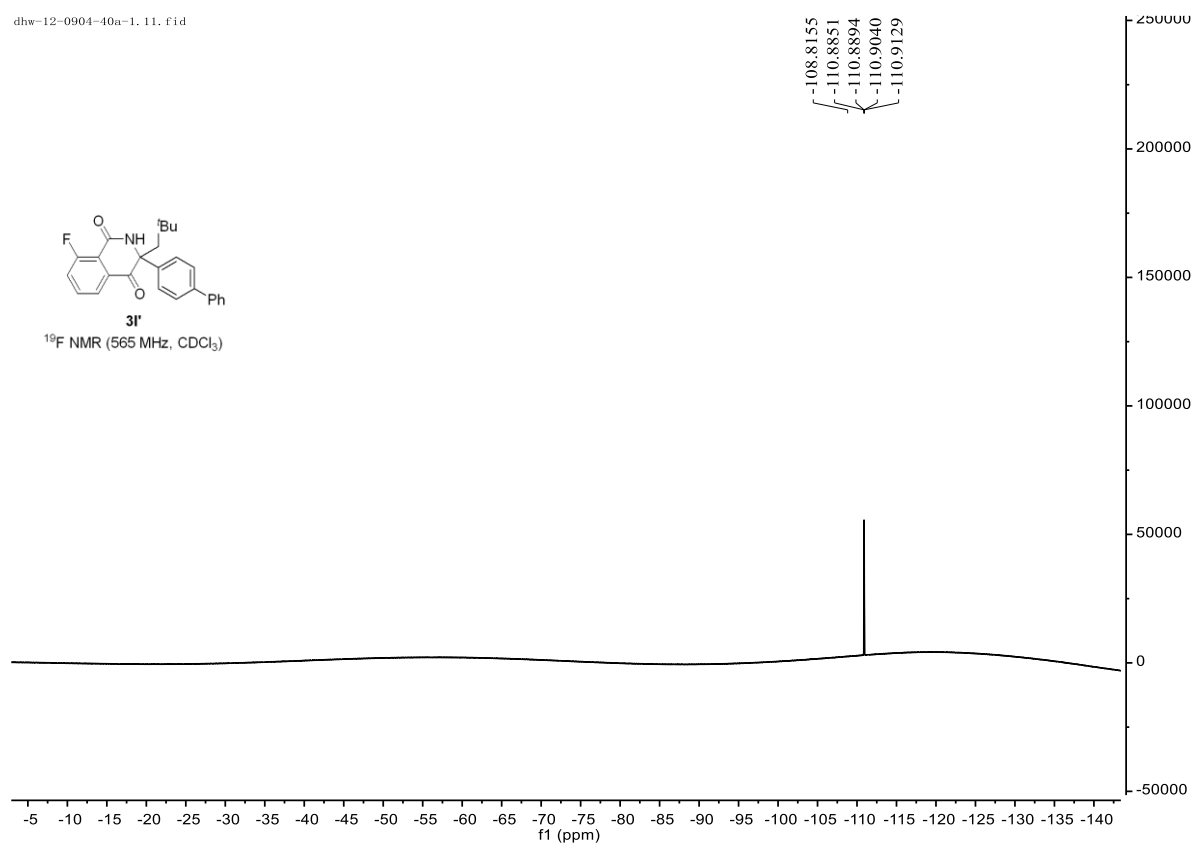
dhw-0504-3k-1.42.fid



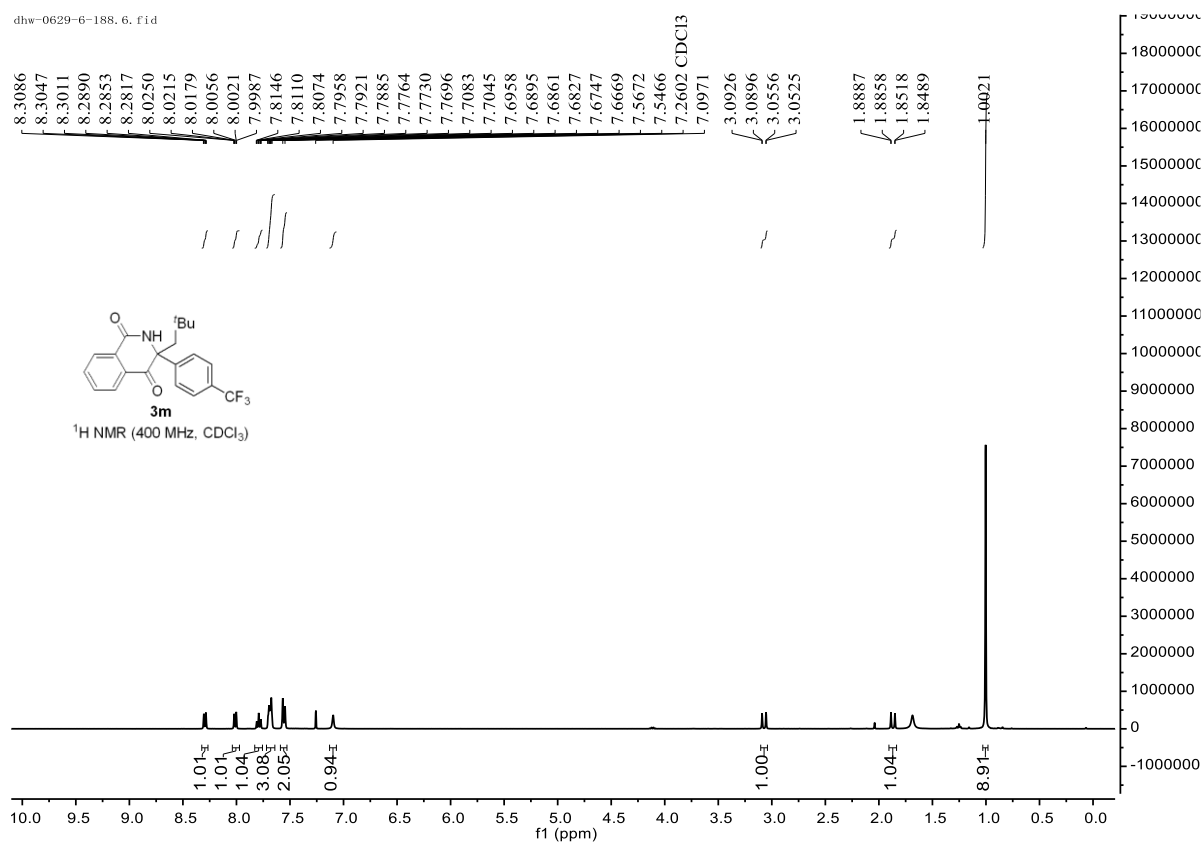
dhw-0504-3k-1c, 10, f1d



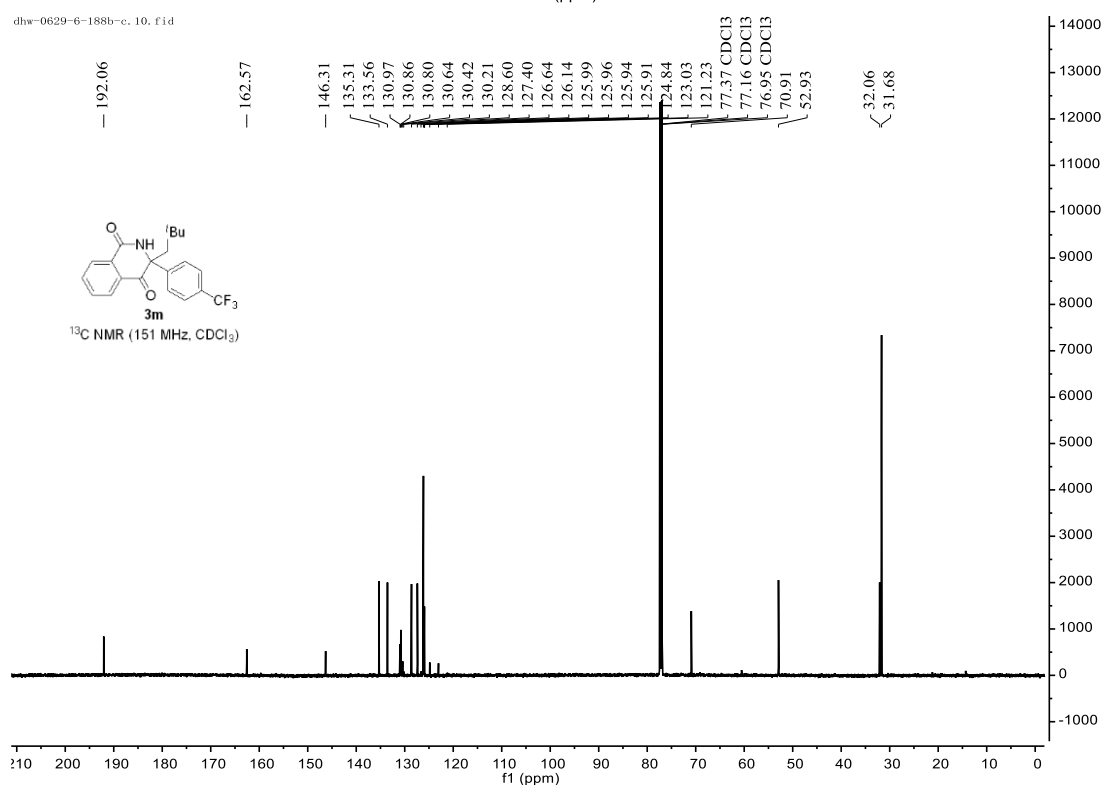
dhw-12-0904-40a-1, 11, f1d



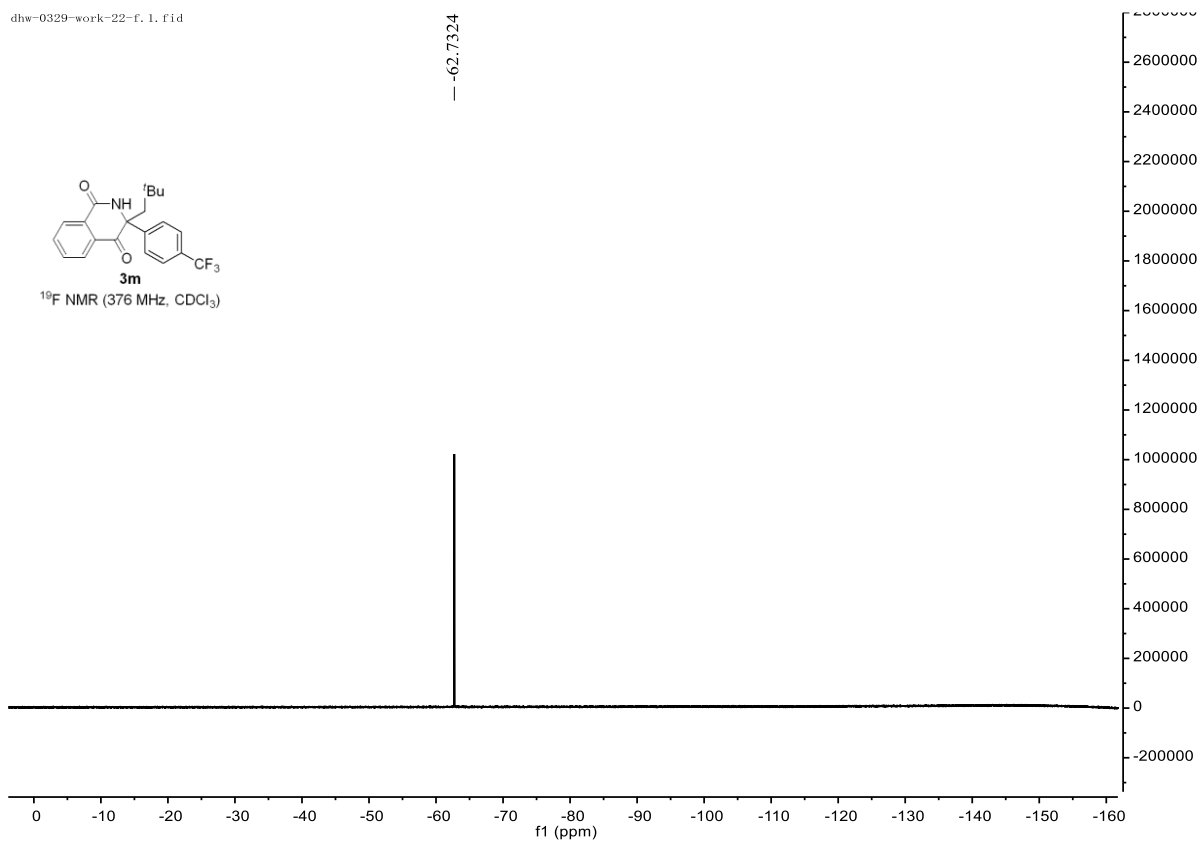
dhw-0629-6-188, 6, f1d



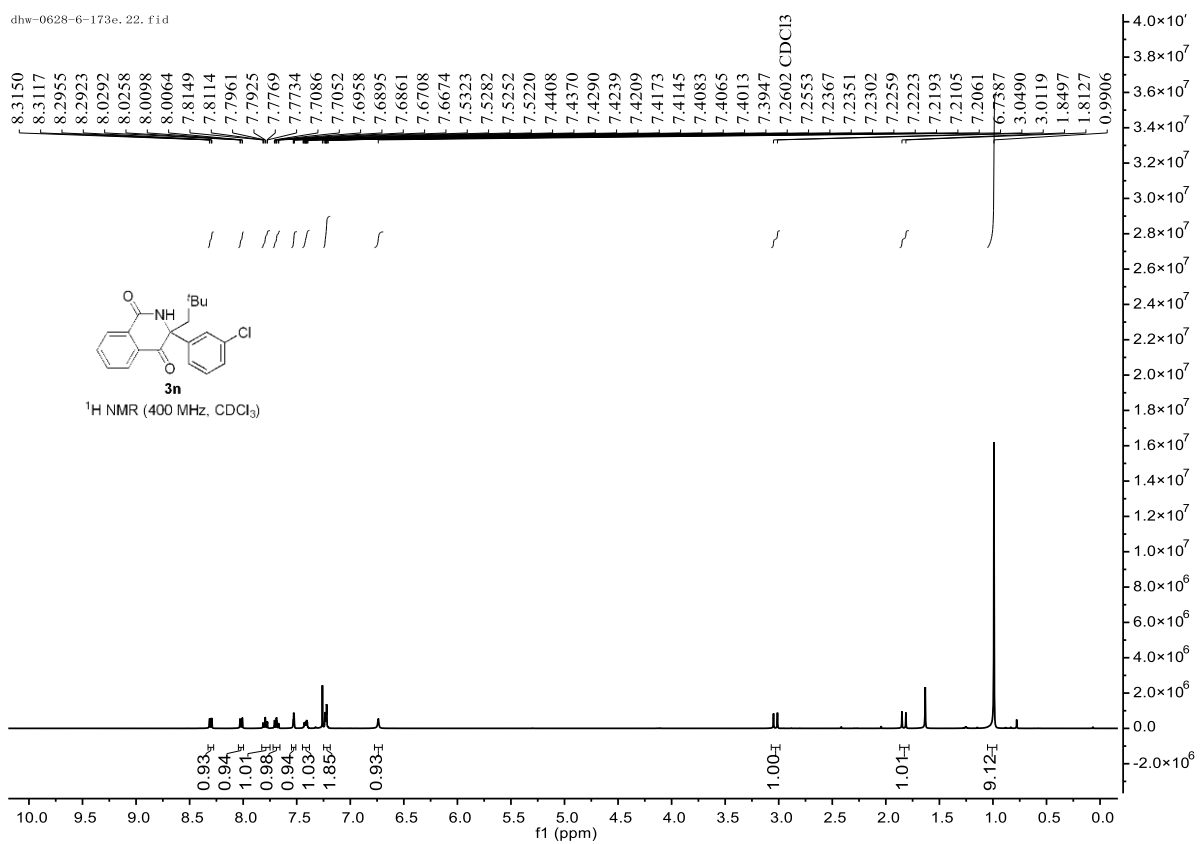
dhw-0629-6-188b-c, 10, f1d



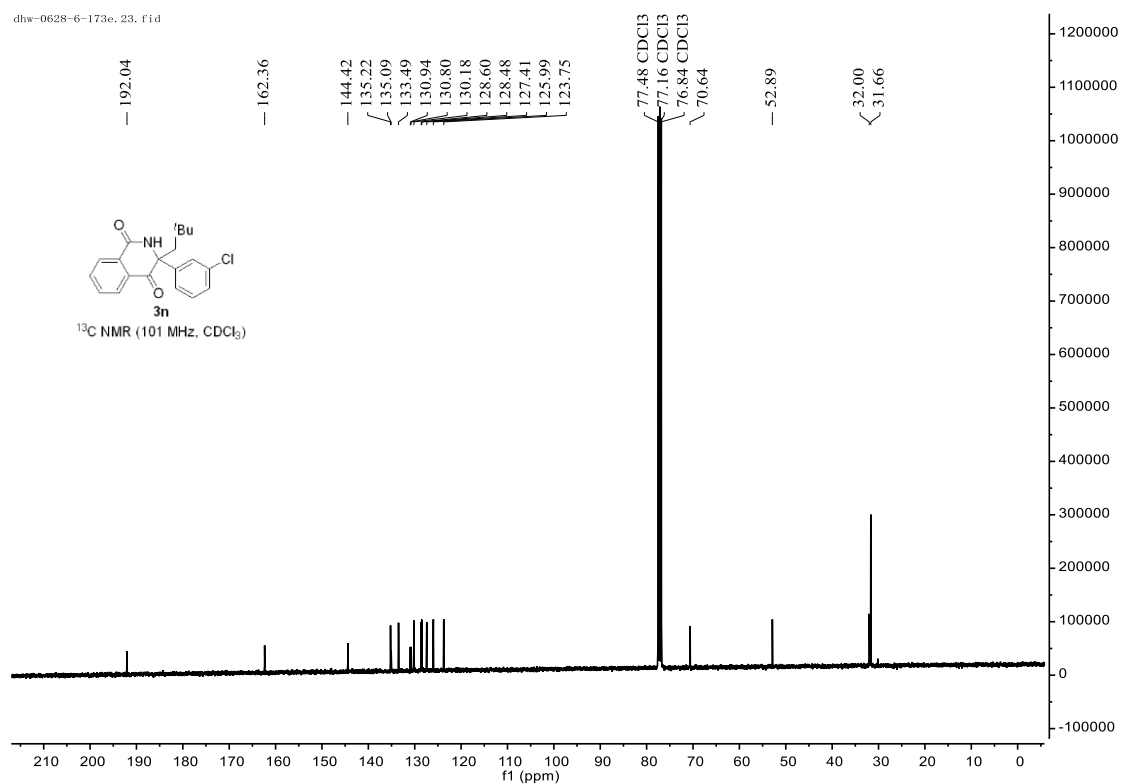
dhw-0329-work-22-f. 1. fid



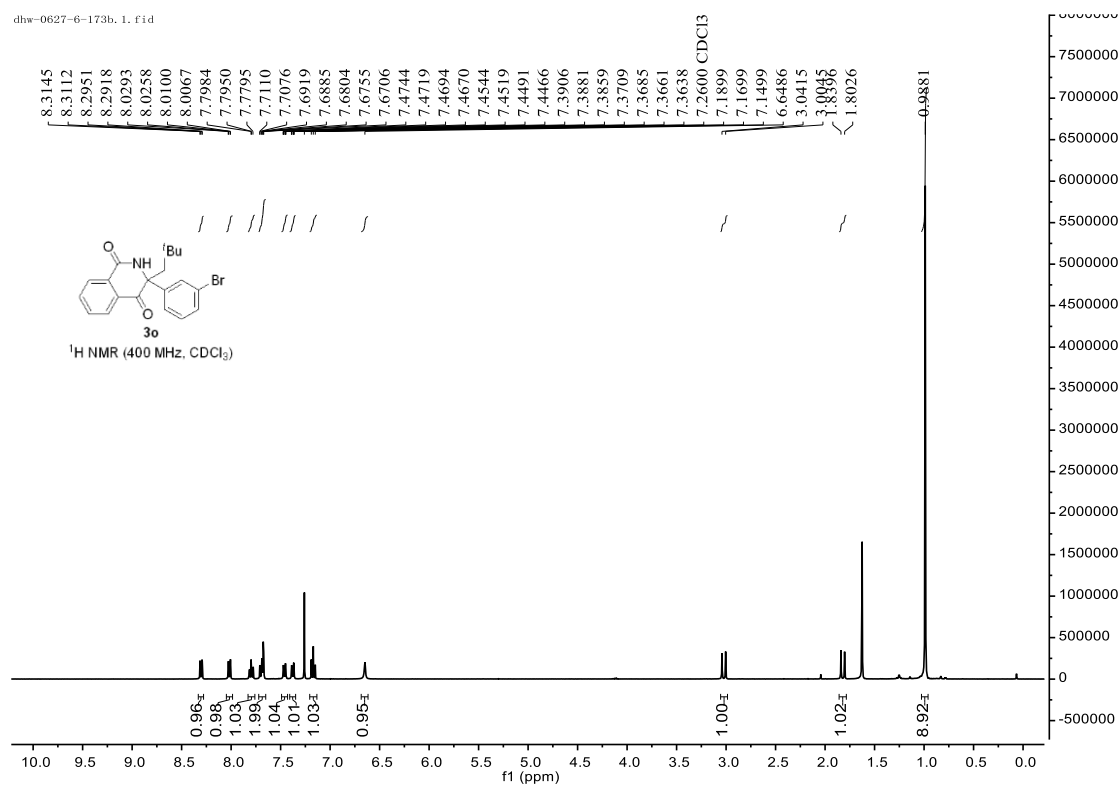
dhw-0628-6-173e. 22. fid



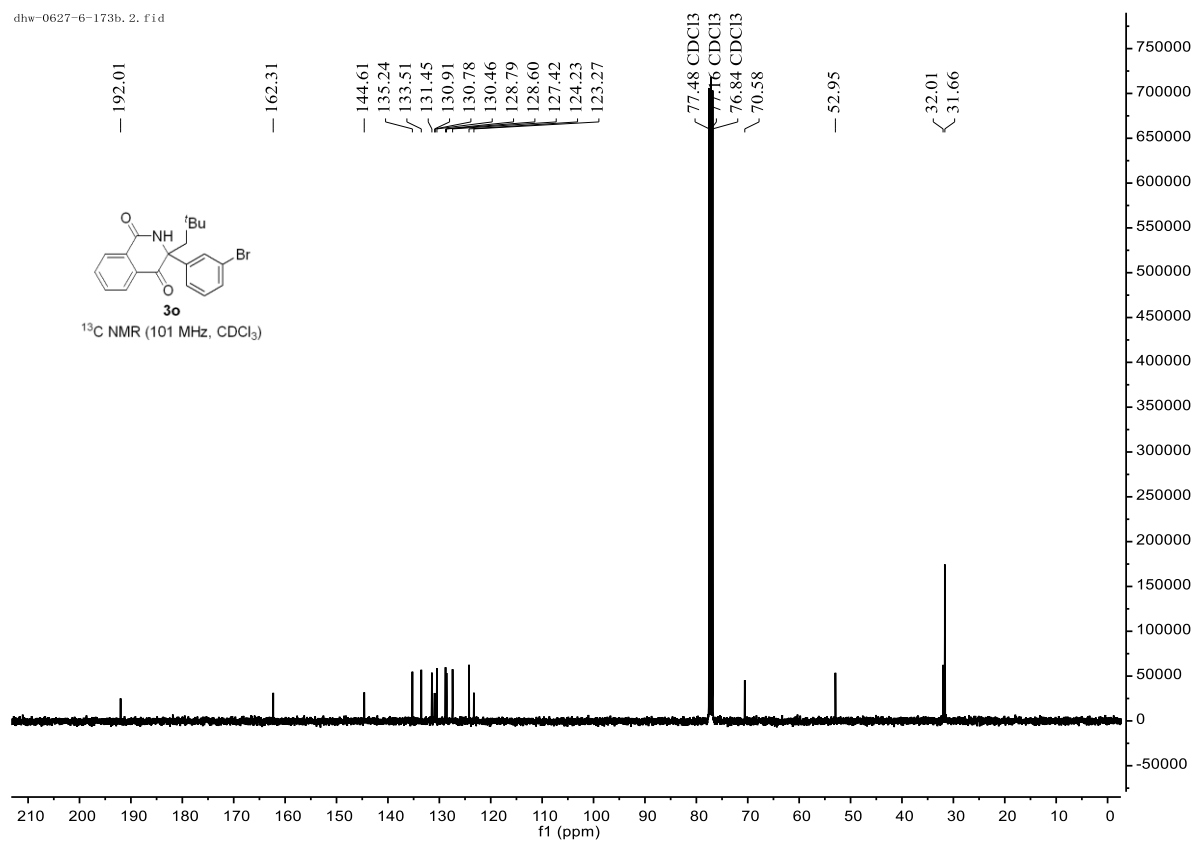
dhw-0628-6-173e, 23, fid



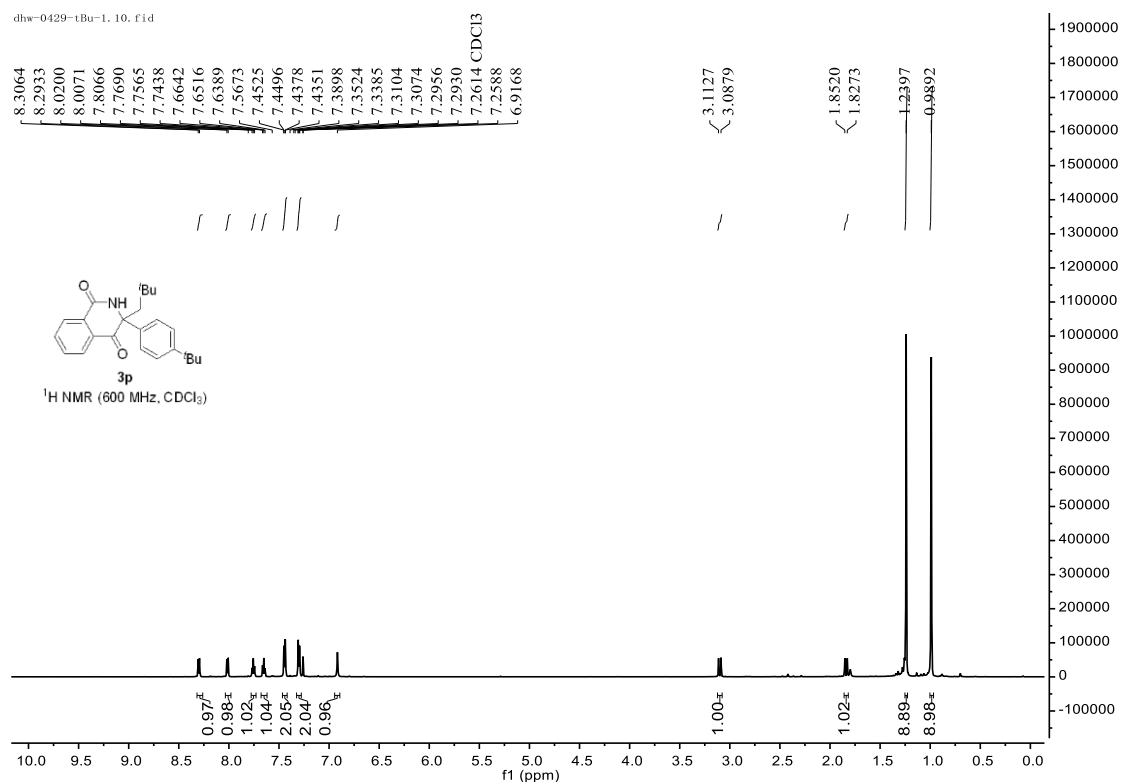
dhw-0627-6-173b, 1, fid



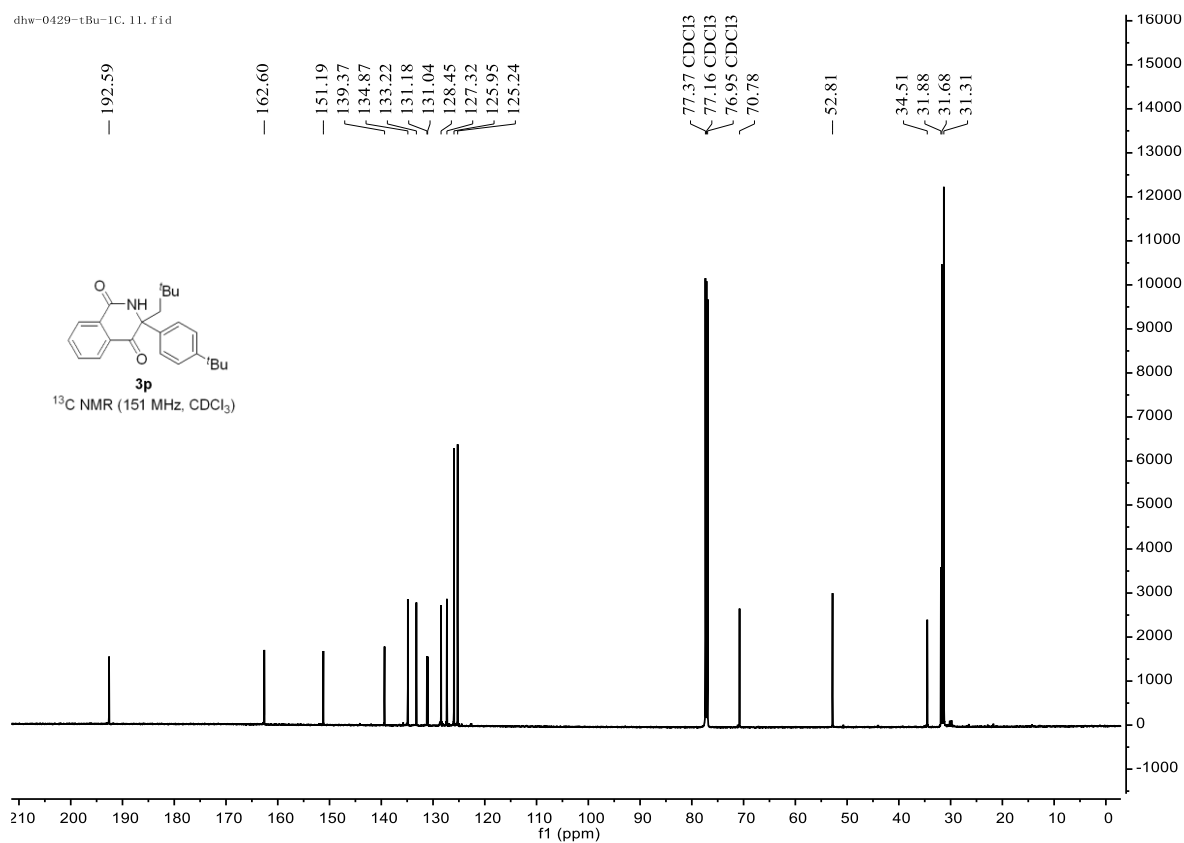
dhw-0627-6-173b. 2. fid



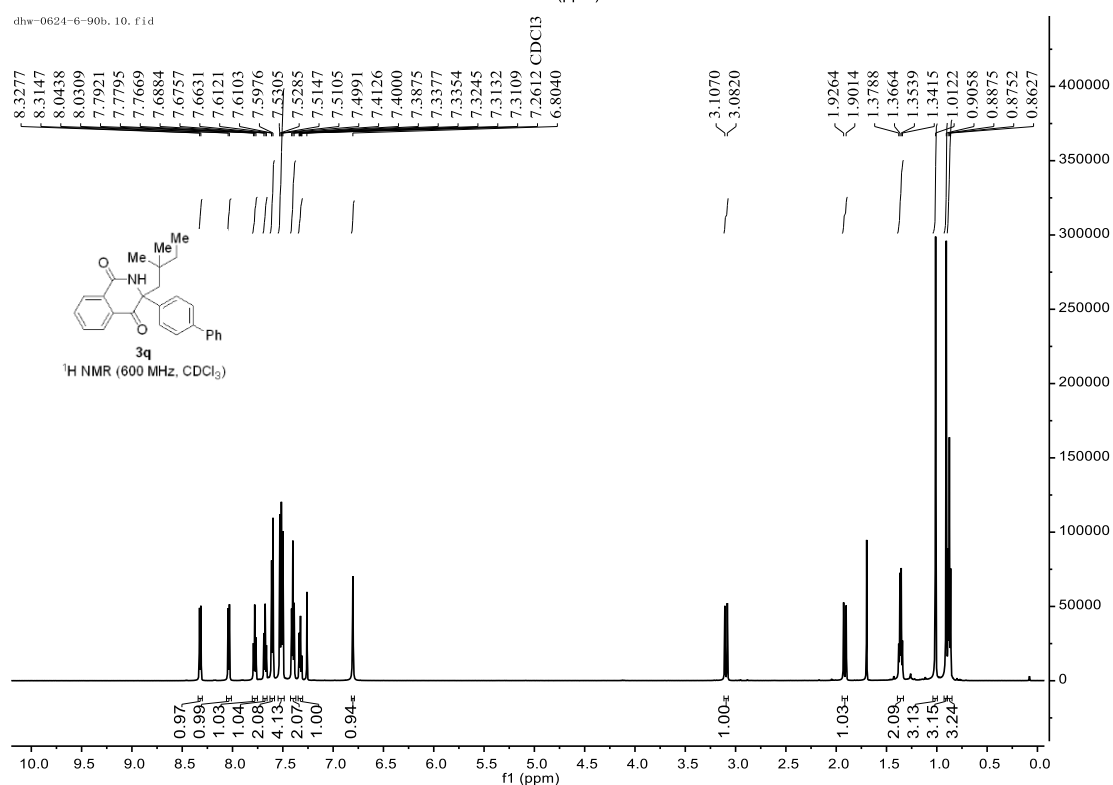
dhw-0429-tBu-1. 10. fid



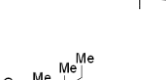
dhw-0429-tBu-1C. 11. f1d



dhw-0624-6-90b. 10. f1d



dhw-0624-6-90b.11.fid


3q

^{13}C NMR (151 MHz, CDCl_3)

192.46
162.61
141.44
141.15
140.19
135.00
133.36
131.08
130.97
128.93
128.51
127.71
127.69
127.40
127.16
126.05
77.37 CDCl₃
77.16 CDCl₃
76.95 CDCl₃
70.83
50.66
37.12
34.46
28.85
27.91
8.64

11000
10000
9000
8000
7000
6000
5000
4000
3000
2000
1000
0
-1000

210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

f1 (ppm)

dhw-0414-3p-1.10.fid

3r

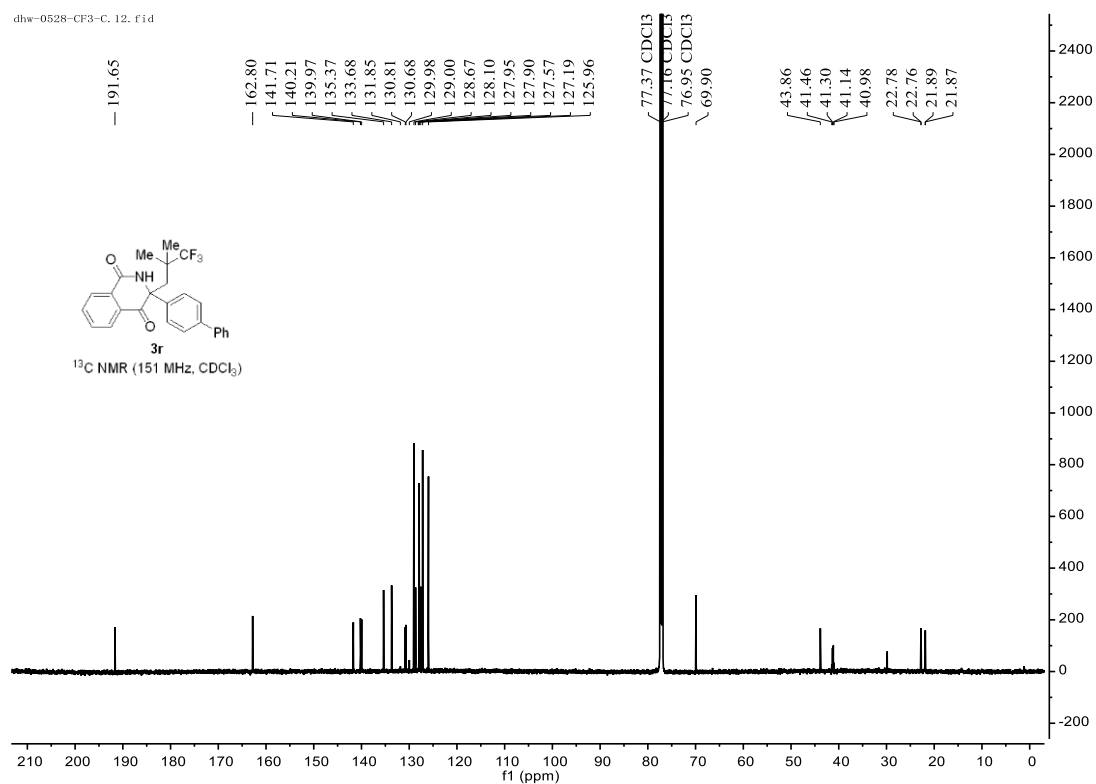
Cc1ccc(cc1)C(=O)N2C(=O)c3ccccc3C2=O

¹H NMR (600 MHz, CDCl₃)

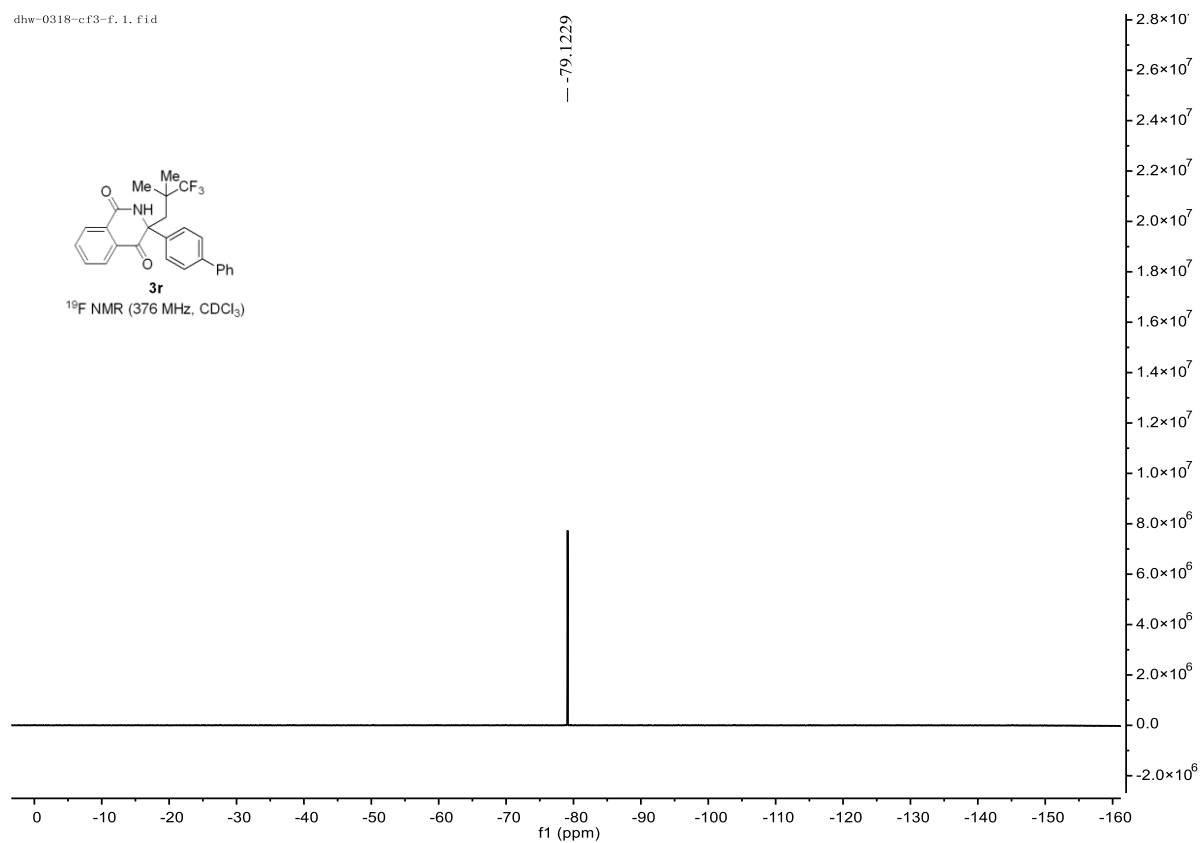
Chemical shift (ppm): 8.3215, 8.3084, 8.0565, 8.0436, 7.8167, 7.8146, 7.8041, 7.8020, 7.7915, 7.7894, 7.7158, 7.7137, 7.7016, 7.6906, 7.6885, 7.5894, 7.5754, 7.5453, 7.5347, 7.5310, 7.5263, 7.5084, 7.5057, 7.4933, 7.4187, 7.4062, 7.3933, 7.3501, 7.3480, 7.3460, 7.3354, 7.3255, 7.3234, 7.3214, 7.2597 CDCl₃, 6.7738, 3.4185, 3.3930, 2.1268, 2.1013, 1.3229, 1.1257.

Integration: 1.00, 1.01, 1.06, 1.09, 2.06, 2.04, 2.02, 1.06, 0.98, 1.06, 1.07, 3.05, 3.00.

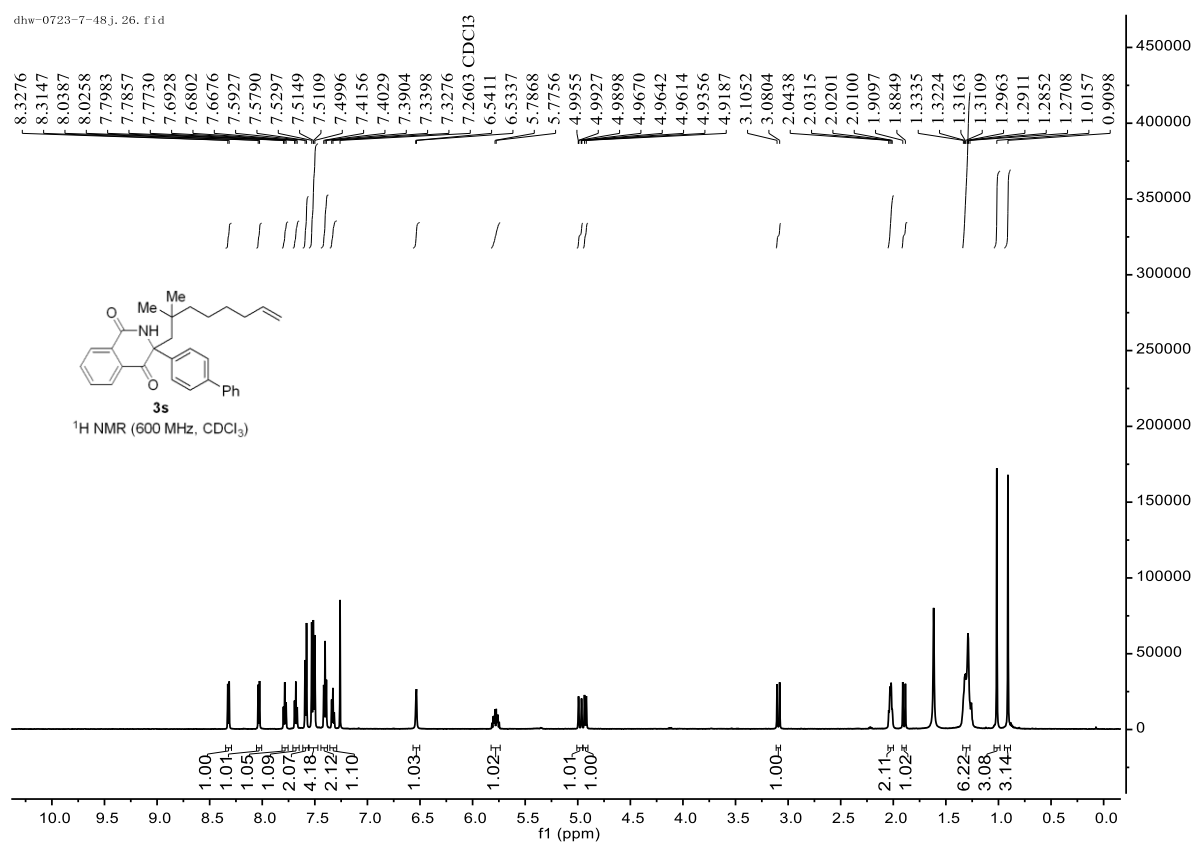
dhw-0528-CF3-C, 12, f1d



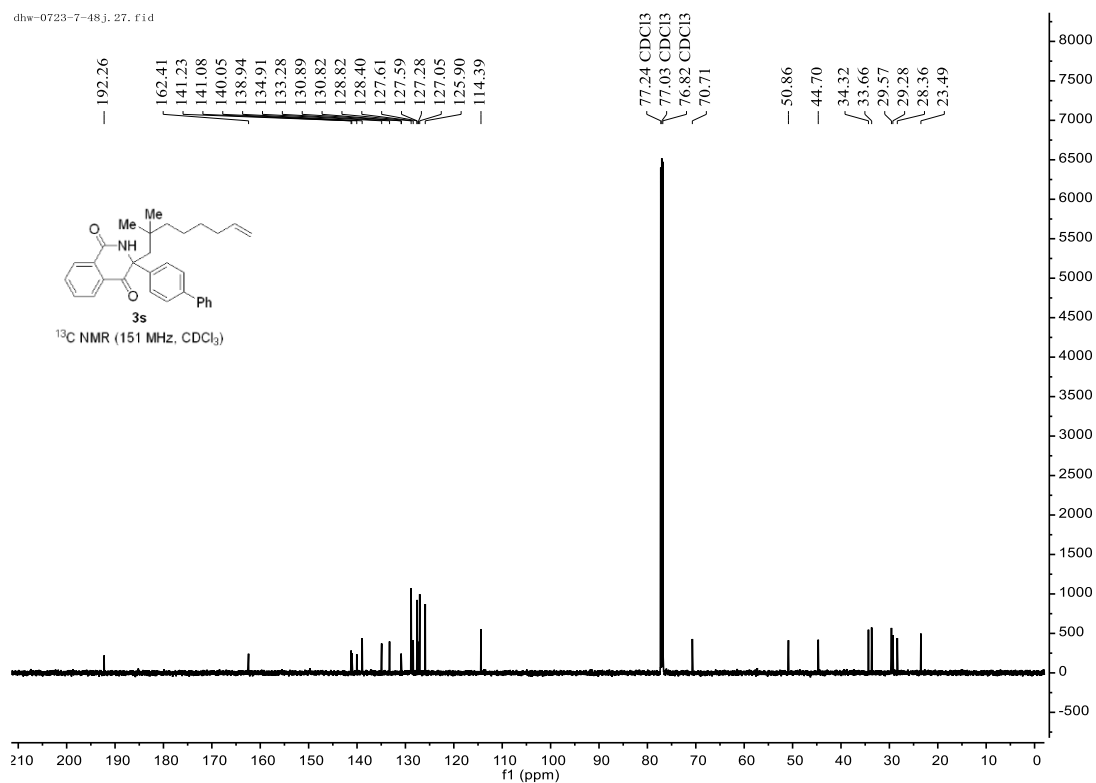
dhw-0318-cf3-f, 1, f1d



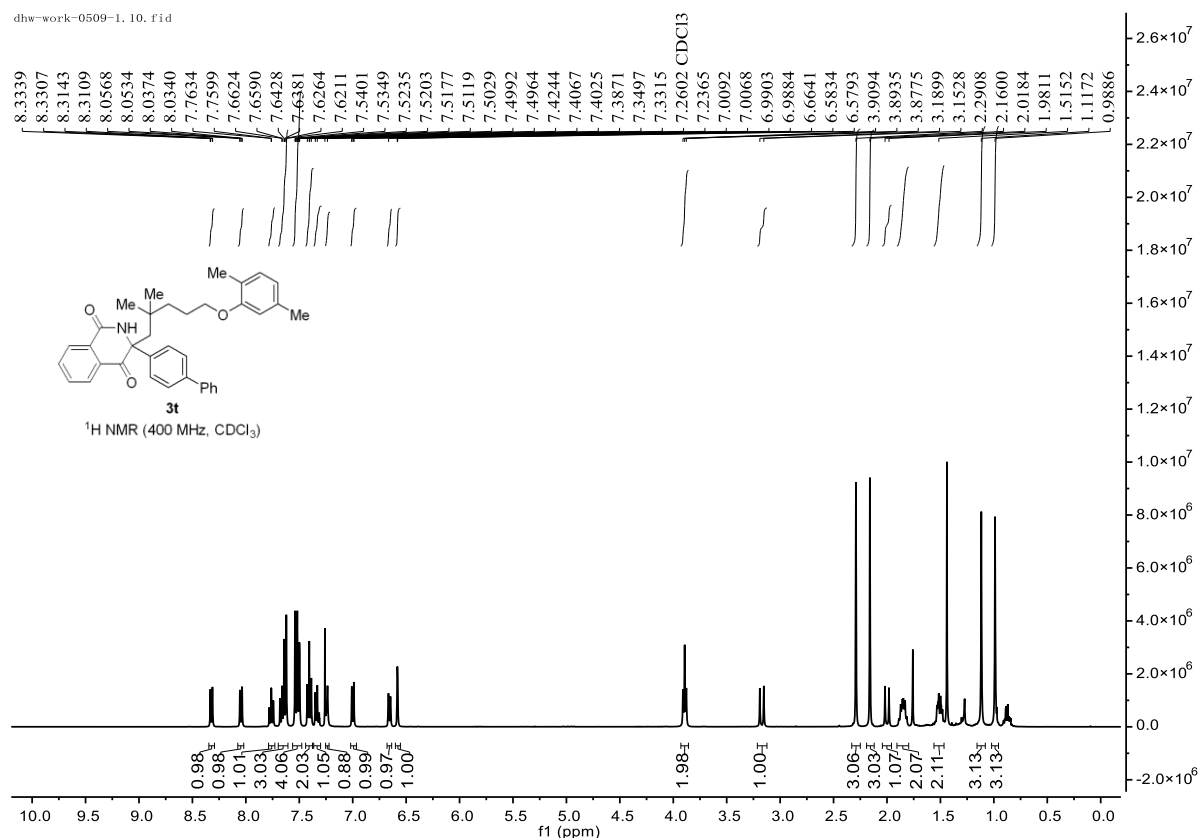
dhw-0723-7-48j, 26. f1d



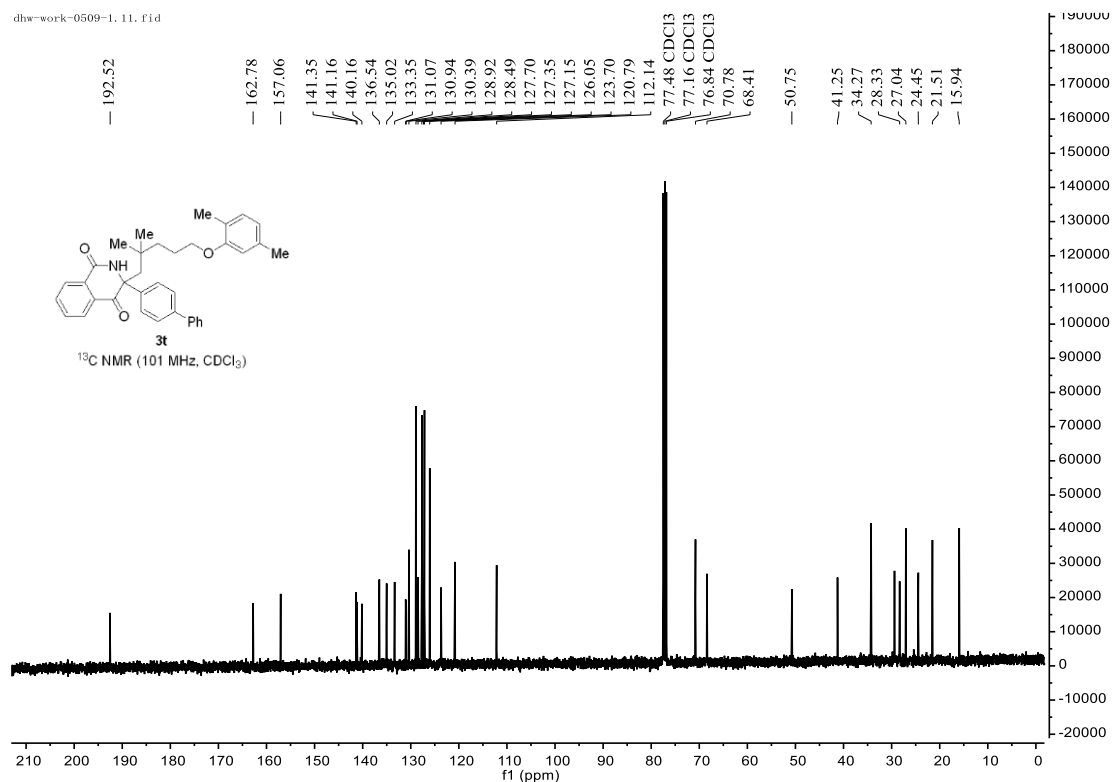
dhw-0723-7-48j, 27. f1d



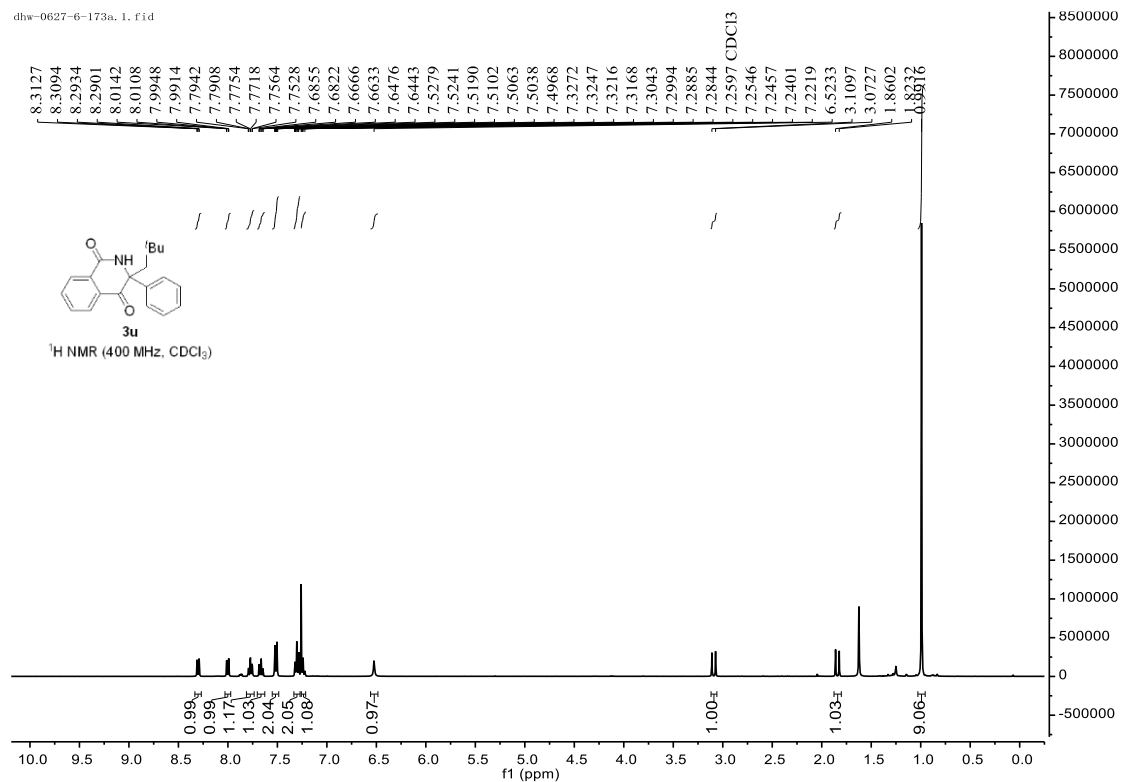
dhw-work-0509-1.10.fid



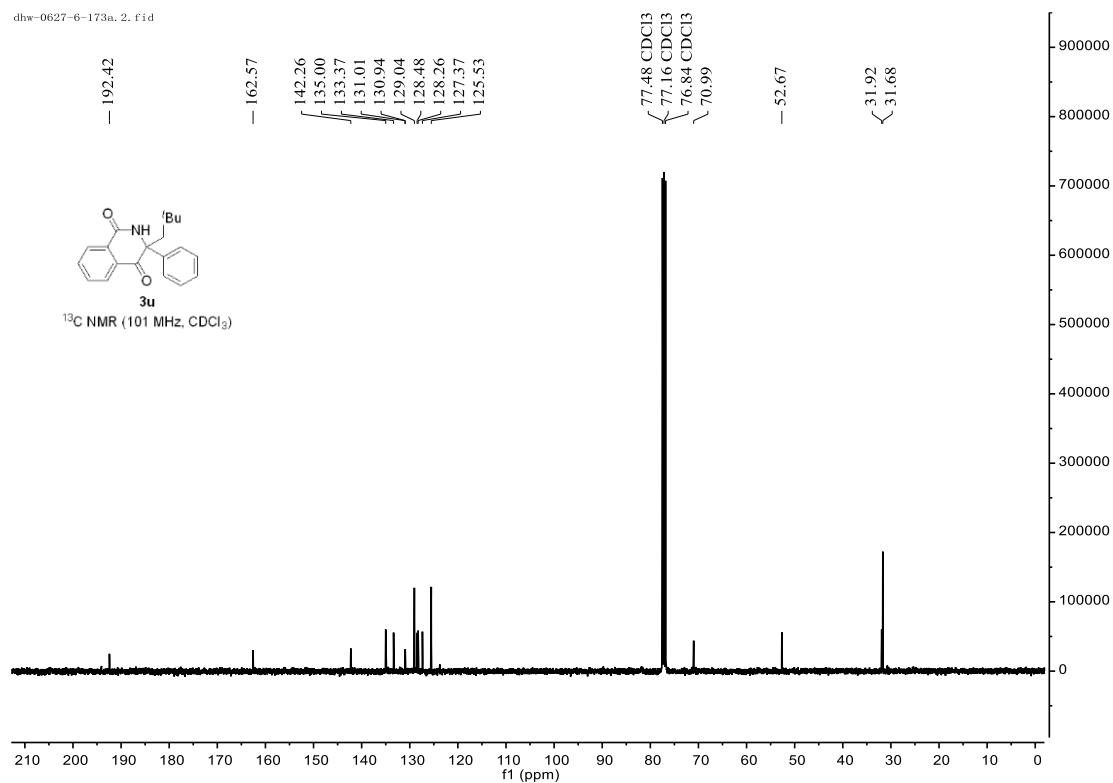
dhw-work-0509-1.11.fid



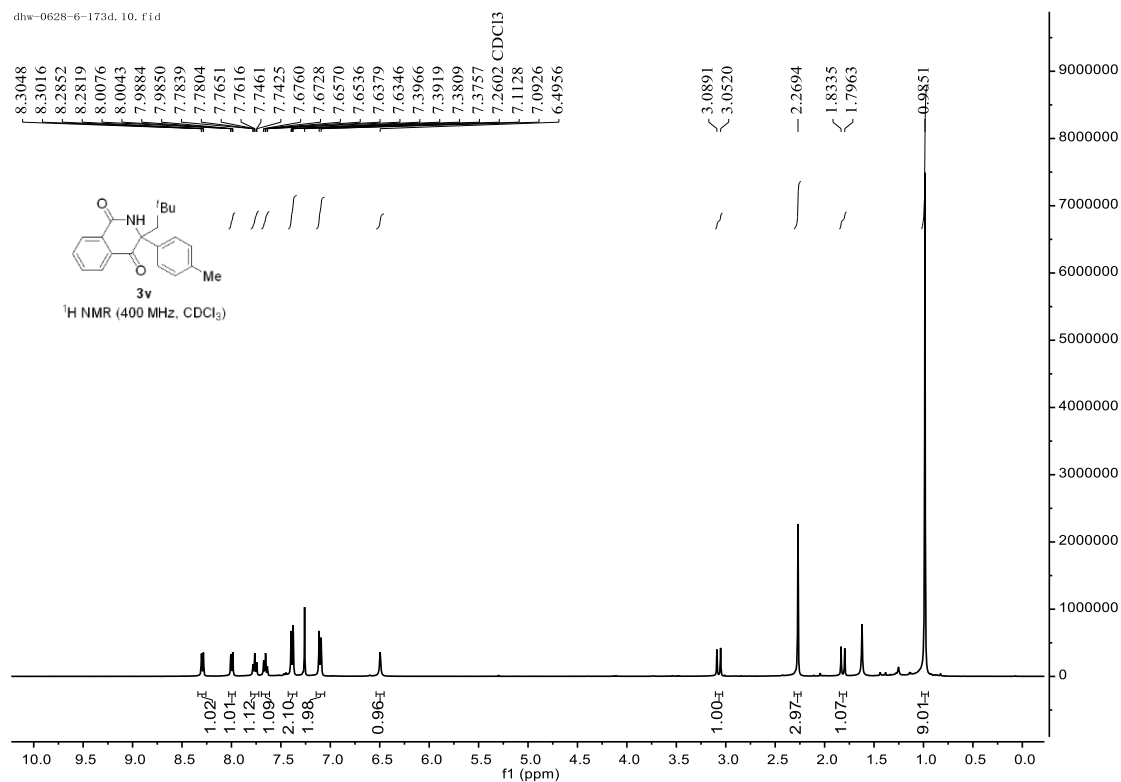
dhw-0627-6-173a, 1, f1d



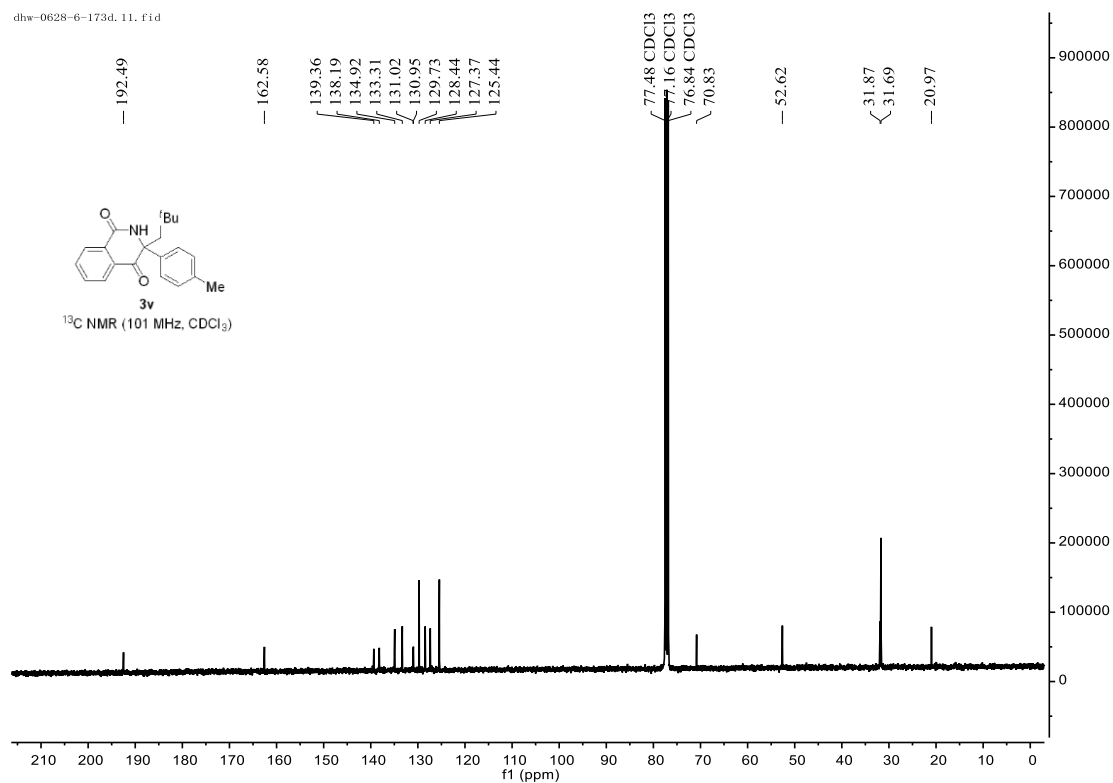
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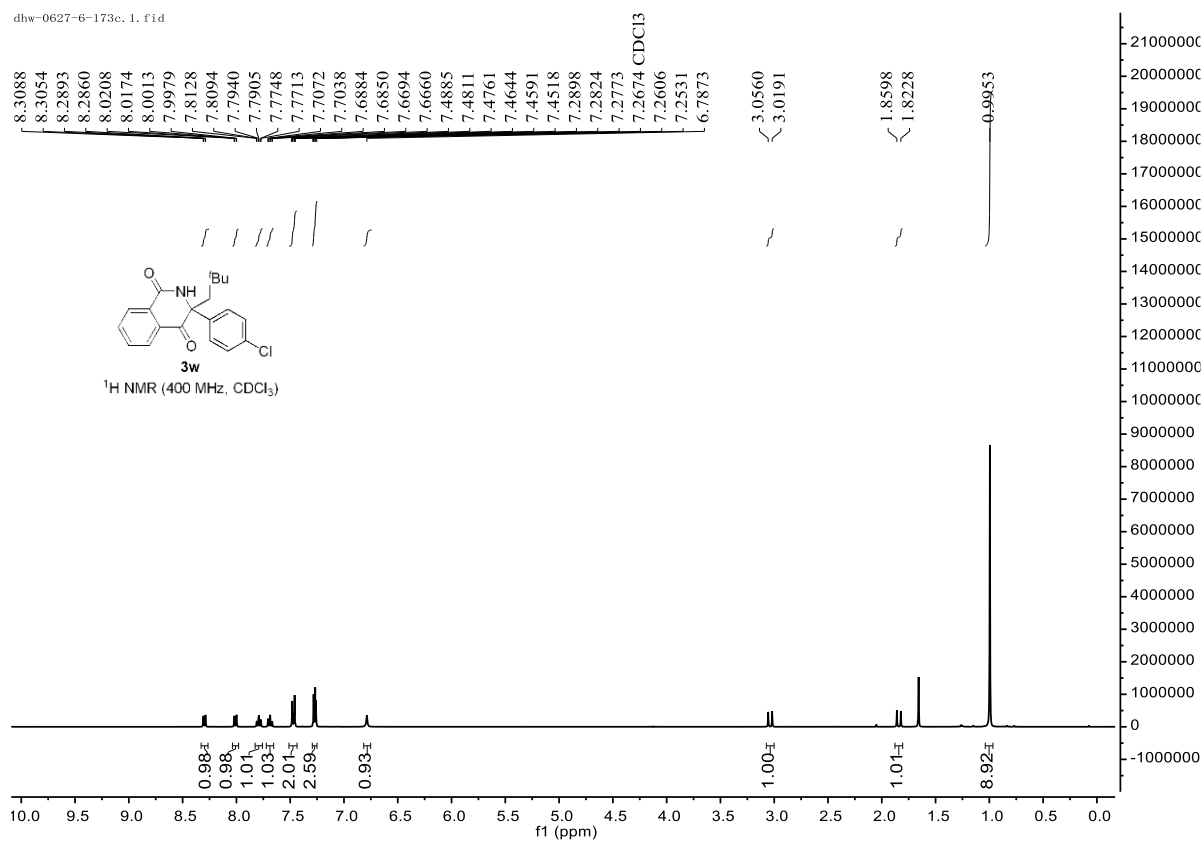
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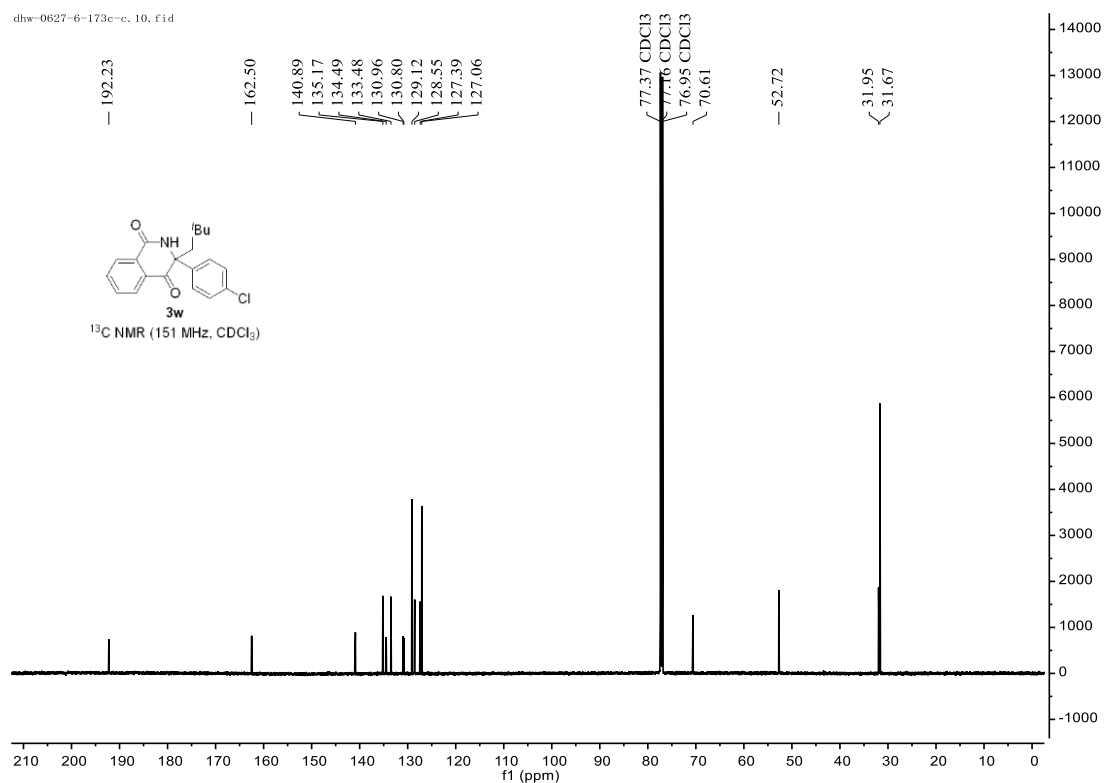
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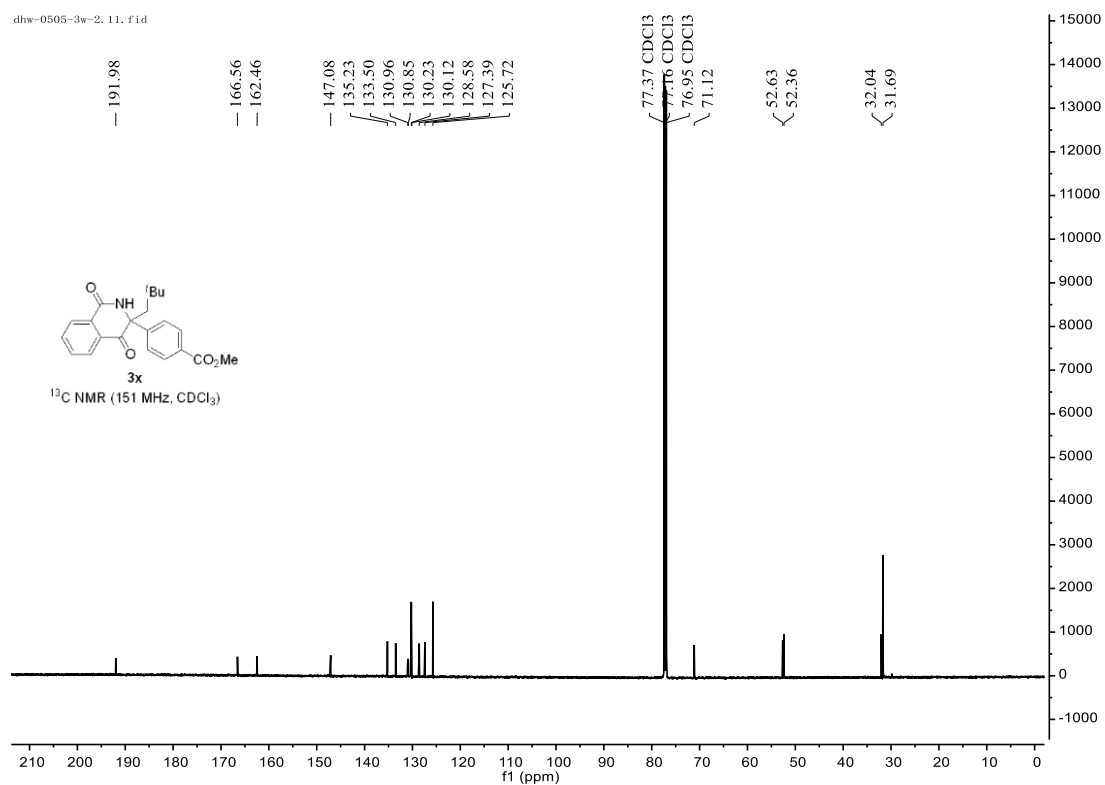
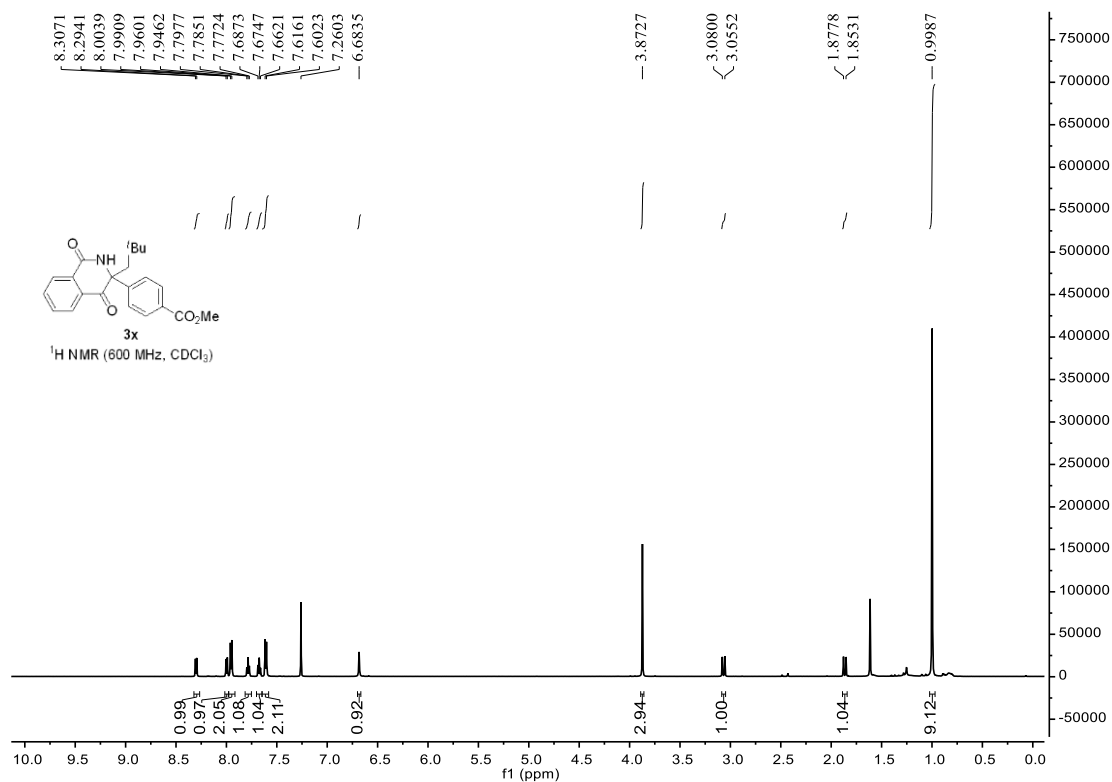


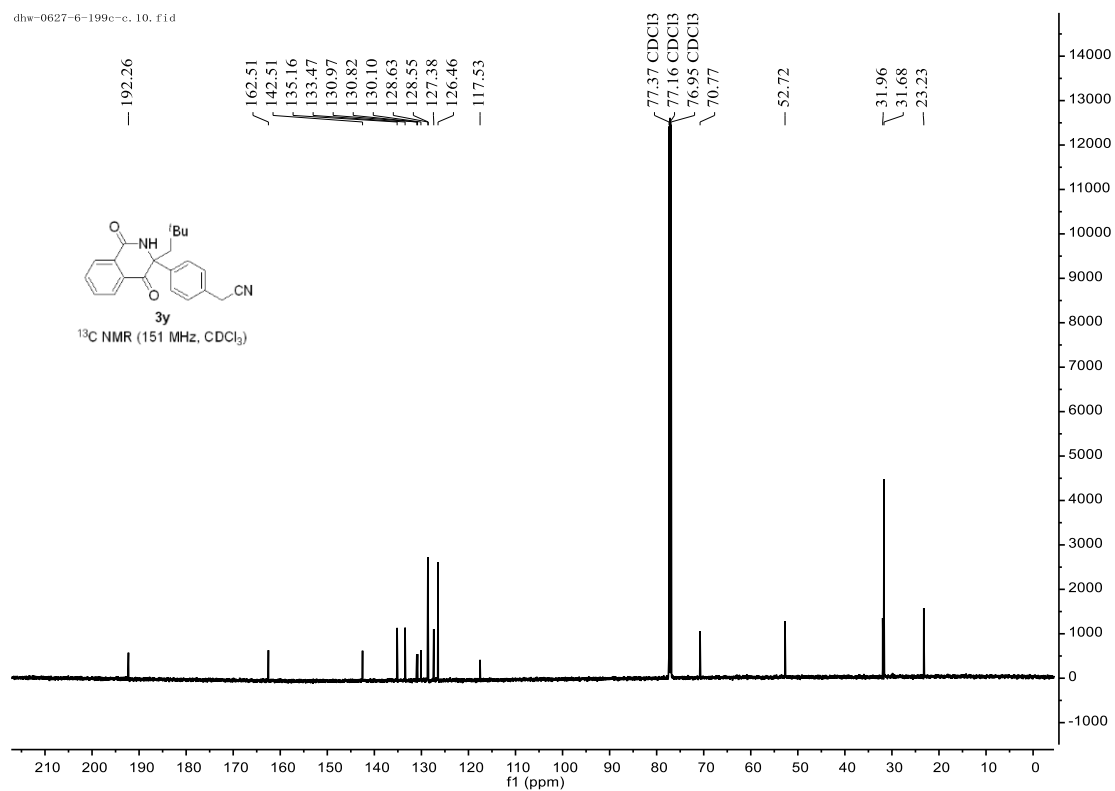
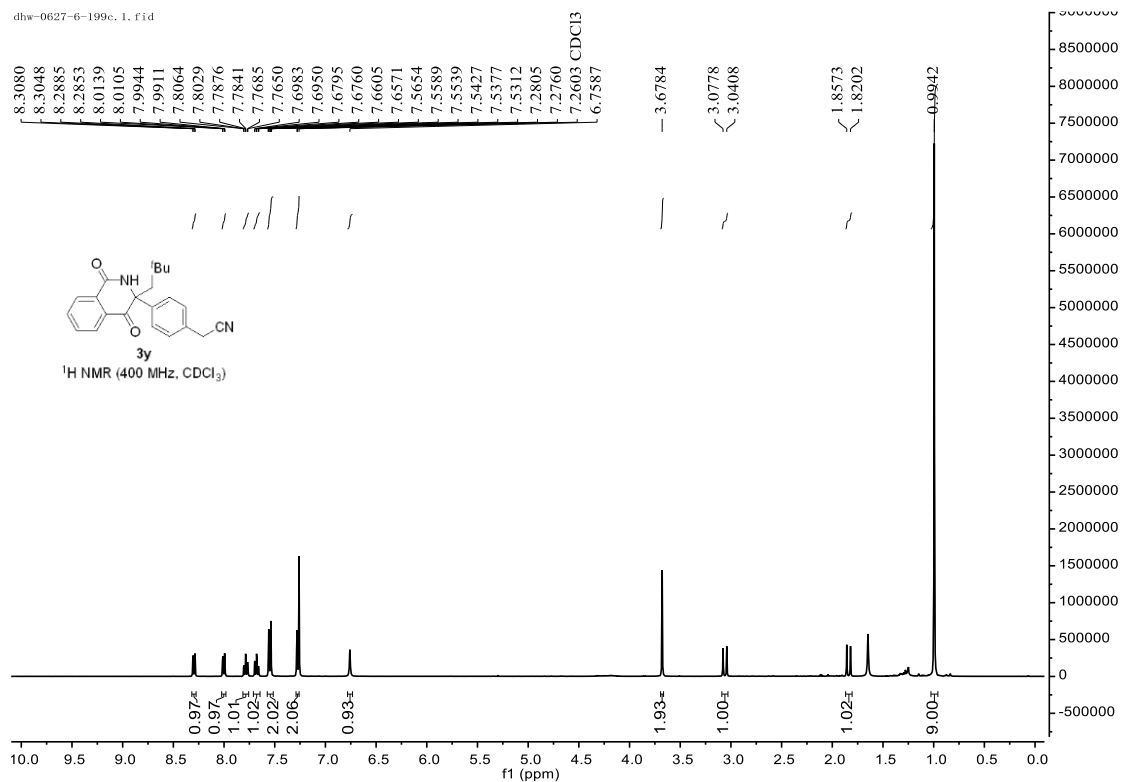
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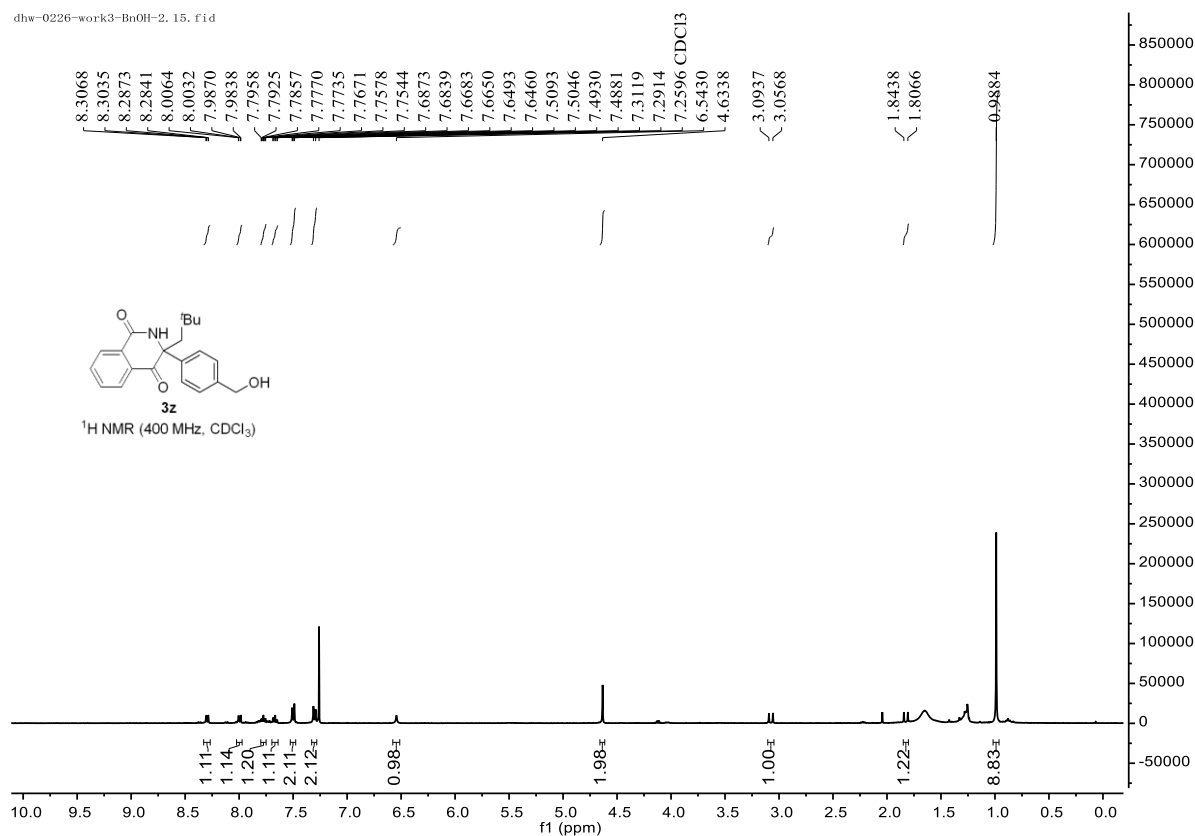
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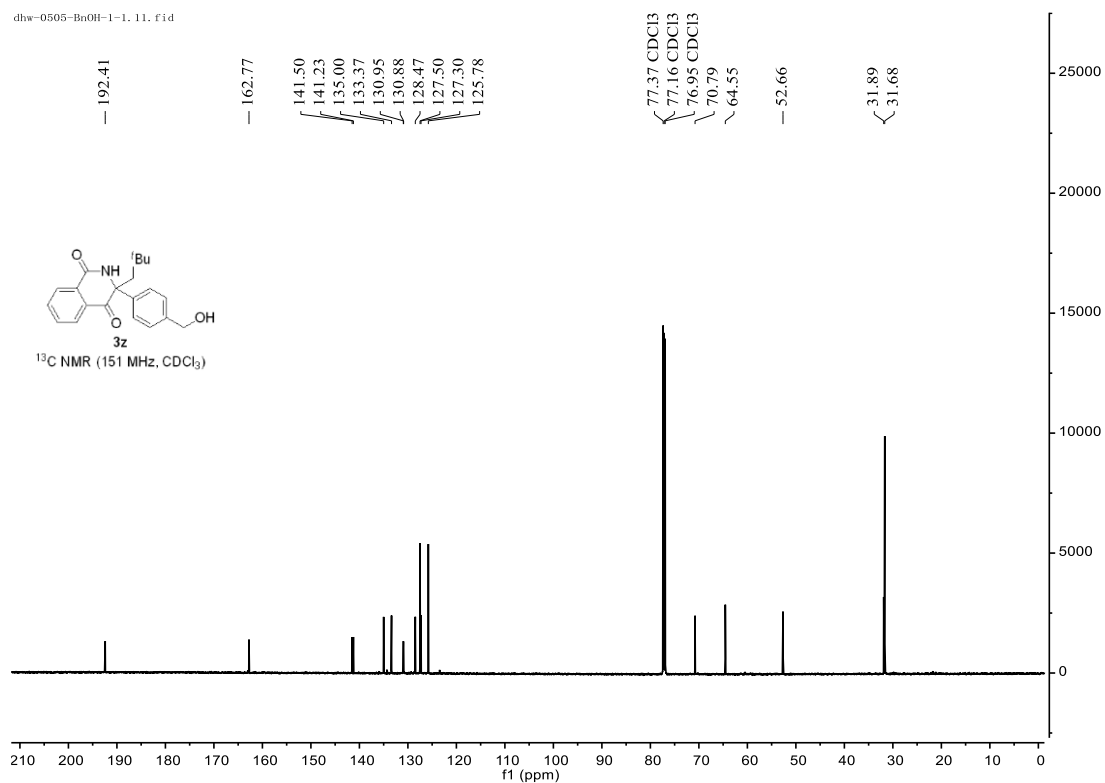




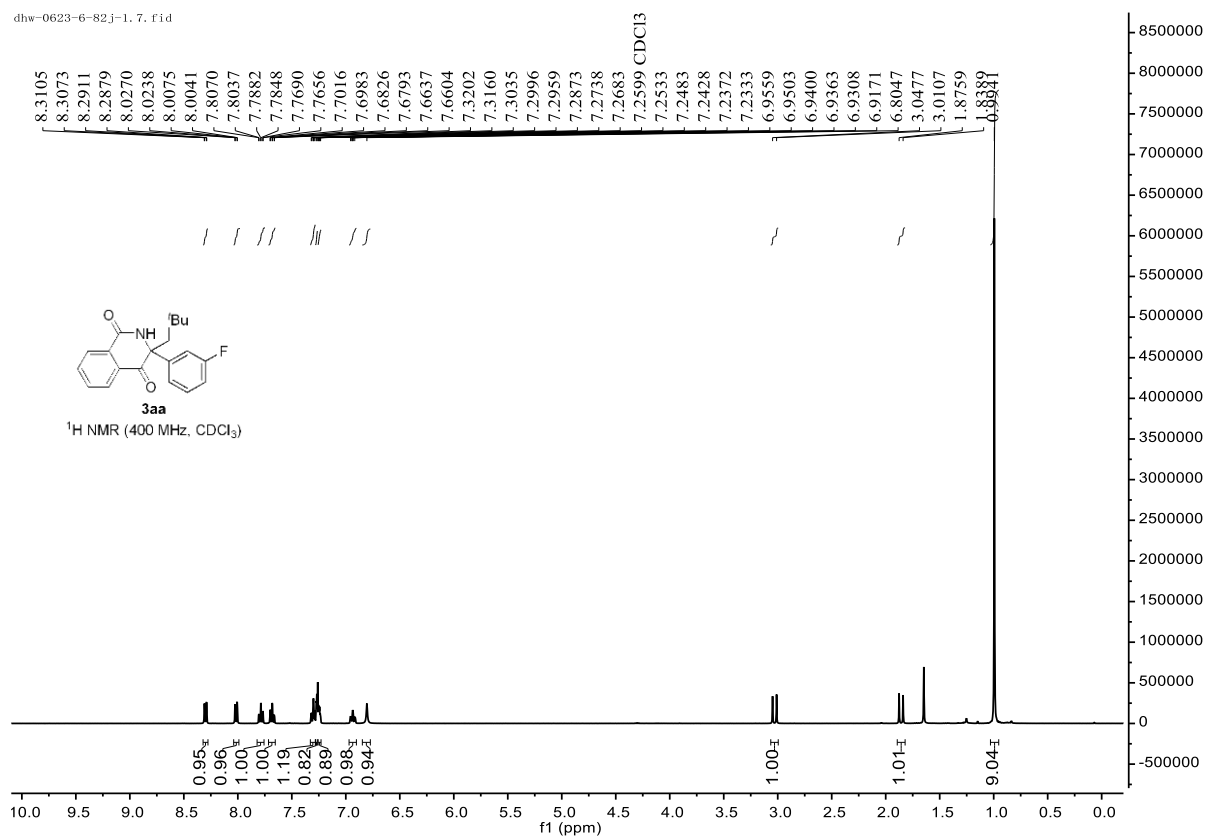
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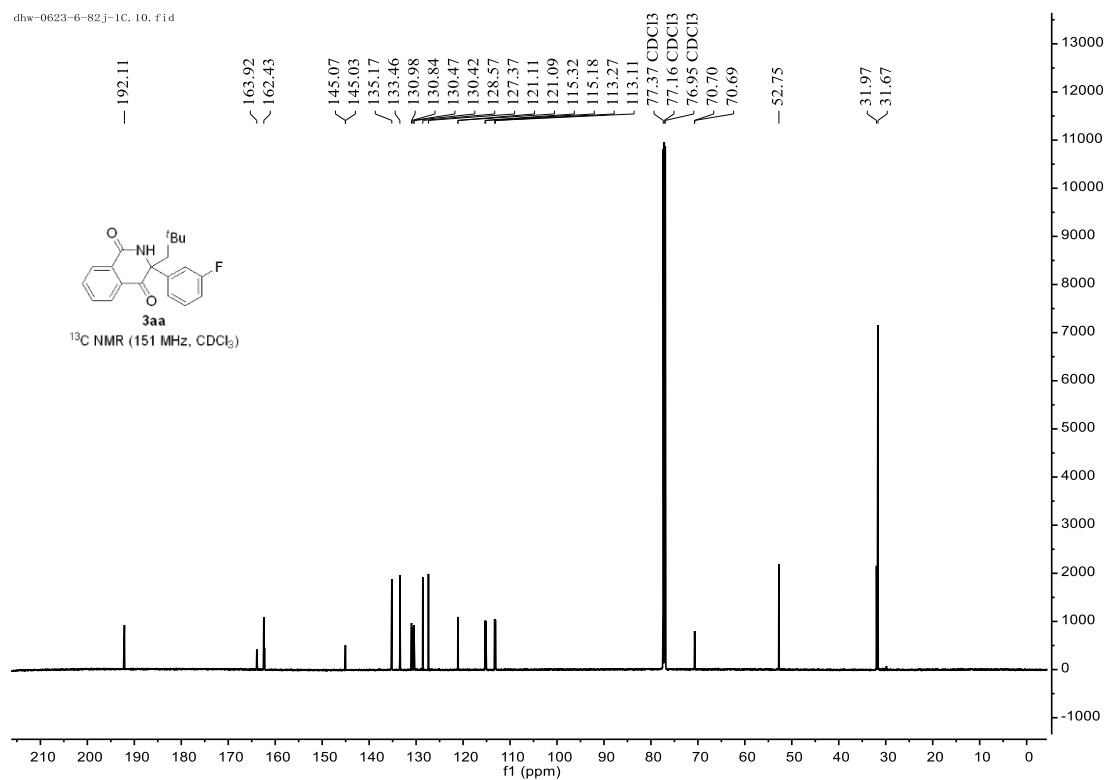
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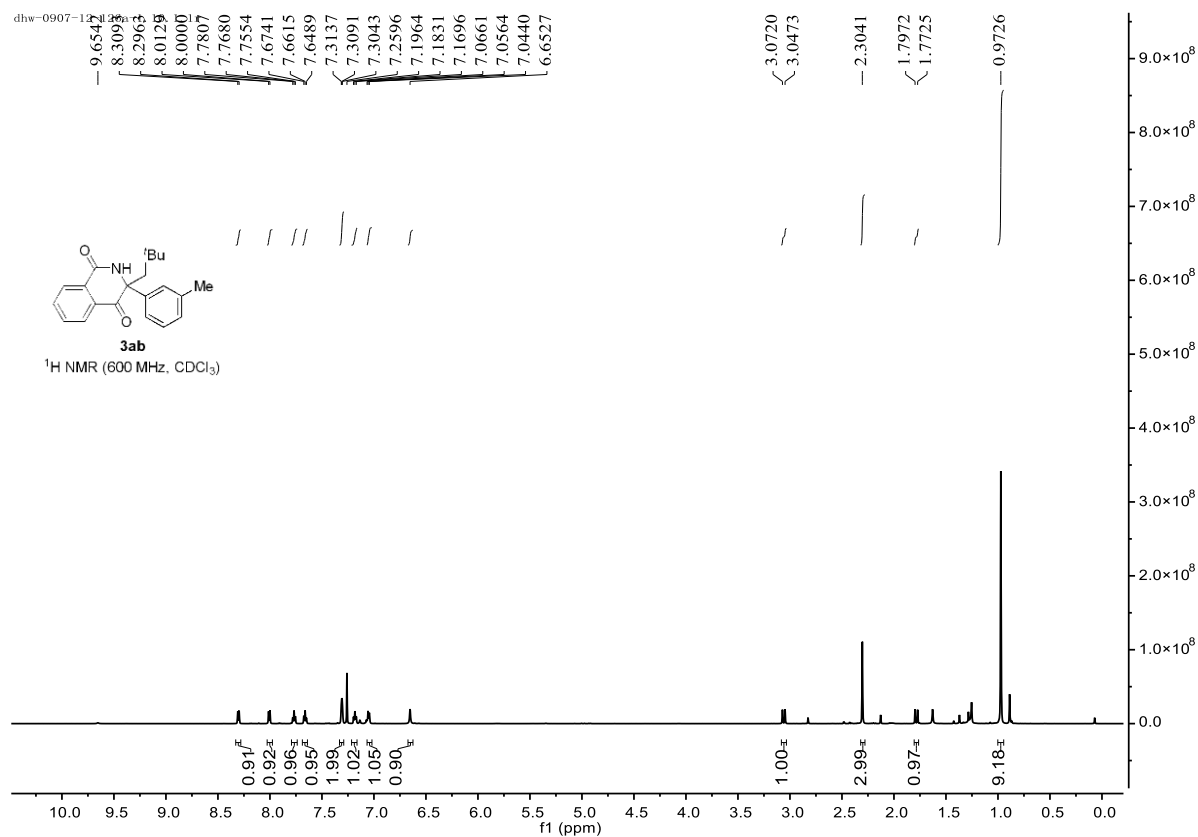
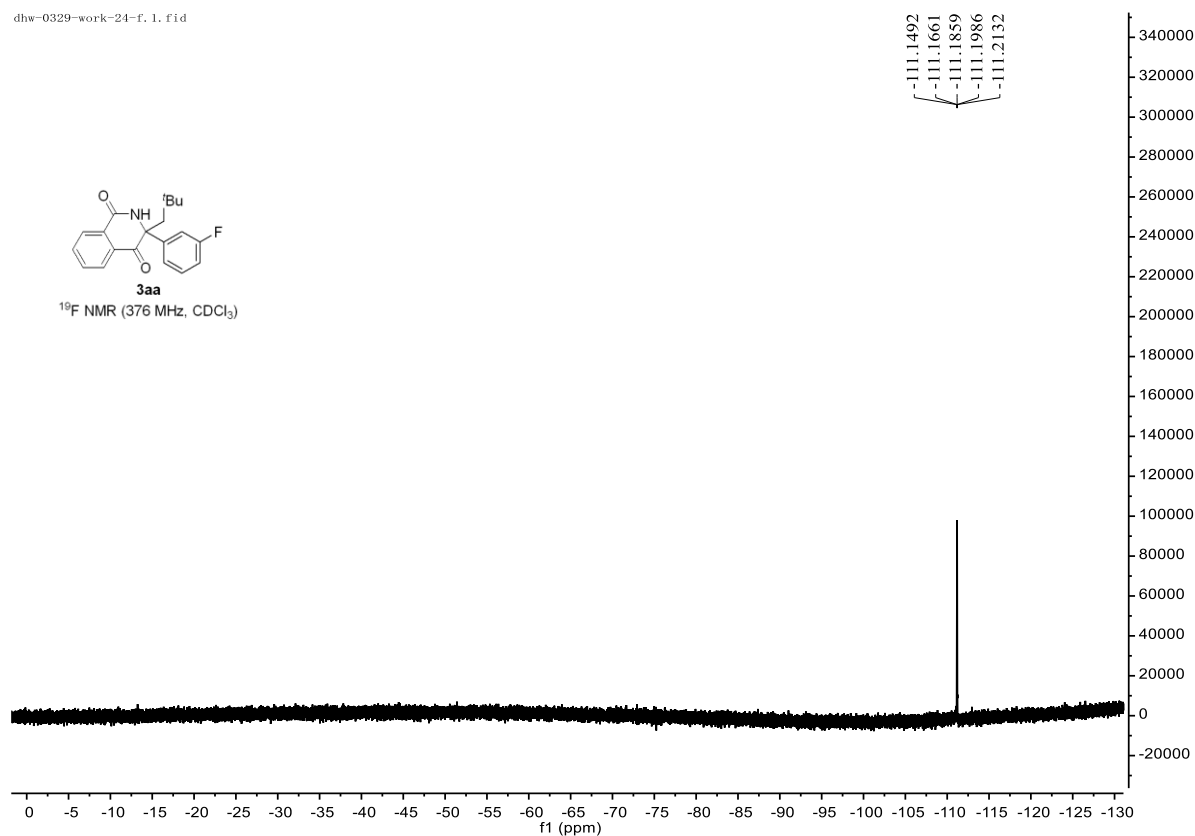
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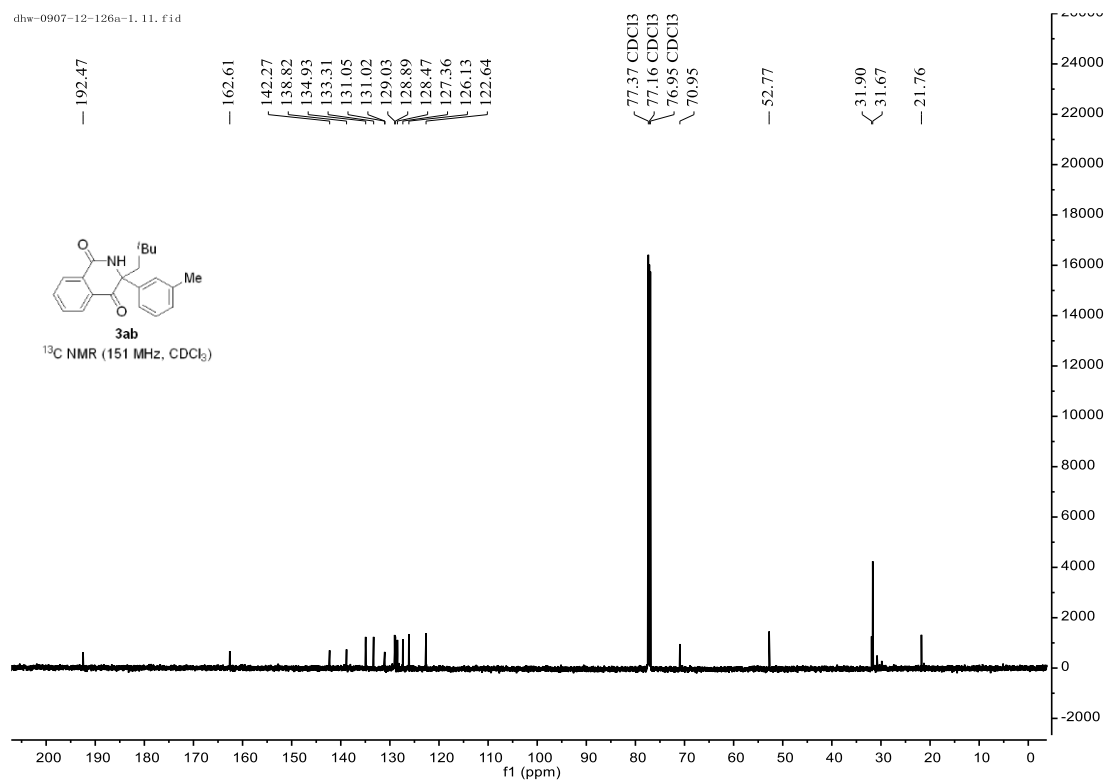
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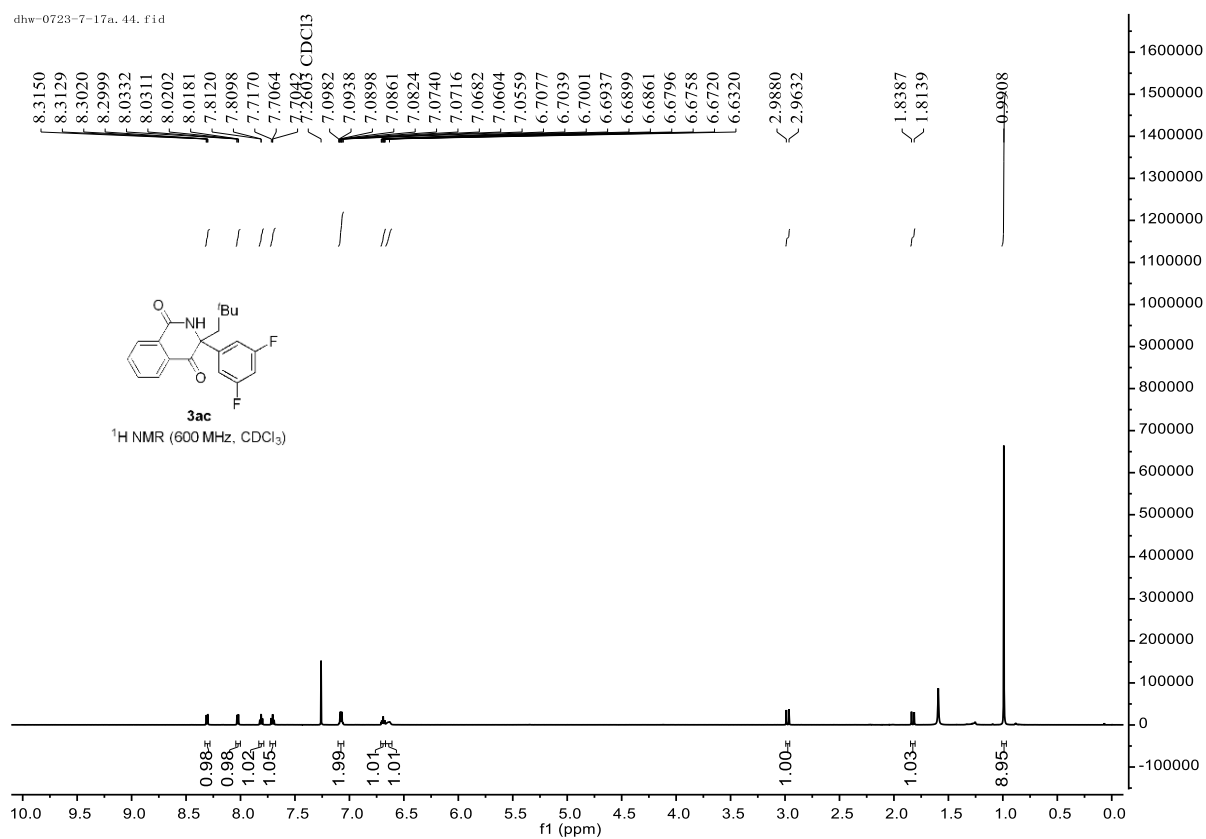
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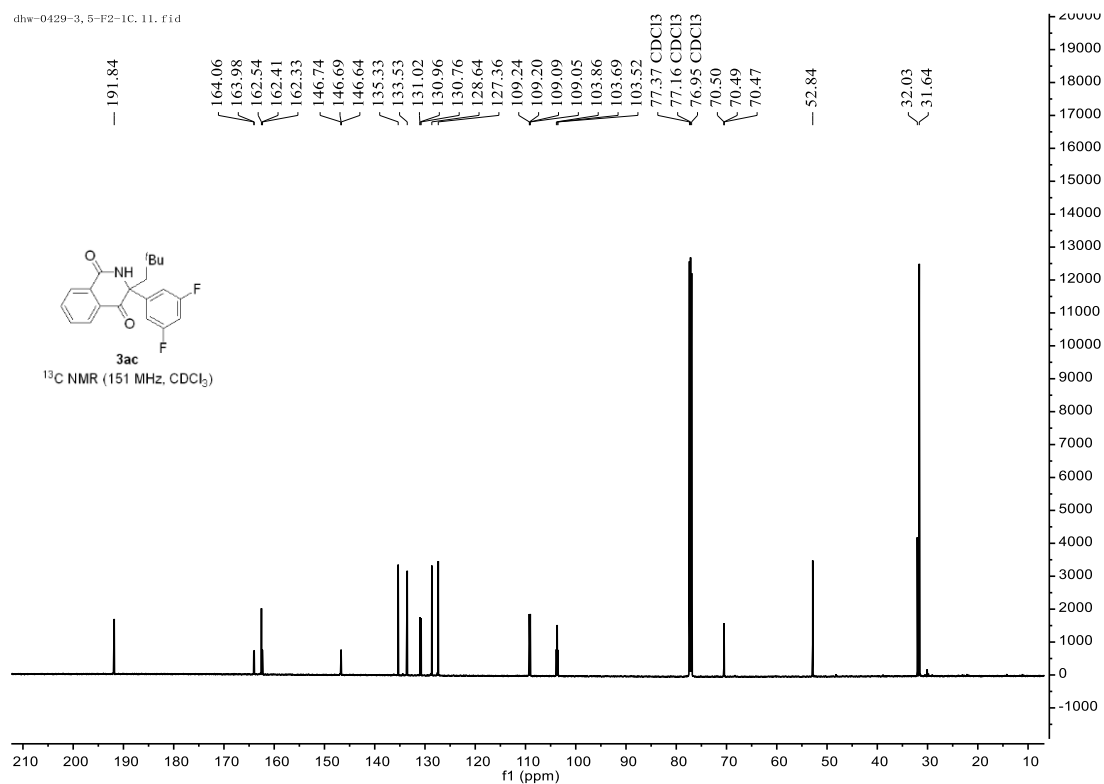
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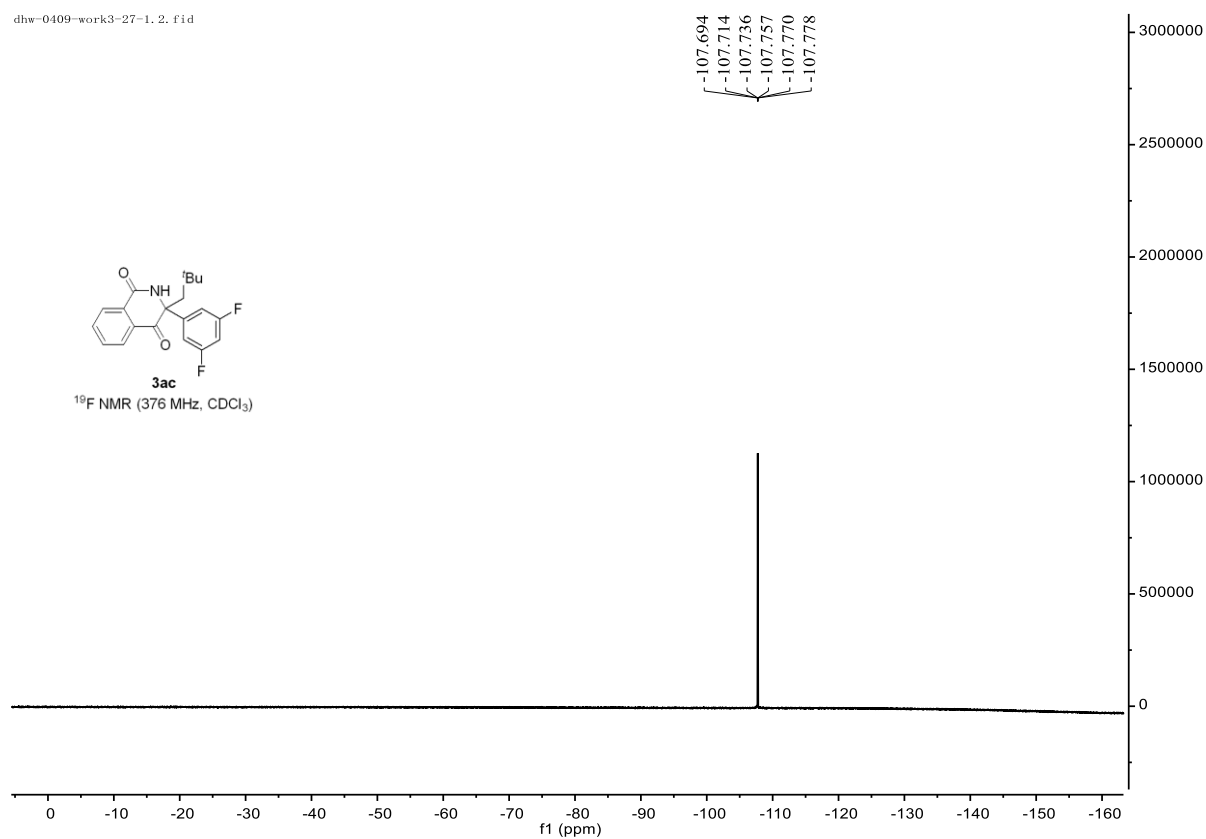
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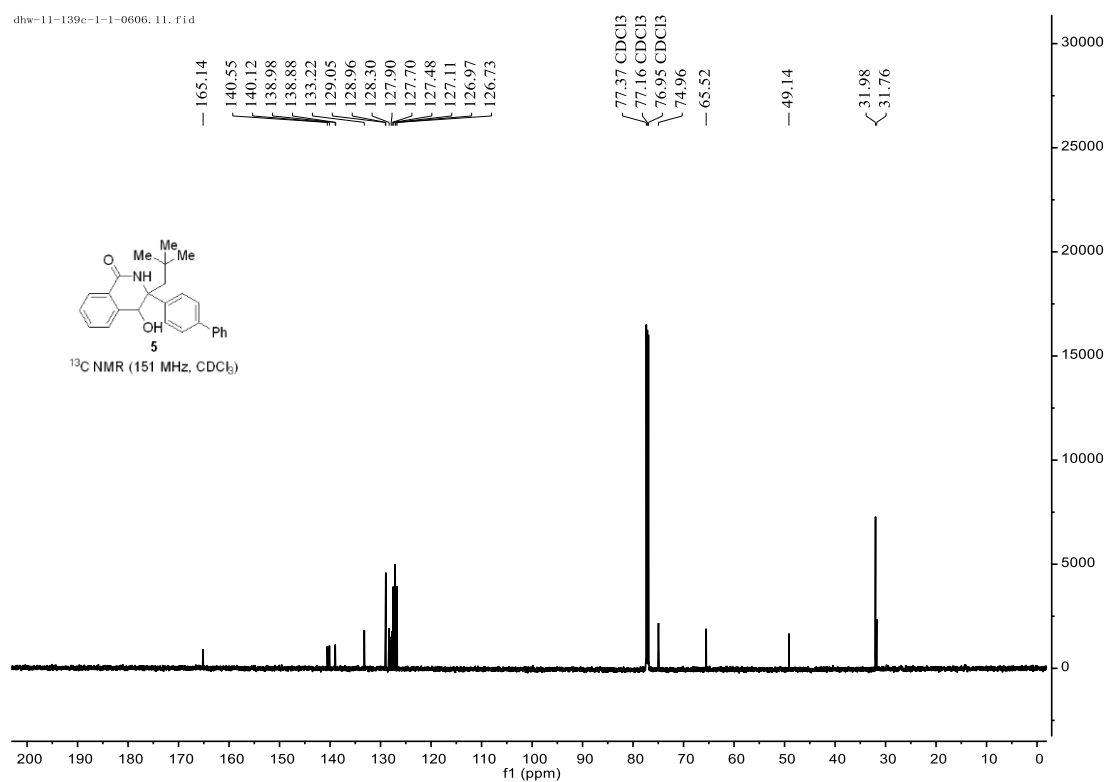
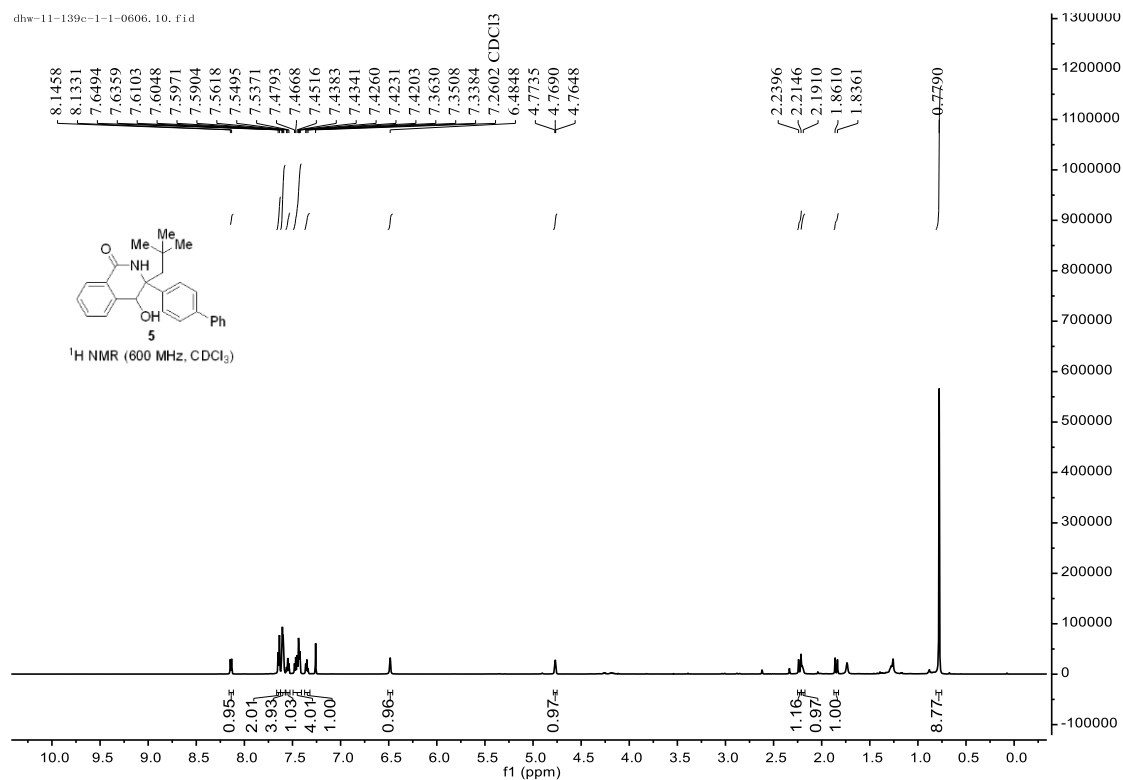


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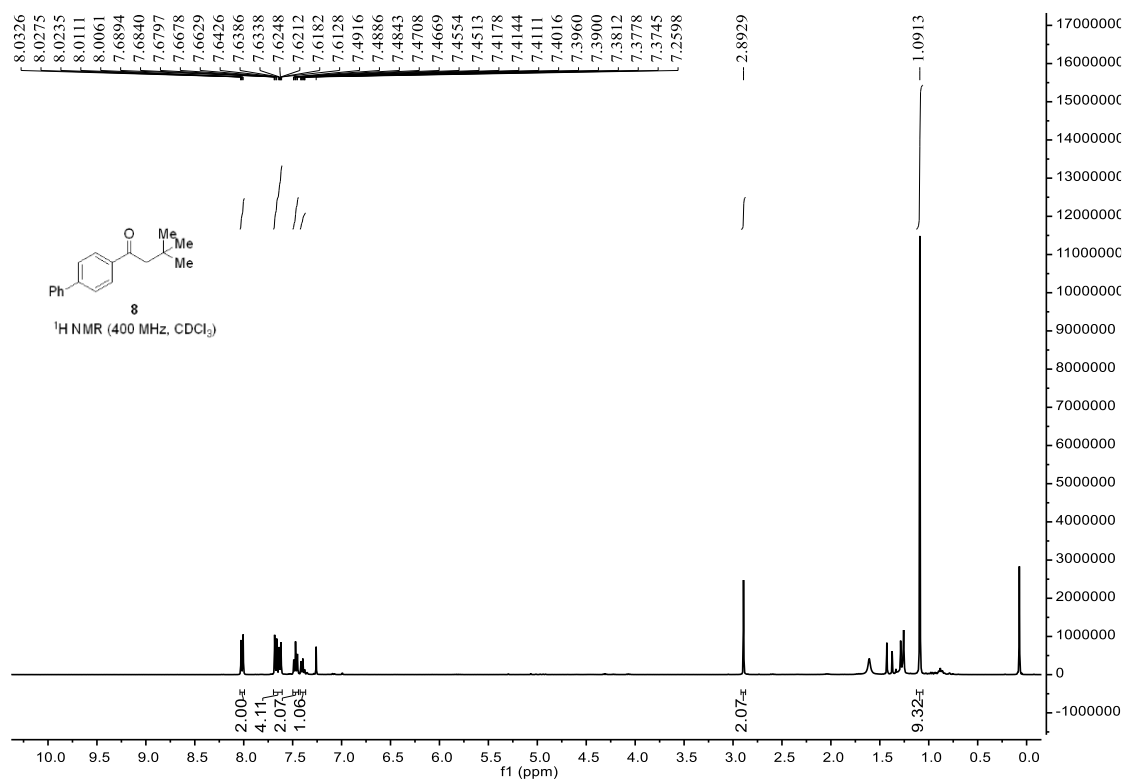
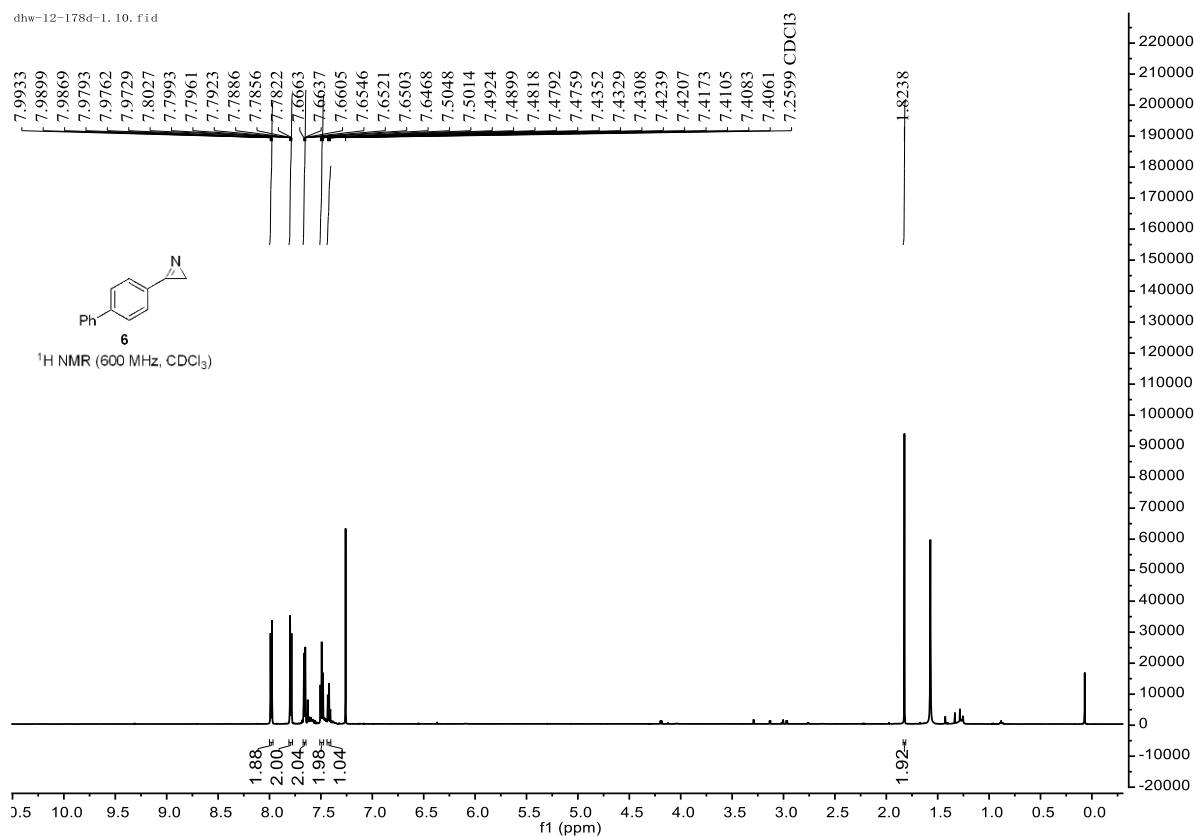


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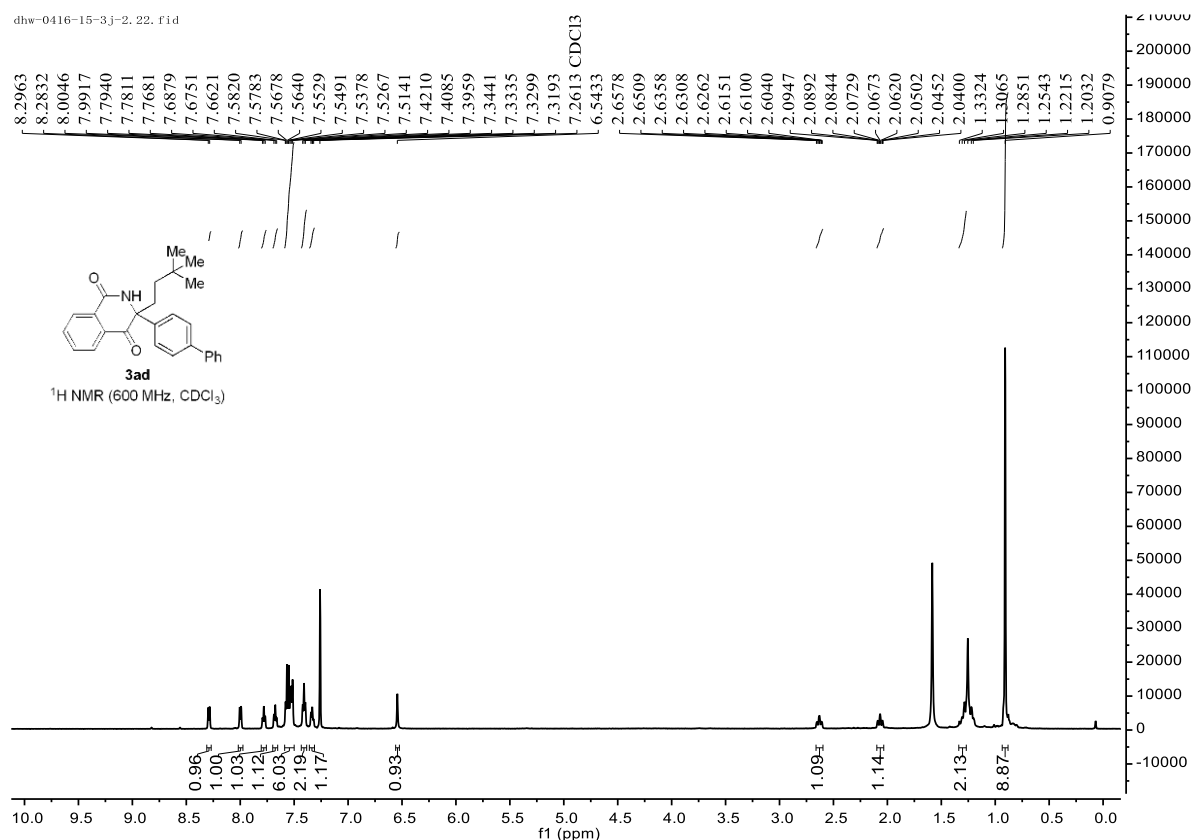




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dhw-0416-15-3j-2.22.fid



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